Check for updates

Пролиферативная активность и дифференцировка стволовых клеток почек при суперинвазионном описторхозе (экспериментальное исследование)

Р.М. Урузбаев^{1,2}, В.Г. Бычков¹, Л.В. Вихарева¹, О.А. Молокова¹

¹Тюменский государственный медицинский университет, Тюмень, Россия;
²Многопрофильный клинический медицинский центр «Медицинский город», Тюмень, Россия

Цель. Выявление реакции репликативного потенциала почек при различных формах описторхоза у лабораторных животных.

Материалы и методы. Эксперимент проведен на 60 сирийских хомяках-самцах. I группа (n = 10) — контрольная, II группа (n = 25) — заражение животных метацеркариями *Opisthorchis felineus*, III группа (n = 25) — моделирование суперинвазионной формы описторхоза: заражение 50 личинками *O. felineus*, повторное заражение 50 метацеркариями через 14 и 25 сут. Из эксперимента хомяки выводились посредством передозировки наркоза с декапитацией на 7, 15, 30 сут опыта. Выделение почек с последующим гистологическим исследованием, в т.ч. с использованием гистохимических и иммуногистохимических методов окрашивания. Далее проводилась микроскопия с последующим статистическим анализом результатов.

Результаты. Выявлены количественные характеристики регионарных стволовых клеток, их пролиферативные тенденции и дифференцировка. В корковом и мозговом веществе почек обнаружена экспрессия маркеров CD117, Oct4, CD34 и CD31-позитивных стволовых клеток, которые в дальнейшем дифференцируются в прогениторные клетки с формированием эпителиальных структур в виде канальцев. В клубочках наблюдали картину васкулогенеза с увеличением сосудистых петель.

Заключение. O. felineus секретом инициирует активацию стволовых клеток в почечных канальцах и перицитах микроциркуляторного русла. Наблюдается выраженная пролиферация переходного эпителия лоханок и начальных отделов мочеточника. Кроме этого, под действием секретома паразита наблюдается выраженная пролиферация стволовых клеток непосредственно в петлях клубочков.

Ключевые слова: почки; описторхоз; 0. felineus; секретом; сирийский хомяк

Как цитировать:

Урузбаев Р.М., Бычков В.Г., Вихарева Л.В., Молокова О.А. Пролиферативная активность и дифференцировка стволовых клеток почек при суперинвазионном описторхозе (экспериментальное исследование) // Российский медико-биологический вестник имени академика И.П. Павлова. 2021. Т. 29. № 2. С. 213–218. DOI: https://doi.org/10.17816/PAVLOVJ57046

Рукопись получена: 27.12.2020



Рукопись одобрена: 11.06.2021

Опубликована: 30.06.2021

Experimental study on the proliferating activity and differentiation of renal stem cells in superinvasive opisthorchiasis

Rinat M. Uruzbaev^{1,2}, Vitaly G. Bychkov¹, Larisa V. Vikhareva¹, Olga A. Molokova¹

¹Tyumen State Medical University, Tyumen, Russia;

²Multipurpose Clinical Medical Center «Medical City», Tyumen, Russia

AIM: This study aimed to identify the replication potential of the kidneys in different forms of opisthorchiasis in laboratory animals.

MATERIALS AND METHODS: An experiment was conducted on 60 Syrian male hamsters. The first group was set as the control (n = 10), the second group (n = 25) was infected with metacercariae (*Opisthorchis felineus*), and the third group (n = 25) was a model of a superinvasive form of opisthorchiasis infection with 50 0. felineus larvae and repeated infection with 50 metacercariae in 14 and 25 days. The hamsters were withdrawn from the experiment on days 7, 15, and 30 via an overdose of narcosis and decapitation. The kidneys were isolated and histologically examined through histochemical and immunohistochemical staining methods. Microscopy was conducted, and results were statistically analyzed.

RESULTS: The quantitative characteristics, proliferation tendencies, and differentiation of regional stem cells were identified. In the cortical and medullary substance of the kidneys, CD117, Oct4, and CD34 markers were expressed, and CD31positive stem cells further differentiated to progenitor cells. Epithelial structures developed in the form of tubules. In the glomeruli, vasculogenesis occurred, and the number of vascular loops increased.

CONCLUSION: 0. felineus secretome initiated the activation of stem cells in the renal tubules and pericytes of a microcirculatory network. The transitional epithelium of the renal pelvis and the initial parts of the ureter proliferated. Under the action of the secretome of parasites, stem cells proliferated directly in glomerular loops.

Keywords: kidneys; opisthorchiasis; Opistorchis felineus; secretome; Syrian hamster

To cite this article:

Uruzbaev RM, Bychkov VG, Vikhareva LV, Molokova OA. Experimental study on the proliferating activity and differentiation of renal stem cells in superinvasive opisthorchiasis. I.P. Pavlov Russian Medical Biological Herald. 2021;29(2):213–218. DOI: https://doi.org/10.17816/PAVLOVJ57046

Accepted: 11.06.2021

214

ORIGINAL STUDIES

Cat liver fluke or Opisthorchis felineus is responsible for the occurrence of a parasitic disease known as opisthorchiasis (a type of trematode infection). The disease has a systemic characteristic: presence of lesions in econiches (liver, pancreas, gallbladder), which is where parasites vegetate [1,2]. Evidenced by clinical and pathoanatomical practice, helminths induce significant alterations in the organs and systems where they do not parasitize, that is, alterations also occur outside the econiches of Opisthorchis spp., for example, in lungs, kidneys, and so on. Manifestations are mostly expressed in hypereosinophilic syndrome resulting from numerous early and late super invasions [3,4]. Opisthorchis spp. contain granulin proteins in their secretome that exerts an evident proliferating effect, which causes mutations in numerous proliferative genes [5,6]. As shown earlier in laboratory animals, Opisthorchis invasion expresses numerous genes, namely c-Kit, APC, K-ras, B-raf, WE6F, VEGFR, and others, which is explained by the reaction of parasites to provide their individuals with a trophic substrate - cholangiocytes. According to the rule of H. Leduc (1964), regional stem cells of the liver (replicative potential) are induced to proliferate and differentiate to cholangiocellular and hepatocellular differons. Simultaneously (15th day of the experiment), the following active processes of neoangiogenesis were observed: vasculogenesis (formation of vessels from progenitor cells) and angiogenesis (formation of vessels from the vascular network) due to the kinetic processes of the endothelium and pericytes of pre-existing capillaries [7].

Clinically, there is a paucity of studies regarding the kidney's condition in opisthorchiasis. Patients with chronic opisthorchiasis present with proteinuria, hematuria, and cylindruria; many researchers assign these manifestations to the allergic reaction to *Opisthorchis spp.* [8,9]. Cases of immune glomerulonephritis with nephrotic syndrome have also been described in acute opisthorchiasis; these conditions were manifested by pronounced nephropathy [10,11]. Morphogenetic studies on the initiation and kinetics of native kidney stem cells in opisthorchiasis have not been conducted, and the reaction of the replicative potential of kidneys to the initiating substrate of *Opisthorchis spp.* – secretome (granulin) – remains unknown. Moreover, this reactions is important for identifying repair processes in the partial resections of this organ.

This study **aimed** to identify the reactions of replicative potential of kidneys in different forms of opisthorchiasis in laboratory animals.

MATERIALS AND METHODS

The experiment was conducted on 60 Syrian male hamsters (*Syrian golden hamster*) weighing 95.0 ± 10.0 g. The animals were divided into three groups:

I – control group – 10 animals;

II – experimental group – 25 hamsters infected with 50 *O. felineus* metacercariae;

III – modeling of super invasive form of opisthorchiasis (SF) – infection with 50 0. felineus larvae followed by repeated infections with 50 metacercariae on the 14^{th} and 25^{th} days.

Metacercariae were isolated from nerflings (Leuciscus idus) of one biotope by G.A. Glazkov's method [12]. All manipulations with the animals were conducted in compliance with the Declaration of Helsinki's guidelines on the humane treatment of animals and Order of HM RF No 267 of June 19, 2003 «On approval of rules of laboratory practice». Hamsters were withdrawn from the experiment by the overdose of narcosis with subsequent decapitation on the 7th, 15th, and 30th days.

After standard histological manipulations, the sections of renal tissue were stained with hematoxylin and eosin by Van Gieson method. De-embedding, antigen retrieval, and immunohistochemical reactions were performed with the use of Bond-Max autostainer (Leica Biosystems, USA) in accordance with the standard protocols. Immunohistochemical examinations were conducted using following markers: CD34 (Lab Vision Corporation, USA; clone - QBEnd/10, Cell Marque), CD31 (Novokastra, USA; clone – JC70, Cell Margue), Oct-4 (Lab Vision Corporation, USA; clone - MRQ-10, Cell Marque), CD117 (Lab Vision Corporation, USA; clone – YR145, Cell Marque), and Ki-67 (RTU, CША; clones – MIB-1, Agilent/Dako). Results of immunohistochemical reactions were evaluated by semi-quantitative and quantitative characteristics: intensity of reactions were assessed on a scale from 0 to 3 points (0 - no reaction)2 - moderate reaction, 3 - evident reaction) and by the number of positively stained cells in 1 microscopic field (microscopic field; 400× magnification). Positively stained cells were counted in 10 microscopic fields at 400× magnification by calculating the arithmetic mean. Coverglass preparations were studied on Axio Lab. A1 (Carl Zeiss Microscopy, Germany) with a further morphometric evaluation of quantitative parameters.

Statistical processing of the results was performed using the variation statistics method with the use of application software package Office Excel 2007 (Microsoft, USA) on IBMPC/AT Pentium IV in the Windows 7.0 environment. The results are presented in shares (%), median (Me), and lower (Q25) and upper (Q75) quartiles. Non-parametric Mann–Whitney method was used for comparison. p < 0.05 was assigned as the level of statistical significance for this study parameters.

RESULTS AND DISCUSSION

On the seventh day after infection, macroscopic and histological examinations revealed minimal alterations in the kidneys. Morphological alterations were similar and corresponded to the morphological picture observed in the control group. In the group with SF, a focal interstitial edema and minor lymphoid infiltration in the cortical substance were noted; moreover, no significant changes were found in the glomeruli.

On the 15th day of the experiment, the microscopic picture of kidneys of animals of groups II and III significantly differed from that of group I (control): morphological changes of different extents of evidence were recorded in the different parts of nephron. An increase in the size of the glomeruli with a moderate compression of the capillary loops and the focal sclerosis of capsule and capillary loops in some of them were observed. In the cortical substance, vacuolization of the cytoplasm in some epithelial cells of convoluted tubules was noted. In the proximal tubules, the signs of hyaline-drop dystrophy and signs of focal necrobiosis were present. A partial desquamation of epithelium with the formation of small denudation foci, as well as the foci of fibrosis and sclerosis was recorded. In SF, up to 28% of glomeruli were hypertrophied, whereas such alteration of glomeruli was observed in late stages in group II. Here, up to 10% of glomeruli were collapsed and had initial signs of glomerulosclerosis accompanied by stasis in capillaries on the renal corpuscle.

An increase in the CD31 marker was observed in the glomerular apparatus in the stroma of kidneys and in the walls of tubules in group II (single infection with metacercariae *O. felineus*). a moderate expression of the CD34 marker indicated the activation of stem cells in these structures (Figure 1A, B, C).



Fig. 1. SF group on the 15th day of the experiment: A – evident proliferation of renal tubules. Hematoxylin and eosin stain. Magnification × 40; B – the membrane expression of CD34 marker in renal tubules. Immunohistochemical reaction with CD34. Magnification × 20; C – cytoplasmic expression of CD31 marker in glomeruli. Immunohistochemical reaction with CD31. Magnification × 10; D – close location of glomeruli with moderate sclerosis. Hematoxylin and eosin stain. Magnification x 20.

Besides, an increase in the number of renal capillaries in the glomerulus itself was detected (group I: 31.00 ± 12.74). In group III (superinvasive form of opisthorchiasis), an evident proliferation of stem and progenitor cells in the forming vessels and proliferation of the epithelial structures of tubules were observed (Table 1).

On the 30th day of the experiment, granular cylinders were observed in the lumen of convoluted tubules. Inflammatory infiltrates were present in the form of lymphomononuclear cells near glomeruli. In the preparations, focal interstitial fibrosis was identified and it was more evident than in the experimental group. The segmentation of glomeruli accompanied by an increase in the number of capillaries of the renal corpuscle was observed. The glomeruli were hypertrophied, located close to each other, and a part of them showed evident sclerosis as compared with the second experimental group (Figure 1D).

In the nephrons, an increase in the diameter of glomerulus and its surface area was recorded as compared to the group of primary infection with *Opisthorchis spp*. (Figure 2A). Here, an evident expression of CD31 and Oct-4 markers (Figure 2A) and evident proliferation of multilayered transitional epithelium in the pelvis (Figure 2D) were noted.

During the experiment, a higher regenerator potential was identified in the walls of renal tubules as compared with glomeruli because in super invasive opisthorchiasis, the expression of CD34, Oct-4, CD117 markers was observed in the walls of renal tubules. In the examination of proliferative activity, the expression of Ki-67 marker in these structures was higher than that in the glomerular apparatus.

217

Markers	Stroma			Parenchyma		
	Group I	Group II	Group III	Group I	Group II	Group III
СD34, кл/мм ²	3.60 ± 1.24	11.70 ± 2.78**	17.80 ± 5.87**	2.70 ± 1.25	10.70 ± 0.43**	11.80 ± 0.54*
	(1.76 - 4.84)	(8.92 - 14.48)	(11.93 - 23.67)	(1.45 - 3.95)	(10.27 - 11.13)	(11.26 - 12.34)
Oct4, кл/мм ²	4.60 ± 3,85	12.70 ± 5.41*	39.50 ± 16.19*	0.40 ± 0.20	1.20 ± 0.09**	1.90 ± 0.95*
	(0.75 - 8,45)	(7.29 - 18.11)	(23.31 - 55.69)	(0.20 - 0.60)	(1.11 - 1.29)	(0.95 - 2.85)
CD117, кл/мм ²	1.06 ± 0.78	7.08 ± 3.48**	19.97 ± 7.93**	1.45 ± 0.68	2.70 ± 1.10*	3.60 ± 3.50**
	(0.28 - 1.84)	(3.60 - 10.56)	(12.04 - 27.90)	(0.77 - 2.13)	(1.60 - 3.80)	(0.10 - 7.10)
CD31, кл/мм ²	2.10 ± 0.74	8.40 ± 0.68**	14.55 ± 5.65*	4.80 ± 0.80	8.40 ± 1.08**	10.80 ± 0.45*
	(1.36 - 2.84)	(7.72 - 9.08)	(8.90 - 20.20)	(4.0 - 5.6)	(7.32 - 9.48)	(10.35 - 11.25)
Ki-67, %	1.6	3.0**	9.0*	0.8	2.4*	7.6**

Notes: * – statistically significant differences as compared to group I, p < 0.05, ** – statistically significant differences as compared to group I, p < 0.01. Each marker was analyzed in 10 fields at × 400 magnification



Fig. 2. SF group on the 30^{th} day of experiment: A – increase in the number of loops in glomeruli. Hematoxylin and eosin stain. Magnification ×40; B – membrane expression of CD31 marker in renal tubules. Immunohistochemical reaction with CD31. Magnification $10 \times$; C – nuclear expression of Oct4 marker in renal glomeruli. III group. Immunohistochemical reaction with Oct4. Magnification ×20; D – proliferation of transitional epithelium of pelvis. Hematoxylin and eosin stain. Magnification ×40.

The practice of postmortem examinations of patients who died with super invasive opisthorchiasis in a hyperendemic focus showed a rare participation of kidneys as the main component of the direct cause of death. An exception is the evident systemic inflammation (sepsis) and the development of chronic kidney disease against the background diffuse immunocomplex glomerulonephritis. It can be suggested that active vasculogenesis, that is, CD-34–positive (endothelial) cells, generally permits the enrichment of glomeruli with additional functional structures and enhance the compensatory potentials of organs. It should be noted that in super invasive opisthorchiasis, neoangiogenesis occurs and is subdivided into angiogenesis – trichotomic branching of existing vascular formations – and vasculogenesis – development of vessels *de novo* from

stem cells. The latter process is observed in various organs, regardless of the place of vegetation of the parasite [2,7].

Pericytes play an important role in organizing the microvasculature of the kidneys. As is it known, they actively express vascular endothelial growth factor; therefore, the identified activity of CD31 marker reaches three points, which confirms the fact that podocytes, forming an expensive network around capillaries, actively participate in the angiogenesis and in the maturation of vessels upon exposure to *0. felineus* secretome. In general, the number of loops in glomeruli increases by 24.3% (67.4 \pm 3.6). Besides with the use of histochemical stains, it was confirmed that moderate sclerotic alterations around glomerular apparatus result from the activation of fibroblastic differon.

CONCLUSION

It was found that O. felineus secretome initiates the activation of stem cells in renal tubules and of the pericytes of microvasculature. The active proliferation of transitional epithelium of pelvis and of initial parts of ureter was found. Besides, under the action of parasite's secretome, a pronounced proliferation of stem cells occurred directly in the glomerular loops that may have exerted a protector effect in

ЛИТЕРАТУРА

1. Бычков В.Г., Иванюженко Н.Д., Шевчук О.Н. Описторхоз в сочетании с алкогольной интоксикацией у людей и в эксперименте. Клинико-морфологическое исследование // Медицинская паразитология и паразитарные болезни. 1986. № 5. С. 30-33.

2. Бычков В.Г., Чернов И.А., Хадиева Е.Д., и др. Закономерности пролиферативных реакций при описторхозе: их роль в канцерогенезе и регенерации // Морфология. 2018. Т. 153, № 3. С. 52.

3. Соловьева О.Г. Клинико-патогенетические особенности заболеваний легких при суперинвазионном описторхозе у населения Среднего Приобья. Дис. ... д-ра мед. наук. Тюмень; 2011.

 Куликова С.В. Структурно-функциональные изменения сердца и антропометрических показателей у больных суперинвазионным описторхозом. Дис.... канд. мед. наук. Тюмень, 2011.
 Пахарукова М.Ю. Структурно-функциональная организация

5. Пахарукова М.Ю. Структурно-функциональная организация системы метаболизма ксенобиотиков у возбудителя описторхоза Opisthorchis felineus (Rivolta, 1884). Дис. ... д-ра биол. наук. Новосибирск; 2016.

6. Помазной М.Ю. Транскриптомный анализ трематоды Opisthorchis felineus. Дис. ... канд. биол. наук. Новосибирск; 2015.

REFERENCES

1. Bychkov VG, Ivanyuzhenko ND, Shevchuk ON. Opistorkhoz v sochetanii s alkogol'noy intoksikatsiyey u lyudey i v eksperimente. Kliniko-morfologicheskoye issledovaniye. *Meditsinskaya Parazitologiya i Parazitarnyye Bolezni.* 1986;(5):30-3. (In Russ).

2. Bychkov VG, Chernov IA, Khadiyeva YeD, et al. Patterns of proliferative reactions in opisthorchiasis: their role in carcinogenesis and regeneration. *Morphology.* 2018;153(3):52. (In Russ).

3. Solov yeva OG. Kliniko-patogeneticheskiye osobennosti zabolevaniy legkikh pri superinvazionnom opistorkhoze u naseleniya Srednego Priob'ya [dissertation]. Tyumen; 2011. (In Russ).

4. Kulikova SV. Strukturno-funktsional'nyye izmeneniya serdtsa i antropometricheskikh pokazateley u bol'nykh superinvazionnym opistorkhozom [dissertation]. Tyumen, 2011. (In Russ).

5. Pakharukova MYu. Strukturno-funktsional'naya organizatsiya sistemy metabolizma ksenobiotikov u vozbuditelya opistorkhoza Opisthorchis felineus (Rivolta, 1884) [dissertation]. Novosibirsk; 2016. (In Russ).

6. Pomaznoy MYu. Transkriptomnyy analiz trematody Opisthorchis felineus [dissertation]. Novosibirsk; 2015. (In Russ).

7. Bychkov VG, Zolotukhina VM, Khadieva ED, et al. Hypereosinophilic

ОБ АВТОРАХ

*Ринат Маратович Урузбаев — к.м.н., доцент кафедры патанатомии и судебной медицины, Тюменский государственный медицинский университет; врач-патологоанатом отделения онкоморфологии и ВТМИ патологоанатомического бюро, Многопрофильный клинический медицинский центр «Медицинский город», Тюмень, Россия. ORCID: https://orcid.org/0000-0001-6883-0543 e-mail: uruzbaevrm@mail.ru

Виталий Григорьевич Бычков — д.м.н., профессор, профессор кафедры патанатомии и судебной медицины, Тюменский государственный медицинский университет, Тюмень, Россия. ORCID: https://orcid.org/0000-0002-0211-2669

Лариса Владимировна Вихарева — д.м.н., профессор, зав. кафедрой анатомии человека, топографической анатомии и оперативной хирургии, Тюменский государственный медицинский университет. Тюмень, Россия. ORCID: https://orcid.org/0000-0001-6864-4417

Ольга Александровна Молокова — д.м.н., доцент, профессор кафедры патанатомии и судебной медицины, Тюменский государственный медицинский университет, Тюмень, Россия. ORCID: https://orcid.org/0000-0002-3736-3089 the critical conditions of an organism of parasite's host.

ADDITIONALLY

Conflict of interests. The authors declare no actual and potential conflict of interests, which should be stated in connection with publication of the article.

Participation of authors: *R.M. Uruzbaev* — collection, translation and analysis of material, writing the text, *V.G. Bychkov, L.V. Vikhareva* — editing, *O.A. Molokova* — concept of the review, editing.

7. Bychkov V.G., Zolotukhina V.M., Khadieva E.D., et al. Hypereosinophilic Syndrome, Cardiomyopathies, and Sudden Cardiac Death in Superinvasive Opisthorchiasis // Cardiology Research and Practice. 2019. Vol. 2019. P. 4836948. doi: 10.1155/2019/4836948

8. Рычкова Е.К., Козина О.И., Сазонова Л.В. Поражение почек при описторхозе. В сб.: Материалы 1-го съезда терапевтов Тюменской области. Тюмень; 1970. С. 88-90.

9. Озерецковская Н.Н., Зальнова Н.С., Тумольская Н.И. Клиника и лечение гельминтозов. Инфекционные и паразитарные заболевания. Л.: Медицина; 1985.

10. Шульцев Т.П., Цаленчук Я.П., Ольхин В.А. Нефротический синдром при описторхозе // Клиническая медицина. 1973. Т. 51, № 8. С. 132-135. 11. Калюжин В.В., Коваль В.В., Калюжина Е.В., и др. Поражение почек при хроническом описторхозе // Медицинская паразитология и паразитарные болезни. 2008. № 4. С. 14-16.

12. Глазков Г.А. Выделение метацеркариев некоторых трематод из пораженной ткани рыб методом переваривания в искусственном желудочном соке. Болезни и паразиты рыб ледовитоморской провинции (в пределах СССР). Томск; 1979. С. 72-82.

Syndrome, Cardiomyopathies, and Sudden Cardiac Death in Superinvasive Opisthorchiasis. *Cardiology Research and Practice*. 2019;2019:4836948. (In Russ). doi: 10.1155/2019/4836948

8. Rychkova EK, Kozina OI, Sazonova LV. Porazheniye pochek pri opistorkhoze. In: *Materialy 1-go s'yezda terapevtov Tyumenskoy oblasti.* Tyumen'; 1970. P. 88-90. (In Russ).

9. Ozeretskovskaya NN, Zal'nova NS, Tumol'skaya NI. *Klinika i lecheniye gel'mintozov. Infektsionnyye i parazitarnyye zabolevaniya.* Leningrad: Meditsina; 1985. (In Russ).

10. Shul'tsev TP, Tsalenchuk YaP, Ol'khin VA. Nefroticheskiy sindrom pri opistorkhoze. *Klinicheskaya Meditsina*. 1973;51(8):132-5. (In Russ).

11. Kalyuzhin VV, Koval^{*} VV, Kalyuzhina EV, et al. Porazheniye pochek pri khronicheskom opistorkhoze. *Meditsinskaya Parazitologiya i Parazitarnyye Bolezni*. 2008;(4):14-6. (In Russ).

12. Glazkov GA. Vydeleniye metatserkariyev nekotorykh trematod iz porazhennoy tkani ryb metodom perevarivaniya v iskusstvennom zheludochnom soke. *Bolezni i parazity ryb ledovitomorskoy provintsii (v predelakh SSSR)*. Tomsk; 1979. P. 72-82. (In Russ).

AUTHORS INFO

*Rinat M. Uruzbaev — MD, Cand.Sci.(Med.), Associate Professor of the Pathological Anatomy and Forensic Medicine Department, Tyumen State Medical University; Pathologist of the Oncomorphology and HTMI Department, the Pathological Bureau Multidisciplinary, Clinical Medical Center Medical City'. ORCID: https://orcid.org/0000-0001-6883-0543 e-mail: uruzbaevrm@mail.ru

Vitaly G. Bychkov — MD, Dr.Sci.(Med.), Professor, Professor of the Pathological Anatomy and Forensic Medicine Department, Tyumen State Medical University, Tyumen, Russia. ORCID: https://orcid.org/0000-0002-0211-2669

Larisa V. Vikhareva — MD, Dr.Sci.(Med.), Professor, Head of the Human Anatomy, Topographic Anatomy and Operative Surgery Department, Tyumen State Medical University, Tyumen, Russia. ORCID: https://orcid.org/0000-0001-6864-4417

Olga A. Molokova — MD, Dr. Sci. (Med.), Associate Professor, Professor of the Pathological Anatomy and Forensic Medicine Department, Tyumen State Medical University, Tyumen, Russia. ORCID: https://orcid.org/0000-0002-3736-3089