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# THE STUDY OF HYBRIDIZATION PROCESSES WITHIN GENUS SPARGANIUM L. SUBGENUS XANTHOSPARGANIUM HOLMB. BASED ON DATA OF NEXT GENERATION **SEQUENCING (NGS)**

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\* The study represents the results of research of intragenic polymorphism in transcribed spacer ITS1 of the 35S rRNA genes in representatives of subgenus Xanthosparganium genus Sparganium which were obtained by means of locus-specific next generation sequencing on the platform Illumina MiSeq. It was shown that ribotype variations in studied samples generally correspond to the division of this genus into three sections - Erecta (subgenus Sparganium), Natantia and Minima (subgenus Xanthosparganium). High level of intragenic polymorphism was revealed in S. hyperboreum, with ribotypes distributed among several groups. Genome of this species includes ribotypes which are typical for other species in subgenus Xanthosparganium. For two investigated S. glomeratum samples, there were no ribotypes similar to such ribotypes in other species of Natantia section. S. glomeratum has got ribotypes identical with S. hyperboreum of Minima section. This feature may be the evidence of ancient intersectional hybridization of these two species. Characteristics of rDNA in S. glomeratum are in favor of putting this species into Minima section. It was suggested that speciation processes within the genus could be based not only on hybridization but also went on in allopatric way. The fist statement is supported by the presence of similar and identical ribotypes in S. emersum, S. × longifolium, S. gramineum and S. hyperboreum, the second – as it was mentioned by other researchers, is due to close relationship between North American and Eurasian taxa.

\* Keywords: Typhaceae; bur-reed; gene systematics; molecular phylogeny; interspecific hybridization; speciation.

## ГИБРИДИЗАЦИОННЫЕ ПРОЦЕССЫ В РАМКАХ РОДА SPARGANIUM L. ПОДРОДА XANTHOSPARGANIUM HOLMB. ПО ДАННЫМ СЕКВЕНИРОВАНИЯ СЛЕДУЮЩЕГО ПОКОЛЕНИЯ (NEXT GENERATION SEQUENCING — NGS)

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🟶 В работе изложены результаты сравнительного исследования внутригеномного полиморфизма транскрибируемого спейсера ITS1 гена 35S рРНК у представителей подрода Xanthosparganium рода Sparganium, полученные методом локус-специфичного секвенирования следующего поколения на платформе Illumina MiSeq. Показано, что вариации риботипов изученных образцов, в целом, соответствуют делению рода на три секции — Erecta (подрод Sparganium), Natantia и Minima (подрод Xanthosparganium). Высокий уровень внутригеномного полимофизма выявлен у S. hyperboreum, риботипы которого были распределены по нескольким группам. В геноме этого вида присутствуют риботипы, характерные для других видов подрода Xanthosparganium. У двух изученных образцов S. glomeratum нами не обнаружено риботипов, сходных с таковыми у других видов секции Natantia. S. glomeratum имеет одинаковые риботипы с S. hyperboreum из секции Minima. Такая особенность может являться признаком древней межсекционной гибридизации этих двух видов. Особенности рДНК S. glomeratum говорят в пользу того, что вид может быть отнесен к секции Minima. Высказано мнение, что видообразовательные процессы в рамках рода могли происходить не только на основе гибридизации, но и аллопатрическим путем. Первое подтверждается наличием близких и одинаковых риботипов у *S. emersum*, *S. × longifolium*, *S. gramineum* и *S. hyperboreum*; второе, как ранее указывалось другими исследователями, кроется в близкой связи североамериканских и евроазиатских таксонов.

**ж Ключевые слова:** Турһасеае; ежеголовник; геносистематика; молекулярная филогения; межвидовая гибридизация; видообразование.

#### INTRODUCTION

The phenomena of interspecific and intergeneric hybridization have recently been considered as an effective way of occupying new ecological niches and formation of species of vascular plants [1-4]. Among 37,000 species of flora in Europe, North America, and parts of Australia, belonging to 3,212 genera of 282 families of vascular plants, the species, which morphology suggests as their hybrid origin, were registered in 40% of families and 16% of genera, with an average frequency of nine species of hybridogenic origin per 100 species of non-hybrid origin [5]. In particular, among aquatic and coastal plants, hybrids were recorded in the genera Potamogeton L. [6-9], Nuphar Smith [10], Nymphaea L. [11, 12], Carex L. [13-15], Ranunculus L. [16, 17], and Typha L. [18, 19]. From this point of view, the genus Sparganium L. (bur-reed), which belongs to the sister family of Typhaceae for Pophae, is of great interest as the object of study [20-24].

The *Sparganium* genome is one of the smallest genomes among vascular plants: 1C = 490 million bps. [25]. It should be noted that, unlike gramineous plants, in which the number of chromosomes in the genome has changed repeatedly [26, 27] as a result of recent acts of polyploidization and secondary diploidization, the number of chromosomes in different species of bur-reed is stable, 2n = 30 [28, 29]. It follows that the average size of the chromosome in bur-reeds is 32 million bps, which is four times lower than in humans, for example.

The Sparganium karyotype comprises two pairs of nucleolus organizer chromosomes [30]. A relatively high number of chromosomes in the karyotype (x = 15) suggests that all the representatives of the genus are poliploids, probably allotatraploids [31]. The available data suggest the distribution of interspecific hybridization in the bur-reed, which contributes to the creation of new genotypes and phenotypes. Because of this, the plants occupy certain ecological niches in different types of water reservoirs in a wide range of natural climatic zones. Thus, Y. Ito et al. [23] revealed the existence of a number of combinations of hybrids, namely, S. angustifolium × S. emersum (S. engleranum Asch. et Graebn.), S. hyperboreum  $\times$  S. natans, S. acaule  $\times$  S. fluctuans, and S. fallax  $\times$  S. japonicum, by comparing the topology of molecular phylogenetic trees constructed on the basis of variations in the sequences of chloroplast DNA and the phytochrome C nuclear gene (phyC). Apparently, that the bur-reed hybrids are homoploid hybrids that preserve the ploidy level of the parent species.

As a rule, this type of hybridization, which is accompanied by the incorporation of a certain part of the genes of one parent species into the genome of another, is called introgressive hybridization [32].

We conducted a comparative analysis of intragenomic polymorphism in the subgenus Xanthosparganium Holmb. of the Sparganium genus. We also searched for traces of hybridization processes in related species of the genus using locus-specific sequencing of the transcribed spacer (ITS1) of 35S rRNA genes repeatedly that are multiplicated in the genome and encode the sequences 18S, 5.8S, and 26S rRNA of the large and small ribosome subunits. In plants, these sequences are transcribed as a single transcription unit; the molecular weight of the transcript is approximately 35S (45S in animals) [33]. In each nucleolus organizer, there may be several thousands of 35S rRNA genes [34, 35]. The study of homoploid hybrids and allopolyploids revealed that the 35SrRNA genes obtained from their ancestors can be preserved in their genome for many generations. Their sequencing enables us to identify recent events of interspecific hybridization in the species [36, 37], which was the aim of our work.

#### **MATERIALS AND METHODS**

Because of the likely hybrid origin of the species, we investigated the intragenomic polymorphism of the ITS1 spacer of the *35S* rRNA gene using locus-specific sequencing on the Illumina MiSeq sequencer. The ITS1 region was selected as a marker due to the fact that, in some cases, rDNA isogenization does not occur completely, resulting in hybrids retaining parental sequences in their genome (for example [38]). A successful study of hybrid origin, using pyrosequencing of the ITS1 region, has previously been conducted on orchids [39].

Genomic DNA was isolated, as described in [40], and the DNA samples were transferred to the Common Use Center of the All-Russia Research Institute for Agricultural Microbiology for library preparation and sequencing.

Next-generation sequencing (NGS) primary data were processed using standard tools, such as FastQC [41], Trimmomatic [42], and Fastq-join [43]. For further data filtering, we used consensus haplotype filtering, calculation of the corresponding haplotype frequencies, sorting by decreasing haplotype frequencies (equal haplotypes are presented in the subsequent analysis as a single copy, but with the indication of a repetition frequency), the exclusion of single and rare haplotypes from the analysis, reference-free alignment, and filtering of contaminants using the GenBank database (BLAST): https://www.ncbi.nlm. nih.gov/genbank/.

The NGS data on intragenomic pools of ITS1 sequences were obtained for the following herbarium specimens of *Sparganium*:

- S. emersum Rehm. (sample no. 55: Russia, Tver Oblast, Andreapolsky District, Parshinskoe Lake, in the outskirts of the village of Parshino and the village of Bologovo, 14.VIII.2016, collected by E. A. Belyakov and E.V. Garin, in the collection of E.A. Belyakov; sample no. 56: the Republic of Belarus, Minsk Oblast, Myadelsky District, the Stracha River, "Golubye Ozyora" Nature Reserve, 23.VI.2016, collected by E.A. Belyakov, E.V. Garin, and A.G. Lapirov, in the collection of E.A. Belyakov; sample no. 59: Russia, Nizhny Novgorod Oblast, Arzamassky District, near the village of Staraya Pustyn, in the channel between the Dolgoye and Parovoe lakes, 14.VIII.2014, collected by E.A. Belyakov, in the collection of E.A. Belyakov);
- *S. gramineum* Georgi (sample no. 61: Tyumen Oblast, Surgut District, 10 km from the village of Ugut, Ekipomykhlor (Kinyaminskoe) Lake, shallow water offshore, 28.VII.2000, collected by A.S. Baikalova, LE);
- S. glomeratum (Laest. ex Beurl.) Neuman (sample no. 52: Russia, Yaroslavl Oblast, Poshekhonsky District, in the outskirts of the village of Golodyayka, in a forest ditch, 10.IX.2016, collected by E.A. Belyakov, in the collection of E.A. Belyakov; sample no. 58: Russia, Vologda Oblast, Cherepovets District, Darwin Nature Reserve, in the outskirts of the village of Muravyovo, a stream, tributary of the River Chimsory, 01.VIII.1997, collected by A.A. Bobrov, IBIW);
- S. hyperboreum Laest. (sample no. 51: Russia, Tyumen Oblast, Tazovsky District, in the outskirts of the village of Tazovsky, in a waterlogged roadside degradation, 01.VIII.2015, collected by S.A. Nikolaenko, in the collection of E.A. Belyakov; sample no. 53: the same, plant sprouted from seeds; sample no. 57: Russia, the Republic of Komi, Izhemsky District, 76 km from the village of Izhma, crossover of B. Izhma, 2.5 km below, 02.VIII.1979, collected by Z.G. Ulle, in the collection of E.A. Belyakov);
- *S. microcarpum* (Neum.) Domin (sample no. 60: the Republic of Belarus, Minsk Oblast, Myadelsky District, the Stracha River, "Golubye Ozyora" Nature Reserve, 23.VII.2016, collected by E.A. Belyakov, E.V. Garin, and A.G. Lapirov, in the collection of E.A. Belyakov); and
- One hybrid *S.* × *longifolium* Turcz. ex Ledeb. (sample no. 62: Russia, Yaroslavl Oblast, Rostov District, shallow water of Lake Chashnitskoye, in the outskirts of the village of Chashnitsy, 16.VIII.2016, collected by E.A. Belyakov, in the collection of E.A. Belyakov).

In total, we obtained data for five species and one hybrid. In addition, some ITS1 sequences were taken from the GenBank database for comparative analysis [41], namely, *S. emersum* (KF265393), *S. glomeratum* (KF265386), *S. gramineum* (KF265381), *S. fluctuans* (KF265378), *S. fallax* (KF265376), *S. subglobosum* (KF265397, KF265387), *S. hyperboreum* (KF265396), *S. natans* (KF265385), and *S. stoloniferum* (KF265395) [22].

## RESULTS

Our results revealed that the ribotypes of the studied samples were distributed into 10 groups (see Fig. 1). Group 1 (SAE) included KF265393, which was declared to GenBank as S. emersum. However, the ribotype of this sample differs from that of our sample of S. emersum, obtained from the Republic of Belarus, by eight nucleotide substitutions. This difference is in accordance with the opinion of Y. Ito et al. [23], that KF265393 was sequenced from a sample that should have been identified as S. acaule [23]. Group 2 (SGH) mainly comprised S. glomeratum and, to a lesser extent, S. hyperboreum ribotypes. The two samples of S. glomeratum had common ribotypes only with the samples of S. hyperboreum. Group 3 (SLG) included the ribotypes of  $S. \times longifolium$  and its parent species, S. gramineum (KF265381). Group 4 (SFS) was represented by S. fluctuans (KF265378 in the GenBank database). Group 5 (SNA), the largest group, included S. emersum, S. glomeratum, S. gramineum, and  $S. \times longifolium$  ribotypes belonging to the Natantia section. Two groups of ribotypes were represented in the Genbank sequenses only. Group 6 (SFX) was represented by S. fallax ribotype (KF265376), and group 7 (SSM) included S. subglobosum ribotype (KF265397 and KF265387). Group 8 (SNH) consisted of both S. natans (KF265385) and S. hyperboreum (KF265396) ribotypes, together with the ribotypes obtained by us from S. hyperboreum samples taken from Tyumen Oblast and the Republic of Komi. Group 10 (SHM) included the ribotypes characteristic of S. hyperboreum only. It should be noted that groups 8 and 10 were distinguished by characteristic informative nucleotide substitutions for 20 mismatched positions. Group 9 (SMH) included ribotypes of representatives of the Erecta section (S. stoloniferum (KF265395) and S. microcarpum from the Republic of Belarus). The presence of rare S. hyperboreum ribotypes was also noted here.

According to a comparative analysis of the intragenomic polymorphism of the ITS1 region, *S. hyperboreum* retained traces of hybridization processes with *S. glomeratum* in its genome. The presence of identical ribotypes in *S. glomeratum* and *S. hyperboreum* may indicate that these two species are close and are included only in rare cases of introgressive hybridization. In contrast, *S. emersum*, *S.* × *longifolium*, *S. gramineum*, and *S. hyperboreum* have many similar, and some identical, ribotypes.

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*S. microcarpum* is the most remote of all the studied samples. The same group includes a sample of *S. stolon-iferum* from GenBank. Both species are part of the subgenus *Sparganium*. It is noted that this group also included single *S. hyperboreum* ribotypes (sample no. E51 from Tyumen Oblast).

## DISCUSSION

Under unstable climatic conditions of the Oligocene and subsequent geological epochs, the degree of the bur-reeds' resistance to cooling and significant temperature fluctuations increased, which contributed to the expansion of the range of individual species. In this case, the overlapping taxon ranges and various critical situations, such as periodic climatic changes and anthropogenic transformation of biotopes, can cause the spontaneous hybridization of species [44–56]. These are hybrid plants that occupy an intermediate position by their characteristics (e. g., by the nature and structure of endocarps [47–48]). They are possibly heterotic in the first generations, highly polymorphic in genome and phenotype, therefore they had a chance to adapt to new environmental conditions.

As our study demonstrated, the presence of close and identical ribotypes in the group of S. emersum, S.  $\times$  longifolium, S. gramineum, and S. hyperboreum indicates the presence of hybridization within the Natantia section of the Xanthosparganium subgenus. The conclusion regarding the hybrid origin of  $S. \times longifolium$  is also consistent with the special aspects of its morphology, as it combines the characters of S. emersum and S. gramineum [49]. Hybridization between these species and the case of return hybrids are quite possible, since the ranges of species partially overlap. In this case, apparently, selective differentiation leads to the replacement of natural (parental) species through genetic assimilation [50]. In addition, hybridization between two highly specialized species can contribute to secondary despecialization, increasing the ecological flexibility of the hybrid. An example of this is the fact that  $S. \times lon$ gifolium eventually displaces the parent species of burreed from the reservoir, gradually occupying their habitat [49].

Our haplotype distribution pattern is consistent with the division of *Sparganium* into two subgenera, *Sparganium* and *Xanthosparganium*. Representatives of the *Natantia* (*S. emersum*, *S.* × *longifolium*, and *S. gramineum*) and *Minima* (*S. hyperboreum* and *S. natans*) sections are well supported within the *Xanthosparganium* subgenus. However, the presence of common ribotypes in the studied samples of *S. hyperboreum* and *S. glomeratum* suggests the transfer of the latter from the *Natantia* section to the *Minima* section. Previously, *S. glomeratum* belonging to the *Natantia* section was primarily due to the usual similarities (size characteristics, leaves with a pronounced fin, etc.) between this plant and *S. emersum*. Note that the lectotype of the *Natantia* section is *S. angustifolium* Michx. [53]. Meanwhile, *S. glomeratum* has an intermediate position along the length of the column (at the fruitlet) between the two abovementioned sections (e. g., in *S. glomeratum*, the length of the column is  $1.3 \pm 0.2$  mm; in *S. emersum*, it is  $3.4 \pm 0.6$  mm; and in *S. hyperboreum*, it is  $0.3 \pm 0.1$  mm), and upper pestillate inflorescences approximate to each other, which is often observed in representatives of the *Minima* section. Therefore, the transfer of *S. glomeratum* to the *Minima* section can be confirmed by both its molecular and morphological characteristics.

The presence of haplotypes common to all studied species of the subgenus *Xanthosparganium* may indicate the prevalence of hybridization in the subgenus. It should be noted that significant intragenomic polymorphism was found in *S. hyperboreum*, which had common haplotypes with the three other species of the *Natantia* section. The largest numbers of common haplotypes were recorded in *S. hyperboreum* and *S. glomeratum*. *S. glomeratum* itself had common haplotypes only with *S. hyperboreum*. This presentation can be explained by the introgressive hybridization between these two species. In addition, *S. hyperboreum* itself may be a species of hybrid origin; the result of the hybridization of species from different sections, *Natantia* and *Minima*.

Speciation within the genus could occur not only on the basis of hybridization but also allopathically. In this regard, confirmation of the close relationship between the North American and Eurasian pairs of species, for example, S. fluctuans-S. gramineum, plays an important role at the molecular level [22, 23]. There are other pairs of species that are of interest to modern researchers (e. g., S. americanum-S. japonicum [54]), the relatedness of which have not yet been sufficiently studied using molecular data [22]. The previously expressed point of view on the formation of taxa allopathically [22, 23] is also confirmed by the fact that, in the Eocene-Oligocene era, there was a union of the continental fragments, which led to a reduction in the number of ecological provinces; namely, Hindustan separated from Africa and joined Asia; in the Cretaceous-Paleogene era (52-60-65 megayears ago), Laurasia split and North America separated from Greenland, and Greenland from Europe, the remnants of Gondwana split, and the separation of Antarctica and Australia occurred [55]. It should be noted that confirmation of the close relationship between North American and Eurasian taxa indicates the monophyletic nature of the genus Sparganium [22, 56].

### CONCLUSIONS

According to the NGS data, a comparative analysis of the intragenomic polymorphism of the ITS1 spacer revealed that the distribution of close haplotypes in the studied samples corresponds to the division of the genus *Sparganium* into two subgenera (*Sparganium* and *Xan-thosparganium*).

Within the *Xanthosparganium* subgenus, the *Minima* section is well supported. As for the *Natantia* section, the two studied samples of *S. glomeratum* did not exhibit the same haplotypes as the other studied species samples in this section; however, there were identical haplotypes with the two studied *S. hyperboreum* samples from the *Minima* section. Therefore, *S. glomeratum* can be relegated to the *Minima* section.

The presence of a large number of hybrid taxa, including intersectional ones, allows us to consider the genus *Sparganium* as an introgressive interspecies complex promising for molecular phylogenetic studies.

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### REFERENCES

- Baack EJ, Rieseberg LH. A genomic view of introgression and hybrid speciation. *Curr Opin Genet Dev.* 2007;17(6):513-518. https://doi.org/10.1016/j. gde.2007.09.001.
- 2. Soltis PS, Soltis DE. The role of hybridization in plant speciation. *Annu Rev Plant Biol.* 2009;60:561-588. https://doi.org/10.1146/annurev. arplant.043008.092039.
- Носов Н.Н., Пунина Е.О., Мачс Э.М., Родионов А.В. Межвидовая гибридизация в происхождении видов растений на примере рода *Poa* sensu lato // Успехи современной биологии. – 2015. – Т. 135. – № 1. – С. 21–39. [Nosov NN, Punina EO, Machs EM, Rodionov AV. Interspecies hybridization in the origin of plant species: cases in the genus *Poa* sensu lato. *Biology Bulletin Reviews*. 2015;5(4):366-382.] https:// doi.org/10.1134/S2079086415040088.
- Родионов А.В., Амосова А.В., Беляков Е.А., и др. Генетические последствия межвидовой гибридизации, ее роль в видообразовании и фенотипическом раз-

нообразии растений // Генетика. — 2019. — Т. 55. — № 3. — С. 255—272. https://doi.org/10.1134/ S0016675819030159. [Rodionov AV, Amosova AV, Belyakov EA, et al. Genetic consequences of interspecific hybridization, its role in speciation and phenotypic diversity of plants. *Russian Journal of Genetics*. 2019;55(3): 278-294.] doi: 10.1134/S1022795419030141.

- 5. Whitney KD, Ahern JR, Campbell LG, et al. Patterns of hybridization in plants. *Perspect Plant Ecol Evol Syst.* 2010;12(3):175-182. https://doi.org/10.1016/j. ppees.2010.02.002.
- 6. Kaplan Z, Fehrer J. Molecular identification of hybrids from a former hot spot of *Potamogeton* hybrid diversity. *Aquat Bot.* 2013;105:34-40. https://doi.org/10.1016/j.aquabot.2012.11.002.
- Iida S, Kadono Y, Kosuge K. Maternal effects and ecological divergence in aquatic plants: a case study in natural reciprocal hybrids between *Potamogeton perfoliatus* and *P. wrightii. Plant Species Biol.* 2013;28(1):3-11. https://doi.org/10.1111/1442-1984.12006.
- Ito Y, Tanaka N, Pooma R, Tanaka N. DNA barcoding reveals a new record of *Potamogeton distinctus* (Potamogetonaceae) and its natural hybrids, *P. distinctus* × *P. nodosus* and *P. distinctus* × *P. wrightii* (*P. malainoides*) from Myanmar. *Biodivers Data J.* 2014;2: e1073. https://doi.org/10.3897/bdj.2.e1073.
- Yang T, Zhang TI, Guo YH, Liu X. Identification of hybrids in *Potamogeton*: incongruence between plastid and its regions solved by a novel barcoding marker *PHYB. PLoS ONE.* 2016;11(11):1-12. https://doi. org/10.1371/journal.pone.0166177.
- 10. Arrigo N, Bétrisey S, Graf L, et al. Hybridization as a threat in climate relict *Nuphar pumila* (Nymphaeaceae). *Biodivers Conserv.* 2016;25(10):1863-1877. https://doi.org/10.1007/s10531-016-1165-z.
- 11.Borsch T, Wiersema JH, Hellquist CB, et al. Speciation in North American water lilies: evidence for the hybrid origin of the newly discovered Canadian endemic *Nymphaea loriana* sp. nov. (Nymphaeaceae) in a past contact zone. *Botany*. 2014;92(12):867-882. https://doi.org/10.1139/cjb-2014-0060.
- 12. Nierbauer KU, Kanz B, Zizka G. The widespread naturalisation of *Nymphaea* hybrids is masking the decline of wild-type *Nymphaea* alba in Hesse, Germany. *Flora*. 2014;209(2):122-130. https://doi.org/10.1016/j. flora.2013.12.005.
- 13. Wiecław H, Koopman J. Numerical analysis of morphology of natural hybrids between *Carex hostiana* and the members of *Carex flava* agg. (Cyperaceae). *Nord J Bot.* 2013;31(4):464-472. https://doi.org/10.1111/ j.1756-1051.2013.00095.x.
- 14. Więcław H, Wilhelm M. Natural hybridization within the *Carex flava* complex (Cyperaceae) in Poland: morphometric studies. *Ann Bot Fenn.* 2014;51(3):129-147. https://doi.org/10.5735/085.053.0101.

- Pedersen AT, Nowak MD, Brysting AK, et al. Correction: hybrid origins of *Carex rostrate* var. *borealis* and *C. stenolepis*, two problematic taxa in *Carex* section *Vesicariae* (Cyperaceae). *PLoS ONE*. 2016;11(10): 1-18. https://doi.org/10.1371%2Fjournal.pone.0165430.
- 16. Zalewska-Gałosz J, Jopek M, Ilnicki T. Hybridization in *Batrachium* group: controversial delimitation between heterophyllous *Ranunculus penicillatus* and the hybrid *Ranunculus fluitans* × *R. peltatus. Aquat Bot.* 2015;120:160-168. https://doi.org/10.1016/j. aquabot.2014.03.002.
- 17. Bobrov AA, Zalewska-Gałosz J, Jopek M, Movergoz EA. *Ranunculus schmalhausenii* (section *Batrachium*, Ranunculaceae), a neglected water crowfoot endemic to Fennoscandia – a case of rapid hybrid speciation in postglacial environment of North Europe. *Phytotaxa*. 2015;233(2):101-138. https://doi.org/10.11646/ phytotaxa.233.2.1.
- 18. Ball D, Freeland J. Synchronous flowering times and asymmetrical hybridization in *Typha latifolia* and *T. angustifolia* in northeastern North America. *Aquat Bot*. 2013;104:224-227. https://doi.org/10.1016/j. aquabot.2012.08.006.
- Freeland J, Ciotir C, Kirk H. Regional differences in the abundance of native, introduced, and hybrid *Typha* spp. in northeastern North America influence wetland invasions. *Biol Invasions*. 2013;15(12):2651-65. https:// doi.org/10.1007/s10530-013-0481-4.
- 20. Givnish TJ, Barfuss MH, van Ee B, et al. Phylogeny, adaptive radiation, and historical biogeography in *Bromaliaceae*: insights from an 8-locus plastid phylogeny. *Am J Bot*. 2011;98(5):872-895. https://doi. org/10.3732/ajb.1000059.
- 21. Givnish TJ, Zuluaga A, Spalink D, et al. Monocot plastid phylogenomics, timeline, net rates of species diversification, the power of multi-gene analyses, and a functional model for the origin of monocots. *Am J Bot.* 2018;105(11):1888-1910. https://doi.org/10.1002/ ajb2.1178.
- 22. Sulman JD, Drew BT, Drummond C, et al. Systematics, biogeography, and character evolution of *Sparganium* (Typhaceae): diversification of a widespread, aquatic lineage. *Am J Bot.* 2013;100(10):2023-2039. https://doi.org/10.3732/ajb.1300048.
- 23. Ito Y, Tanaka N, Kim C, et al. Phylogeny of Sparganium (Typhaceae) revisited: non-monophyletic nature of *S. emersum* sensu lato and resurrection of *S. acaule. Plant Syst Evol.* 2016;302(1):129-135. https://doi. org/10.1007/s00606-015-1245-7.
- 24. Su T, Yang JX, Lin YG, et al. Characterization of the complete chloroplast genome of *Sparganium stoloniferum* (Poales: Typhaceae) and phylogenetic analysis. *Mitochondrial DNA B Resour.* 2019;4(1): 1402-1403. https://doi.org/10.1080/23802359.2019. 1598798.

- 25.Leitch IJ, Johnston E, Pellicer J, et al. Plant DNA C-values database. Release 7.1, April 2019. Available from: https://cvalues.science.kew.org/.
- 26. Родионов А.В., Ким Е.С., Пунина Е.О., и др. Эволюция хромосомных чисел в трибах Aveneae и Poeae по данным сравнительного исследования внутренних транскрибируемых спейсеров ITS1 и ITS2 ядерных генов 45S рРНК // Ботанический журнал. – 2007. – Т. 92. – № 1. – С. 57–71. [Rodionov AV, Kim ES, Punina EO, et al. Evolution of chromosome numbers in the tribes Aveneae and Poeae inferred from the comparative analysis of the internal transcribed spacers ITS1 and ITS2 of nuclear 45S rRNA genes. Botanicheskii zhurnal. 2007;92(1):57-71. (In Russ.)]
- 27. Salse J. Deciphering the evolutionary interplay between subgenomes following polyploidy: a paleogenomics approach in grasses. *Am J Bot*. 2016;103(7):1167-1174. https://doi.org/10.3732/ajb.1500459.
- 28. Болховских З.В., Гриф В.Г., Захарьева О.И., Матвеева Т.С. Хромосомные числа цветковых растений. Л.: Наука, 1969. 926 с. [Bolkhovskikh ZV, Grif VG, Zakharyeva OI, Matveeva TS. Khromosomnyye chisla tsvetkovykh rasteniy. Leningrad: Nauka; 1969. 926 р. (In Russ.)]
- 29. Агапова Н.Д., Архарова К.Б., Вахтина Л.И., и др. Числа хромосом цветковых растений флоры СССР: Moraceae-Zygophyllaceae. — СПб.: Наука, 1993. — 427 с. [Agapova ND, Arkharova KB, Vakhtina LI, et al. Chisla khromosom tsvetkovykh rasteniy flory SSSR: Moraceae-Zygophyllaceae. Saint Petersburg: Nauka; 1993. 427 p. (In Russ.)]
- 30. Kim CS, Kim SY, Meon MO. A new record for the Korean flora: *Sparganium fallax* Graebn. (Sparganiaceae). *Korean J Plant Taxonomy*. 2010;40(3):169-173. https://doi.org/10.11110/kjpt.2010.40.3.169.
- Goldblatt P. Polyploidy in angiosperms: monocotyledons. In: Polyploidy. Boston (MA): Springer; 1980. pp. 219-239.
- 32. Anderson E. Introgressive hybridization. London–New York: Hafner Publ. Co.; 1969. 109 p.
- 33. Seitz U, Seitz U. Molecular-weight of ribosomal-RNA precursor molecules and their processing in higherplant cells. Z Naturforsch C Biosci. 1979;34(3-4): 253-258. https://doi.org/10.1515/znc-1979-3-416.
- 34. Garcia S, Kovařík A, Leitch AR, Garnatje T. Cytogenetic features of rRNA genes across land plants: analysis of the plant rDNA database. *Plant J.* 2017;89(5):1020-1030. https://doi.org/10.1111/tpj.13442.
- 35. Родионов А.В., Гнутиков А.А., Коцинян А.Р., и др. Последовательность ITS1-5.8S рДНК-ITS2 в генах 35S рРНК как маркер при реконструкции филогении злаков (сем. Роасеае)// Успехи современной биологии. – 2016. – Т. 136. – № 5. – С. 419–437. [Rodionov AV, Gnutikov AA, Kotsinyan AR, et al. ITS1-5.8S rDNA-ITS2 sequence in 35S rRNA genes as

marker for reconstruction of phylogeny of grasses (Poaceae family). *Biology Bulletin Reviews*, 2017, 7(2):85-102. (In Russ.)]. https://doi.org/10.1134/S2079086417020062.

- 36. Egan AN, Schlueter J, Spooner DM. Applications of next-generation sequencing in plant biology. *Am J Bot.* 2012;99(2):175-185. https://doi.org/10.3732/ ajb.1200020.
- 37. Пунина Е.О., Мачс Э.М., Крапивская Е.Е., Родионов А.В. Полиморфные сайты в транскрибируемых спейсерах генов 35Sp РНК пионов как индикатор происхождения сортов // Генетика. — 2017. — Т. 53. — № 2. — С. 181-191. [Punina EO, Machs EM, Krapivskaya EE, Rodionov AV. Pilymorphic sites in transcribed spacers of 35S rRNA genes as an indicator of origin of the Paeonia cultivars. *Russian Journal of Genetics*. 2017;53(2):202-212.] https:// doi.org/10.1134/S1022795417010112.
- 38. Пунина Е.О., Мачс Э.М., Крапивская Е.Е., и др. Межвидовая гибридизация в роде *Paeonia* (Paeoniaceae): полиморфные сайты в транскрибируемых спейсерах генов 45S pPHK как индикаторы происхождения природных и искусственных гибридов пионов // Генетика. – 2012. – Т. 48. – № 7. – С. 812. [Punina EO, Machs EM, Krapivskaya EE, et al. Interspecific hybridization in the genus *Paeonia* (Paeoniaceae): polymorphic sites in transcribed spacers of the 45S rRNA genes as indicators of natural and artificial peony hybrids. *Russian Journal of Genetics*. 2012;48(7):684-697.] https://doi.org/10.1134/ S1022795412070113.
- 39. Андронова Е.В., Мачс Е.М., Филиппов Е.Г., и др. Филогеография таксонов рода *Сургередіит* (Orchidaceae) на территории России // Ботанический журнал. 2017. Т. 102. № 8. С. 1027-1059. [Andronova EV, Machs EM, Filippov EG, et al. Phylogeography of the Genus *Cyprepedium* (Orchidaceae) taxa in Russia. *Botanicheskii zhurnal.* 2017;102(8): 1027-1059. (In Russ.)]. https://doi.org/10.1134/S0006813617080014.
- 40. Doyle JJ, Doyle JL. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical Bulletin.* 1987;19(1):11-15.
- 41.National Center for Biotechnology Information. Gen-Bank Overview. Available from: https://www.ncbi.nlm. nih.gov/genbank/.
- 42. Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics*. 2014;30(15):2114-2120. https://doi.org/10.1093/ bioinformatics/btu170.
- 43. Aronesty E. Comparison of sequencing utility program. *Open Bioinformatics J.* 2013;7:1-8. https://doi.org/1 0.2174/1875036201307010001.
- 44. Taylor SA, Larson EL, Harrison RG. Hybrid zones: windows on climate change. *Trends Ecol Evol.*

2015;30(7):398-406. https://doi.org/10.1016/j. tree.2015.04.010.

- 45. Vallejo-Marin M, Hiscock SJ. Hybridization and hybrid speciation under global change. *New Phytologist*. 2016;211(4):1170-1187. https://doi.org/10.1111/nph.14004.
- Preston CD, Pearman DA. Plant hybrids in the wild: evidence from biological recording. *Biol J Linn Soc Lond*. 2015;115(3):555-572. https://doi.org/10.1111/ bij.12577.
- 47.Дорофеев П.И. Третичные флоры Западной Сибири. – М., Л.: Издательство Академии наук СССР, 1963. – 346 с. [Dorofeev PI. Tretichnyye flory Zapadnoy Sibiri. Moscow-Leningrad: Izdatel'stvo Akademii Nauk SSSR; 1963. 346 р. (In Russ.)]
- 48.Дорофеев П.И. К систематике третичных Sparganium // Советская палеокарпология: итоги и перспективы / Сб. статей под ред. Г.И. Горецкого, В.П. Гричук. М.: Наука, 1979. С. 53–75. [Dorofeev PI. K sistematike tretichnykh Sparganium. In: (Collected papers) Sovetskaya paleokarpologiya: itogi i perspektivy. Ed. by G.I. Goretskiy, V.P. Grichuk. Moscow: Nauka; 1979. pp. 53-75. (In Russ.)]
- 49.Беляков Е.А., Щербаков А.В., Лапиров А.Г., Шилов М.П. Морфология и экологические особенности Sparganium × longifolium (Typhaceae) в центре Европейской части России // Biosystems Diversity. – 2017. – Vol. 25. – № 2. – Р. 154–161. [Belyakov EA, Shcherbakov AV, Lapirov AG, Shilov MP. Morphology and ecological characteristics of Sparganium × longifolium (Typhaceae) in the Central part of European Russia. Biosystems Diversity. 2017;25(2):154-161. (In Russ.)]. https://doi. org/10.15421/011723.
- 50. Виноградова Ю.К., Галкина М.А. Гибридизация как фактор инвазионной активности чужеродных видов золотарника (Solidago) // Журнал общей биологии. – 2019. – Т. 80. – № 1. – С. 43–56. [Vinogradova YuK, Galkina MA. Hybridization as a factor of invasive activity of alien species of goldenrods (Solidago). *Journal of general biology*. 2019;80(1):43-56. (In Russ.)]. https://doi.org/10.1134/S004445961901007X.
- 51. Tamura K, Nei M. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Mol Biol Evol.* 1993;10(3):512-526. https://doi.org/10.1093/oxfordjournals.molbev.a040023.
- 52. Kumar S, Stecher G, Tamura K. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Mol Biol Evol*. 2016;33(7):1870-1874. https://doi.org/10.1093/molbev/msw054.
- 53. Цвелев Н.Н. Заметки о некоторых гидрофильных растениях флоры СССР // Новости систематики высших растений. 1984. Т. 21. С. 232–242.

[Tzvelev NN. Notulae de florae URSS plants hydrophilis nonnulis. *Novitates systematicae plantarum vascularium*. 1984;21:232-242. (In Russ.)]

- 54. Cook CD, Nicholls MS. A monographic study of the genus *Sparganium*. Part 2: Subgenus *Sparganium*. *Bot Helv*. 1987;97(1):1-44.
- 55. Снакин В.В. Динамика биоразнообразия, дрейф материков и глобализация // Век глобализации. -

2015. –  $\mathbb{N}_{2}$  1. – C. 66–74. [Snakin VV. The Dynamics of biodiversity, continental drift and globalization. *Age of Globalization*. 2015;(1):66-74. (In Russ.)]

56. Chaw SM, Zharkikh A, Sung HM, et al. Molecular phylogeny of extant gymnosperms and seed plant evolution: analysis of nuclear 18S rRNA sequences. *Mol Biol Evol.* 1997;14(1):56-68. https://doi.org/10.1093/oxfordjournals.molbev.a025702.

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