

DOI: <https://doi.org/10.17816/PED1535-25>

Diagnosis and treatment of infection specific to the perinatal period (Draft clinical recommendations for discussion by neonatologists and pediatricians)

Dmitry O. Ivanov¹, Alexandra S. Panchenko¹, Larisa A. Fedorova¹, Ekaterina N. Balashova², Elena V. Bem¹, Nazar M. Zelenin¹, Irina V. Myznikova¹, Marina I. Levadneva¹, Anna S. Nabieva¹, Svetlana E. Pavlova¹, Larisa A. Romanova¹, Galina N. Chumakova¹, Ekaterina E. Yakovleva¹, Tamara V. Belousova³, Irina Ya. Izvekova³, Dmitry Yu. Ovsyannikov⁴

¹ Saint Petersburg State Pediatric Medical University, Saint Petersburg, Russia;

² Research Center for Obstetrics, Gynecology and Perinatology, Moscow, Russia;

³ Novosibirsk State Medical University, Novosibirsk, Russia;

⁴ Peoples' Friendship University of Russia, Moscow, Russia

ABSTRACT

Perinatal infections occupy a leading place among the causes of neonatal morbidity, maternal and perinatal mortality. Infections are among the main causes of termination of pregnancy and premature birth. The practical recommendations presented in the work are intended for doctors of obstetric institutions in order to make a clinical diagnosis of an infection specific to the perinatal period, the tactics of examination and treatment of newborn children. The clinical recommendations correspond to the latest scientific data on the topic, contain information that is applied to the practical activities of a neonatologist, intensive care specialist and pediatrician. These clinical recommendations contain information about infections specific to the perinatal period, including the definition, frequency of occurrence, etiology of infections, pathogenetic mechanisms of disease development. Numerous high-risk factors for infection of the fetus and newborn are described in detail. The document discusses and proposes the classification of the disease, the criteria for the adoption of the diagnosis. The features of the clinical picture of the disease are described, it is noted that the inflammatory process in a newborn child can be localized in any organ or acquire a systemic (generalized) character, in some cases, the ingress of an infectious agent into a macroorganism is not necessarily accompanied by clinical manifestations, which indicates an asymptomatic or subclinical course of infection. The recommendations provide advanced laboratory and instrumental diagnostics. The stages of treatment are described, including the choice and correction of antibacterial therapy, taking into account the peculiarities of the mother's anamnesis, the child's gestation period and the etiology of the disease. These clinical recommendations have been prepared taking into account the level of credibility of the recommendations and the level of reliability of the evidence. These practical recommendations are offered for public discussion and are posted in full on the website of the Ministry of Health of the Russian Federation.

Keywords: newborns; infection specific to the perinatal period; diagnosis; treatment; practical guidelines; clinical guidelines.

To cite this article

Ivanov DO, Panchenko AS, Fedorova LA, Balashova EN, Bem EV, Zelenin NM, Myznikova IV, Levadneva MI, Nabieva AS, Pavlova SE, Romanova LA, Chumakova GN, Yakovleva EE, Belousova TV, Izvekova IYa, Ovsyannikov DYU. Diagnosis and treatment of infection specific to the perinatal period (Draft clinical recommendations for discussion by neonatologists and pediatricians). *Pediatrician (St. Petersburg)*. 2024;15(3):5–25. DOI: <https://doi.org/10.17816/PED1535-25>

DOI: <https://doi.org/10.17816/PED1535-25>

Диагностика и лечение инфекции, специфичной для перинатального периода (Проект клинических рекомендаций для обсуждения неонатологами и педиатрами)

Д.О. Иванов¹, А.С. Панченко¹, Л.А. Федорова¹, Е.Н. Балашова², Е.В. Бем¹, Н.М. Зеленин¹, И.В. Мызникова¹, М.И. Леваднева¹, А.С. Набиева¹, С.Е. Павлова¹, Л.А. Романова¹, Г.Н. Чумакова¹, Е.Е. Яковлева¹, Т.В. Белоусова³, И.Я. Извекова³, Д.Ю. Овсянников⁴

¹ Санкт-Петербургский государственный педиатрический медицинский университет, Санкт-Петербург, Россия;

² Национальный медицинский исследовательский центр акушерства, гинекологии и перинатологии им. акад. В.И. Кулакова, Москва, Россия;

³ Новосибирский государственный медицинский университет, Новосибирск, Россия;

⁴ Российский университет дружбы народов им. Патриса Лумумбы, Москва, Россия

АННОТАЦИЯ

Перинатальные инфекции занимают ведущее место среди причин неонатальной заболеваемости, материнской и перинатальной смертности. Инфекции относятся к основным причинам прерывания беременности и преждевременных родов. Практические рекомендации, представленные в работе, предназначены для врачей родовспомогательных учреждений с целью постановки клинического диагноза инфекции, специфичной для перинатального периода, тактики обследования и лечения новорожденных детей. Клинические рекомендации соответствуют последним научным данным по теме, содержат информацию, которая носит прикладной характер для практической деятельности врача неонатолога, реаниматолога и педиатра. В данных клинических рекомендациях содержится информационная справка об инфекциях, специфических для перинатального периода, включающая в себя определение, частоту встречаемости, этиологию инфекций, патогенетические механизмы развития заболевания. Подробно описаны многочисленные факторы высокого риска инфицирования плода и новорожденного. В документе обсуждается и предлагается классификация заболевания, критерии установления диагноза. Описаны особенности клинической картины болезни, отмечено, что воспалительный процесс у новорожденного ребенка может локализоваться в каком-либо органе или приобретать системный (генерализованный) характер, в ряде случаев попадание инфекционного агента в макроорганизм не обязательно сопровождается клиническими проявлениями, что свидетельствует о бессимптомном или субклиническом течении инфекции. В рекомендациях представлена расширенная лабораторная и инструментальная диагностика. Расписаны этапы лечения, включая выбор и коррекцию антибактериальной терапии, с учетом особенностей анамнеза матери, срока гестации ребенка и этиологии заболевания. Данные практические рекомендации подготовлены с учетом уровня убедительности рекомендаций и уровня достоверности доказательств, предлагаются к обсуждению общественности и в полном виде представлены на сайте Минздрава России.

Ключевые слова: новорожденные; инфекция, специфичная для перинатального периода; диагностика; лечение; практические рекомендации; клинические рекомендации.

Как цитировать

Иванов Д.О., Панченко А.С., Федорова Л.А., Балашова Е.Н., Бем Е.В., Зеленин Н.М., Мызникова И.В., Леваднева М.И., Набиева А.С., Павлова С.Е., Романова Л.А., Чумакова Г.Н., Яковлева Е.Е., Белоусова Т.В., Извекова И.Я., Овсянников Д.Ю. Диагностика и лечение инфекции, специфичной для перинатального периода (Проект клинических рекомендаций для обсуждения неонатологами и педиатрами) // Педиатр. 2024. Т. 15, № 3. С. 5–25.
DOI: <https://doi.org/10.17816/PED1535-25>

INTRODUCTION

Infections specific to the perinatal period are one of the most burning issues in modern perinatology, as they are the main cause of morbidity, as well as of maternal and perinatal mortality. Within the context of modern realities, a physician at an obstetric facility needs clear algorithms of actions for an accurate diagnosis, as well as patient treatment strategy that comply with the principles of evidence-based medicine. The recommendations below were compiled exactly to address this issue.

DEFINITION

Infections specific to the perinatal period (IPPs) are infectious processes (inflammations) occurring in various organs and/or systems of the fetus during the period from the 22nd full week (154th day) or the newborn up to the 7th day of extrauterine life (168 hours). These infections are characterized by clinical and pathomorphological changes typical for infectious diseases, which can be detected either antenatally or after birth [5, 16, 17, 30].

According to the International Statistical Classification of Diseases and Related Health Problems, 10th revision, IPPs are coded as P39.8 Other specified infections specific to the perinatal period; P39.9 Infection specific to the perinatal period, unspecified; P37.9 Congenital infectious or parasitic disease, unspecified.

When a perinatal, congenital, or parasitic infection is identified by etiology, the corresponding nosological codes are used: P23, P37.0–37.5, and P39.0–39.4.

EPIDEMIOLOGY

From 2017 to 2020, the incidence of perinatal infections (IPPs) in newborns in the Russian Federation was approximately 1.4% [18]. In obstetric hospitals, the infectious morbidity rate for premature infants weighing 1000 g or more was 8%, while for infants with extremely low birth weight (ELBW), it was around 27%. The mortality rates were 1.6% and 21%, respectively. Infections specific to the perinatal period are reported in 2.3% of live-born children and account for 6% of early neonatal morbidity [33]. IPPs are to be recorded and registered in the infectious disease register at the location of detection, within medical institutions and territorial bodies responsible for federal state sanitary and epidemiological supervision.

DISEASE ETIOLOGY AND PATHOGENESIS

Perinatal infection is the leading cause of morbidity, maternal and perinatal mortality. Infection is the main cause of pregnancy termination and premature birth.

The term “perinatal infections” is usually applied to infections transmitted from a mother to a child during intrauterine development (intrauterine/congenital), during childbirth (perinatal, or intranatal), immediately after childbirth (postnatal) with their development in the early neonatal period of life.

The causative agents of perinatal infections are diverse [5, 18, 24, 33, 74, 81–83, 119, 122]. These may be bacteria (Group *B streptococcus*, *Escherichia coli*, *Enterobacter aerogenes*, *Klebsiella* spp., *Pseudomonas aeruginosa*, *Staphylococcus epidermidis*, *Staphylococcus aureus*, *Haemophilus influenza*, *Klebsiella pneumoniae*, *Streptococcus pneumoniae*, *Treponema pallidum*, *Listeria monocytogenes*, *Mycobacterium tuberculosis*), viruses (*Herpes simplex virus*, *Cytomegalovirus*, *Respiratory syncytial virus*, *Rubella*, etc.), atypical pathogens (*Chlamydia trachomatis*, *Ureaplasma urealyticum*, *Mycoplasma hominis*, *Mycoplasma pneumonia*, etc.), fungi (*Candida* spp.), protozoa (*Toxoplasma gondii*).

The fundamental difference of IPP is its proven relation of the infection to the mother's body. In a normal pregnancy, the fetus is well protected by the placenta and amniotic membranes from any various pathogenic and opportunistic microorganisms. A pathological course of pregnancy, caused primarily by infectious causes, facilitates the penetration of pathogens into the fetus, crossing the placental barriers (transplacentally or ascendingly), contributing to the development of an inflammatory reaction in the fetus in microbial invasion.

In the vast majority of cases, fetal infection begins with damage to the placenta. The infectious and inflammatory process in the placenta and in the amniotic membranes — intra-amniotic infection — adversely affects vital functions of the fetus. Through the bloodstream, pathogenic microorganisms enter the chorionic villi of the placenta, where they become fixed and form an inflammation focus.

The negative effect of antenatal infection on the fetoplacental complex depends on several factors, including the gestational age at the time of infection, the type of pathogen, its virulence, the extent of seeding, whether the infection is primary or secondary in the pregnant woman, the route of infection to the fetus, the degree of prevalence, the intensity of the inflammatory process, and the severity and nature of changes in the immune response of a pregnant woman.

Antenatal infectious agent achieves the fetus hematogenously (transplacentally) or through infected amniotic fluid by an ascending (from the cervical canal) or descending (from the fallopian tubes) route, by transmembranal route (through the amniotic membranes in endometritis, placentitis), iatrogenically (during medical manipulations), or by contact [13, 32, 33]. Most microorganisms, having entered the uterine cavity, stimulate the infiltration and activation of neutrophils and lead to

an increase in the synthesis and release of proinflammatory cytokines, prostaglandins and matrix metalloproteinases. These changes contribute to cervical maturation, membrane rupture, uterine contractions, and premature amniotic fluid discharge [4, 5, 33, 41].

Perinatal infection and the development of a systemic inflammatory process in a fetus with activation of its immune system often leads to premature birth and proven intra-amniotic infection [76, 77, 95, 114]. Newborns from mothers with chorioamnionitis are at high risk in terms of development of neonatal sepsis, bronchopulmonary dysplasia, intraventricular hemorrhage, periventricular leukomalacia, and neonatal death.

High-risk factors of fetal infection are inflammatory diseases of the uterus and its appendages, colpitis, bacterial vaginosis, oligohydramnios or polyhydramnios, complicated obstetric and gynecological history (repeated artificial termination of pregnancy with complicated post-abortion period, habitual miscarriage, congenital malformations and antenatal death of a fetus), placental insufficiency, intrauterine growth and fetal development retardation, use of immunosuppressive therapy, surgical correction of isthmic-cervical insufficiency, immunodeficiency conditions [25, 30, 96].

The development of the infectious process can manifest in congenital malformations (CM), premature delivery, growth and development retardation, intrauterine death, and clinical signs of infection immediately after birth and/or after several hours or days (if the infection occurred intranatally).

The inflammatory process can be localized in any organ or become systemic (generalized); in some situations, the penetration of an infectious agent into a macroorganism is not necessarily accompanied by clinical manifestations, which indicates an asymptomatic or subclinical course of infection [4, 6, 15–18, 30, 117, 118]. Transformation of an infection into a generalized process is determined by factors that reduce the body reactivity, namely the development of intrauterine and intranatal hypoxia, primary and secondary immunodeficiencies [27–29].

To prevent intrauterine infection, all triggering factors are taken into account. Their possible impact shall be excluded starting from the pregravid stage and throughout pregnancy and until childbirth.

CLASSIFICATION

There is no approved general classification of IPP. The ICD code X P39.8 should be used to encode infections if a newborn in the first week of life shows clinical and laboratory signs of an inflammatory reaction with the identification of a specified pathogen [viral, bacterial, parasitic, fungal, mixed (polymicrobial, viral-bacterial)] and/or with the presence of an infectious focus, the case

of which is not specified in the clinical recommendations and ICD X.

Code P37.9, P39.9 Infection, unspecified should be used to encode a case when a newborn in the first week of life shows clinical, laboratory and/or morphological signs of an inflammatory reaction without identification of the pathogen and localization of the inflammatory focus.

CLINICAL PATTERN

Early clinical symptoms of congenital/perinatal infections usually do not have specific manifestations. Immediately after birth, certain conditions may be detected in a newborn that indicate an unfavorable course of the intrauterine period (signs of intrauterine growth retardation), morphofunctional immaturity, congenital malformations, multiple dysmorphias, birth in a state of asphyxia against the background of chronic intrauterine hypoxia). From the first hours or in the first days of life (72 h), signs of deterioration in the condition increase as manifestations of infectious toxicosis: impaired thermal regulation [unstable temperature ($\geq 38.5^{\circ}\text{C}$ or $\leq 36.0^{\circ}\text{C}$), inability to retain heat independently], “marbling,” pale skin with a grayish shade, perioral cyanosis and/or acrocyanosis, sclerema, jaundice of unknown origin, early and prolonged yellowness of the mucous membranes and skin, polymorphic hemorrhagic rash (isolated, punctate petechiae, ecchymosis, confluent erythema, large hemorrhagic and necrotic foci, from birth or in the early stages, of different localization), other manifestations of hemorrhagic syndrome (gastric, pulmonary hemorrhage, macrohematuria, bleeding from skin puncture sites). A newborn may have a decreased or absent sucking reflex, refusal to eat, lethargy, muscle hypotonia, hyperesthesia, excitability or depression. Episodes of hypoglycemia or hyperglycemia, edema syndrome, respiratory disorders (apnea and/or tachypnea, increased need for oxygen, respiratory support), manifestations of cardiovascular failure [bradycardia (average heart rate (HR) less than 110 per minute) and/or tachycardia (average HR over 180 per minute)], other rhythm disturbances, arterial hypotension (mean arterial pressure less than the 5th percentile for the gestational age) are reported. Dysfunction of the gastrointestinal tract is possible (intolerance to enteral nutrition, abdominal distension, weakened or absent peristalsis during auscultation).

In the presence of a perinatal-specific manifest/severe infection, the following symptoms and syndromes may be present: sepsis-like syndrome (differential diagnosis is carried out with early neonatal sepsis), hepatosplenomegaly, cytopenia (usually monoleukopenia, thrombocytopenia), pneumonitis, hydrothorax, hepatitis, often cholestatic, pathological jaundice, enterocolitis/hemocolitis, and ascites.

Clinical manifestations of infection in intrauterine (antenatal) or intranatal infection in a newborn child most often occur in the first 72 hours of the child's life (early infections); in the group of children born with an extremely low body weight, manifestation may be delayed up to 5–7 days of life. In 85% of cases, symptoms of an infectious disease appear in the first 24 hours of life, often 6–8 hours after birth (very early infections), in 5% within 24–48 hours, in 10% signs of infection appear on the 2–3rd day of life. In premature babies, the first clinical signs appear from the moment of birth with the manifestation of respiratory disorders, which masks them as respiratory distress syndrome (RDS) of the newborn.

General clinical signs in the diagnosis of infectious and inflammatory process in newborns caused by perinatal infection have non-specific symptoms. It is important to conduct differential diagnosis with other infectious nosological entities, including, first of all, early neonatal sepsis and congenital pneumonia.

To confirm the diagnosis of PIS, the requirements are:

- thorough collection of the mother's perinatal history;
- physical examination of a newborn with identified one or more symptoms of the disease (see the Clinical Pattern section);
- laboratory and instrumental studies to exclude the source of infection [lungs, urinary system, gastrointestinal tract (GI), central nervous system (CNS)].

It is recommended to study the mother's medical history to identify the risk group for IPP development [5, 11, 30, 39, 46, 49, 52, 57, 58, 62–65, 68, 75–77, 97, 99, 109, 110, 119, 120]: the grade of recommendations is C (the level of evidence is 3).

Comments. Maternal risk factors for the development of IPP include:

- the presence of an acute infectious and inflammatory disease or exacerbation of a chronic infectious and inflammatory disease;
- invasive obstetric diagnostic or therapeutic procedures;
- the presence of clinical signs of acute and persistent, including bacterial, infections before or during childbirth;
- prolonged and frequent hospitalization of a mother during this pregnancy, multiple courses of antibacterial, hormonal and/or cytotoxic therapy;
- laboratory data of a mother before delivery: elevated C-reactive protein (CRP) level, leukocytosis (excluding leukocytosis after recent administration of corticosteroids);
- detection of pathogenic microorganisms in the mother's birth canal, primarily group B streptococcus or its antigens;
- pre-delivery discharge of amniotic fluid (anhydrous interval ≥ 18 hours);

- increase in maternal body temperature during delivery $\geq 38^\circ\text{C}$ for more than 2 h;
- intrauterine interventions during pregnancy;
- antibiotic therapy of a mother immediately before delivery or during delivery with protected penicillins or reserve antibacterial drugs;
- clinical manifestations of chorioamnionitis or other intra-amniotic infection;
- consumption of raw meat, raw eggs, raw milk, or contaminated vegetables and fruits, or contact with cat feces during pregnancy (*Toxoplasma gondii*) by a mother;
- vaginal delivery in the presence of primary maternal infection induced by herpes simplex virus types 1 or 2 (*Herpes simplex virus types 1, 2*);
- seronegative mothers who develop primary infection during pregnancy (cytomegalovirus) or exacerbation of cytomegalovirus infection in seropositive pregnant women (see Clinical Guidelines on Congenital Cytomegalovirus Infection¹);
- human immunodeficiency virus infection in a mother;
- consumption of dairy products and food without proper thermal treatment (*Listeria monocytogenes*);
- bacteriuria during pregnancy.

The main symptoms of chorioamnionitis in any combination include febrile fever (body temperature 38.0°C), maternal tachycardia (100 beats/min), fetal tachycardia (160 beats/min), purulent or purulent-bloody vaginal discharge, sometimes with a foul odor [65, 67]. In terms of diagnosis, histological examination of the placenta is of great importance with the detection of typical inflammatory changes in the vessels of the fetal part of the placenta and the wall of the umbilical cord (deciduitis, funisitis, vasculitis, placental tissue infiltration), which suggests the possible development of an infectious process in a newborn and serves as an additional criterion in verifying the diagnosis of a viral infection or IPP (placental examination is mandatory to confirm the diagnosis) [9, 28, 29, 43, 51, 56]. Signs of chorioamnionitis: the presence of inflammation of the fetal membranes, amniotic fluid and decidual tissue.

Additional risk factors for the development of IPP include: preeclampsia and other pregnancy complications in a mother; vitamin D deficiency; premature rupture of membranes; meconium-stained amniotic fluid with a specific odor; premature birth; fetal distress; perinatal hypoxia and asphyxia during childbirth; death of children in the family from severe bacterial infections at the age of up to 3 months (suspected primary immunodeficiency).

When collecting the mother's medical history, it is important to confirm the infection by microbiological (cultural) examination of the discharge from the female genital organs (and/or amniotic fluid) for aerobic and

¹ https://cr.minzdrav.gov.ru/recomend/260_2

facultative anaerobic microorganisms, and to conduct reasonable antibiotic therapy in order to reduce neonatal morbidity and purulent-septic complications in the mother.

It is also necessary to study mother's vaccination information, epidemiological history, occupational hazards (work in children's groups, work with animals, etc.), traveling, especially during pregnancy (typical for infections caused by the Epstein-Barr virus, malaria, dengue fever, Zika virus, etc.), the presence of diseases with exanthema during pregnancy; features of the current pregnancy: thrombocytopenia of unspecified etiology, threat and premature birth, congenital malformations, fetal growth retardation, fetal hydrops, previous missed miscarriages and antenatal death, placental insufficiency, polyhydramnios and oligohydramnios.

Neonatal factors include prematurity and intrauterine growth retardation, especially of the dysplastic type, multiple dysmorphias, congenital malformations and structural anomalies.

LABORATORY AND INSTRUMENTAL DIAGNOSTIC STUDIES

For a newborn with a suspected IPP, it is recommended to conduct a general (clinical) blood test, detailed with leukocyte and platelet counts, neutrophil index (NI), absolute neutrophil count, to detect any inflammatory changes with a repeat test at the age of 48–72 hours and at the end of the antibiotic therapy to decide on its cancellation or continuation (change) [11, 12, 14, 71, 82, 87, 94, 98, 101, 106, 115]: the grade of recommendations is B (the level of evidence is 3).

Comments. An elevated NI level and a low absolute neutrophil count are predictors of infection in newborns. The sensitivity of the absolute neutrophil count is 78%, the specificity is 73%, the sensitivity of NI is 78%, the specificity is 75%.

For a newborn with a suspected IPP, it is recommended to conduct a microbiological (culture) test of blood for sterility from the umbilical cord or peripheral vein and determine the sensitivity of microorganisms to antimicrobial chemotherapeutic drugs in order to detect and identify the pathogen, exclude sepsis in a newborn and determine the strategy of the antibiotic therapy [21, 23, 42, 45, 66, 82, 89, 91, 101, 106, 116, 120, 128]: the grade of recommendations is B; the level of evidence is 3.

Comments. A quick culture method (QCM, Shell vial assay) is preferred, if a medical institution (HF) has the possibilities. Umbilical blood culture has high sensitivity and specificity for diagnosing intrauterine infection of bacterial etiology. Modern microbiological studies can distinguish true bacteremia from contamination in blood culture (given the MI's possibilities). Blood culture is considered negative for gram-negative microorganisms

if there is no growth within 48 hours, and for gram-positive microorganisms if there is no colony growth within 72 hours. Upon that, blood culture is highly likely to be contaminated if colony growth occurs after 72 hours of incubation. Modern systems can identify a pathogen in 77, 89, 94% of cases 24, 36 and 48 hours after blood sampling, respectively. The absence of a positive result of blood culture of a pathogen DOES NOT exclude the presence of an infectious process in a newborn (given the MI's possibilities)

In case of tracheal intubation for a newborn with a suspected course of IPP, it is recommended to conduct a microbiological (cultural) study of sputum for aerobic and facultative-anaerobic microorganisms, sputum for fungi (yeast and mycelial) and determine the sensitivity of microorganisms to antimicrobial chemotherapeutic drugs to identify the pathogen, exclude pneumonia and determine the strategy of antimicrobial therapy [22, 44, 59, 91, 101, 106, 116, 120, 128]: the grade of recommendations is A (the level of evidence is 3).

Comments. A quick culture method (QCM, Shell vial assay) is preferred, given the MI's possibilities.

In the presence of risk factors for the development of IPP or clinical and/or laboratory signs of infection in a mother, for a newborn with a suspected IPP it is recommended to perform selective identification of DNA of *Epstein-Barr virus*, *Cytomegalovirus*, *Parvovirus B19*, herpes virus type 6 (*HHV6*), toxoplasma (*Toxoplasma gondii*), RNA of *Rubella virus* by polymerase chain reaction (PCR) in peripheral and umbilical cord blood, quantitative examination, determine DNA of *Treponema pallidum*, chlamydia (*Chlamydia* spp.), streptococci (*Streptococcus agalactiae*, *SGB*, *Streptococcus pyogenes*, *SGA*), *Haemophilus influenzae*, *Varicella-Zoster virus* and lichen in blood by PCR, quantitative study of herpes simplex types 1 and 2 (*Herpes simplex virus types 1, 2*), listeria (*Listeria monocytogenes*), *Pseudomonas aeruginosa*, by PCR in blood, quantitative examination, molecular biological study of urine for ureaplasma (*Ureaplasma* spp.) with species specification, molecular biological study of upper respiratory tract secretions for *Mycoplasma hominis*, molecular biological study of bronchoalveolar lavage fluid, sputum, endotracheal aspirate [if a child is on artificial ventilation (ALM)] for methicillin-sensitive and methicillin-resistant *Staphylococcus aureus*, methicillin-resistant coagulase-negative *Staphylococcus* spp., molecular biological examination of sputum, bronchoalveolar lavage fluid (if a child is on mechanical ventilation) for *Cytomegalovirus* to identify pathogens of IPP [4, 10, 18, 19, 20, 33, 66, 101]: the grade of recommendations is B (the level of evidence is 2).

Comments. Conducting a blood test by the PCR method depends on the technical capabilities of the MI's

laboratory. If there is no possibility to conduct a quantitative study, a qualitative study is acceptable. Upon receipt of the results of molecular genetic tests confirming the origin of the infectious process, further strategy of managing a child is implemented in accordance with the existing clinical guidelines for specific nosologies.

For a newborn with a suspected course of IPP, it is recommended to conduct study of the CRP level in blood serum with a control after 48–72 hours to identify signs of systemic inflammatory response syndrome (SIRS) and at the end of the course of antimicrobial therapy to determine the strategy of ABT (cancellation or continuation of therapy) [47, 48, 60, 67, 70, 86, 92, 103, 111, 134]: the grade of recommendations is B (the level of evidence is 2).

Comments. Reference values are determined by the method and type of analyzer used in the MI. CRP production starts 4–6 hours after development of the infectious process, it doubles after 8 hours and reaches the maximum peak after 36–48 hours. The assessment of CRP in the first 6–8 hours after birth has low sensitivity of 35–50% and false positive results in 30% [69, 70, 107, 112, 134]. In this regard, the study of the CRP level should be carried out 24–48 hours after birth (earlier if indicated), and monitoring shall be performed no earlier than 24–36 hours over time, which increases the sensitivity of this study to 74–98%, specificity to 71–94% [18, 47, 70, 92, 103, 112]. Non-infectious conditions of a newborn may influence the increase in the CRP level in the first 24–48 hours after birth: trauma, meconium aspiration syndrome, ischemic tissue damage and hemolysis [47, 60, 92, 103, 131]. An increase in the CRP level is an early sign of bacterial infection in full-term infants; in premature infants, such dependence has not been clearly proven (the sensitivity is 68.5%, the specificity is 85.5%) [47].

Determination and assessment of other SIRS markers to exclude a generalized infectious process (neonatal sepsis) is carried out according to indications (given the MI's possibilities):

- determination of the presepsin level in blood (sensitivity 80–94%, specificity 75–100%). It is known that the presepsin level does not depend on: gestational age, body weight, early postnatal age, method of delivery. Studies of the level of presepsin and interleukin6 (IL6) are not used to determine further strategy of antimicrobial therapy (cancellation, change, prolongation of the course), but only for early diagnosis of an infectious disease [3, 40, 79, 84, 85, 100, 104, 105];
- determination of IL6 levels in blood (sensitivity 83–95%, confidence interval 71–90%, specificity 87–95%, confidence interval 78–93%) [105, 124];

- determination of procalcitonin (PCT) levels in blood is assessed in accordance with the threshold value depending on the age (hours) after birth (sensitivity 87–94%, specificity 74–90%). In newborns, a physiological increase in PCT is observed during the first 48 hours of life, a sign of PCT infection of more than 2.5 ng/mL in the first 72 hours, after 72 hours — more than 2.0 ng/mL [50]. The PCT level increases in the first days after injury, surgery, severe burns, in patients with invasive fungal infections [79, 80].

Determination and correct assessment of inflammation markers, use of a combination of the above-specified markers in the diagnostic process increases the probability of identification of an infectious disease in a newborn [38, 40, 47, 48, 60, 67, 70, 79, 84, 85, 87, 92, 100, 103, 104, 105, 111, 124, 128, 127, 134].

For a newborn with a suspected IPP and the presence of neurological disorders typical for an infectious lesion of the central nervous system, it is recommended to perform a lumbar puncture and microscopic examination of the cerebrospinal fluid, cell counting in a counting chamber (determination of cytosol, protein level) to exclude meningitis/encephalitis [97]: the grade of recommendation is B (the level of evidence is 2).

Comments. Prior to performing a spinal puncture, it is necessary to stabilize the condition of a newborn (respiratory therapy, treatment of shock, seizures, hemorrhagic syndrome).

For a newborn with a suspected IPP and the presence of neurological disorders typical for an infectious lesion of the central nervous system, it is recommended to conduct a microbiological (cultural) study of the cerebrospinal fluid for aerobic and facultative-anaerobic opportunistic pathogens, determine DNA of the herpes simplex virus types 1 and 2 (*Herpes simplex virus types 1, 2*) in the cerebrospinal fluid by PCR [97]: the grade of recommendation is B (the level of evidence is 2).

Comments. Conducting a blood test of the cerebrospinal fluid by the PCR method depends on the technical capabilities of the MI's laboratory. If there is no possibility to conduct a quantitative study, a qualitative study is acceptable. Upon receipt of the results of molecular genetic tests confirming the origin of the infectious process, further strategy of managing a child is implemented in accordance with the existing clinical guidelines for specific nosologies.

For a newborn with a suspected IPP, a general (clinical) urine analysis is recommended to exclude urinary tract infection [97]: the grade of recommendation is B (the evidence level is 2).

For a newborn with a suspected IPP, it is recommended to conduct a microbiological (cultural) urine test for sterility [97] to exclude a urinary tract infection in the presence of pathological changes in the general (clinical) urine analysis: the grade of recommendations is B (the level of evidence is 2).

For a newborn with a suspected IPP and respiratory disorders, a chest X-ray is recommended to exclude pneumonia [18–22, 33, 34, 106]: the grade of recommendations is B (the level of evidence is 3).

Comments. It is necessary to determine the cause of respiratory disorders in a newborn and establish the appropriate diagnosis: congenital pneumonia, respiratory distress syndrome of the newborn, meconium aspiration syndrome, congenital malformations of the heart, congenital malformations of the lungs, interstitial lung diseases, etc.

For a newborn with a suspected IPP, is recommended to perform echocardiography (EchoCG), neurosonography (NSG), ultrasound examination (US) of the abdominal cavity, kidneys and adrenal glands to assess the function of organs and systems [18–21, 33, 34, 108]: the grade of recommendations is B (the level of evidence is 3).

Comments. In case of concomitant disorders of various organs and systems, EchoCG, NSG, ultrasound, ECG can facilitate timely prescription and correction of the symptomatic therapy. It is necessary to exclude various somatic diseases, primarily congenital malformations of the heart, lungs, intestines, kidneys.

For newborns with dysfunction of organs and systems, in order to control vital functions and differential diagnosis, it is recommended to conduct daily bedside monitoring of the heart rate, respiratory rate, blood pressure (including systolic), SpO₂, body temperature, diuresis rate [18–21, 33, 34, 55, 118, 133]: the grade of recommendations is B (the level of evidence is 3).

Comments. In case of concomitant disorders of the function of various organs and systems, the above studies can facilitate timely prescription and correction of the syndrome-based and symptomatic therapy.

In the early neonatal period, it is necessary to differentiate IPP from the following conditions and nosological entities with confirmed etiology: intrauterine infections (congenital cytomegalovirus infection, herpes simplex virus types 1, 2, 6, toxoplasmosis); neonatal sepsis; RDS of newborns; congenital pneumonia; meningitis; carditis; necrotizing enterocolitis; congenital malformations of the heart, lungs, intestines, kidneys; diaphragmatic hernia, hereditary metabolic diseases; congenital metabolic disorders; asphyxia; transient tachypnea of the newborn; neonatal meconium aspiration; persistent pulmonary hypertension of the newborn.

Treatment of children with IPP includes conservative therapy:

1. Etiotropic empirical ABT is prescribed to newborns with clinical and/or laboratory and instrumental signs of probable or proven IPP, but without specified etiology.

2. Justified intensive (syndrome-based) therapy is carried out for indications: correction of metabolic, hemostatic disorders, manifestations of organ dysfunction.

3. Symptomatic therapy.

4. Reasonable feeding (total parenteral nutrition, partial parenteral nutrition, breastfeeding, feeding with breast milk substitutes, including adapted formulas).

For newborns with clinical and anamnestic risk factors, 1–2 or more clinical symptoms and/or laboratory and instrumental signs of probable or proven congenital (perinatal) infection, but without specified etiology, it is recommended to prescribe empirical ABT at early stages [1, 2, 7, 8, 26, 31, 35–37, 46, 53, 61, 66, 72, 78, 80, 90, 93, 99, 102, 121, 126, 130, 132]: the grade of recommendations is A (the level of evidence is 1).

Comments. Antibacterial therapy (ABT) in case of suspected development of IPP is indicated at early stages after birth for the following categories of children: patients with very low birth weight (VLBW) and extremely low birth weight (ELBW); newborns requiring invasive ALV since birth due to the severity of condition; newborns with neonatal seizures. It is recommended to start ABT no later than 2 hours of life, for newborns with ELBW — in the delivery room. For newborns weighing more than 1500 g at birth, ABT is prescribed for indications based on the results of the initial clinical and laboratory examination. ABT started if development of a perinatal-specific infection is suspected in the first day of life is cancelled in the absence of clinical, laboratory and instrumental data confirming the infection within 48–72 hours of life (after determining the CRP level, for PCT indication). If a diagnosis of a perinatal-specific infection is confirmed, the empirical ABT regimen is continued until the results of the microbiological study and the evaluation of the results of the clinical, laboratory and instrumental examination are obtained, with a subsequent decision on the cancellation or further prescription of targeted ABT (in accordance with the sensitivity of the isolated microflora). When the levels of SIRS markers and the results of the clinical, laboratory and instrumental examination of the newborn are normalized, ABT is canceled.

Initial ABT regimens:

Regimen A: provides for ABT for newborns whose mothers have an uncomplicated medical history. It is recommended to prescribe empirical ABT using broad-spectrum penicillins (ATC code J01CA; ampicillin) [126, 129] in combination with other aminoglycosides (ATC code J01GB; gentamicin, amikacin, netilmicin) or monotherapy with a combination of penicillins,

including combinations with beta-lactamase inhibitors (ATC code J01CR; ampicillin + sulbactam). In case of renal dysfunction, it is reasonable to decide on the cancellation of other aminoglycosides (ATC code J01CA) on an individual basis, taking into account the available medical history data and results of laboratory and microbiological examination of a patient;

Regimen B: involves ABT in newborns whose mothers have a history of aggravating factors: chorioamnionitis, intrauterine interventions, prolonged anhydrous interval (more than 18 hours), elevated CRP, fever during labor lasting more than 2 hours, antibiotic therapy in a mother immediately before labor and during labor, culturing of group B streptococcus from the cervical canal. Regimen B can also be considered in cases where there are risk factors for infection on the part of a newborn (for example, VLBW, ELBW, artificial ventilation). In this case, it is reasonable to prescribe a combination of penicillins, including combinations with beta-lactamase inhibitors (ATC code J01CR; ampicillin + sulbactam) and other aminoglycosides (ATC code J01GB; gentamicin, amikacin, netilmicin);

Preference is given to parenteral administration of systemic antimicrobial drugs (ATC code J; intravenous administration of drugs). It is not recommended to prescribe drugs containing amoxicillin + clavulanic acid (ATC code J01CR) due to a possible adverse effect of clavulanic acid on the intestinal wall, especially in premature infants. It is unreasonable to include cephalosporins of the 1st (ATC code J01DB), 2nd (ATC code J01DC), 3rd (ATC code J01DD) and 4th (ATC code J01DE) generations in the initial antibacterial therapy regimen due to the lack of activity against *Listeria monocitogenes* and *Enterococcus* spp., as well as the risk of development of necrotizing enterocolitis and invasive candidiasis in newborns with ELBW;

Regimen C: targeted ABT. Targeted ABT is used if a mother has flora that is resistant to the drugs of the initial antibiotic therapy regimens "A" and "B" and/or after receiving the results of a microbiological examination of a newborn with determined sensitivity of microorganisms to systemic antimicrobial drugs (ATC code J).

In case of absence of sensitivity of the isolated pathogens to systemic antibacterial drugs (ATC code J01) of the initial regimen, it is necessary to change to systemic antibacterial drugs (ATC code J01), the sensitivity to which was revealed, or switch to local protocols taking into account the microbiological monitoring of the department where the patient stays.

In case of an increase in laboratory activity, as well as in case of a suspected nosocomial infection secondary to the conducted starting therapy, it is recommended to study the patient's biological material from all available loci, a microbiological (culture) blood test for sterility is mandatory, after which it is necessary to correct the ABT.

The duration and strategy of ABT are determined in each case individually and depend on the severity of the child's condition and the normalization of clinical, laboratory and instrumental data. When prescribing ABT, instructions for drug use shall be followed. If it is necessary to prescribe an antibacterial drug in accordance with the sensitivity of the isolated microflora for vital indications outside the instructions for human use (off-label), it is recommended to conduct a medical panel/consultation and obtain an informed consent from the patient's legal representative.

Pathogenetically justified intensive therapy for a newborn with IPP is carried out in accordance with clinical/methodological recommendations depending on the existing comorbid disease/condition (normalization of acid-base composition, correction of respiratory and hemodynamic disorders, etc.) [18, 20, 33, 34, 88, 113].

Symptomatic therapy includes the administration of drugs depending on the clinical manifestations of the infectious process (hemostatic, anticonvulsant, sedative, etc.).

Early start of enteral nutrition is recommended (preference is given to breast milk); according to indications, total parenteral nutrition, partial parenteral nutrition, feeding with breast milk substitutes, including adapted milk formulas is performed [52, 73, 123, 131].

The therapeutic and protective regimen involves creating optimal conditions for newborns nursing. Depending on the severity of a condition, a newborn with a suspected IPP should be transferred to the neonatal intensive care unit (NICU), intensive care unit (ICU) or neonatal pathology unit. A premature baby should be kept in a thermally neutral environment, sensory stimulation should be limited (protection from light, noise, touch), body temperature should be monitored depending on thermal regulation, and pain syndrome should be prevented.

Medical rehabilitation is carried out depending on the concomitant pathology and complications by specialized specialists (neurologist, ophthalmologist, etc.).

Prevention of perinatal-specific infection: timely detection and treatment of infectious diseases in a mother during pregnancy, vaccination of mothers [11, 12, 50, 54, 64, 125, 135].

Compliance with the rules and regulations of SanPiN 1.2.3685–21 (sanitary and epidemiological regime in obstetric and neonatal departments).

Treatment of newborns with a perinatal-specific infection is carried out in the 24-hour in-patient settings (NICU, ICU, department of pathology of newborns and premature babies, neonatal unit in children's hospitals).

Indications for patient discharge from the medical institution:

- 1) stabilization of the child's condition is achieved, no signs of organ and system failure;
- 2) normalization of inflammation markers;
- 3) cancellation of antibacterial therapy at least 24 hours before discharge with monitoring of the general blood test and its mandatory interpretation on the day before discharge;
- 4) absence of other contraindications to discharge.

The disease prognosis may vary, depending on the gestational age at the time of birth, the severity of the infectious process, its duration, and comorbidities. In extremely premature infants, immunocompromised patients who had a perinatal-specific infection, the risk of developing periventricular leukomalacia, retinopathy of prematurity, bronchopulmonary dysplasia, disability, or death increases.

ADDITIONAL INFO

Authors' contribution. All authors made a substantial contribution to the conception of the study,

acquisition, analysis, interpretation of data for the work, drafting and revising the article, final approval of the version to be published and agree to be accountable for all aspects of the study.

Funding source. This study was not supported by any external sources of funding.

Competing interests. The authors declare that they have no competing interests.

ДОПОЛНИТЕЛЬНАЯ ИНФОРМАЦИЯ

Вклад авторов. Все авторы внесли существенный вклад в разработку концепции, проведение исследования и подготовку статьи, прочли и одобрили финальную версию перед публикацией.

Источник финансирования. Авторы заявляют об отсутствии внешнего финансирования при проведении исследования.

Конфликт интересов. Авторы декларируют отсутствие явных и потенциальных конфликтов интересов, связанных с публикацией настоящей статьи.

REFERENCES

1. Ivanov DO, Petrenko YuV, Yakovleva EE, et al. *Algorithms of antibiotic prophylaxis and antibacterial therapy in structural units of a multidisciplinary children's hospital: a textbook*. St. Petersburg: SPbSPMU, 2024. 192 p. (In Russ.)
2. Alexandrovich YuS, Ivanov DO, Pshenishnov KV. *Sepsis of newborns*. Saint Petersburg: SPbSPMU, 2018. 174 p. (In Russ.)
3. Velkov VV. Use of presepsin biomarker for early and highly specific diagnostics of sepsis. *Wounds and wound infections*. *The prof. B.M. Kostyuchenok journal*. 2015;2(1):53–82. EDN: TUGSBZ
4. Zlokazov MD, Lyubimova AV, Tekhova IG, et al. The problems of detection and registration of infections specific to the perinatal period in neonates. *Epidemiology and Vaccine Prophylaxis*. 2018;17(5): 71–77. EDN: YQXVCP doi: 10.31631/2073-3046-2018-17-5-71-77
5. Zlokazov MD. *Epidemiological features of the occurrence of bacterial infections specific to the perinatal period in newborn children* [Dissertation]. St. Petersburg, 2020. 131 p. EDN: EJIEEN Available from: <https://www.dissercat.com/content/epidemiologicheskie-osobennosti-vozniknoveniya-bakterialnykh-infektsii-spetsifichnykh-dlya>
6. Ivanov DO, Petrenko YuV, Kurzina EA, Petrova NA. performance characteristics of clinical analysis of blood in infants who developed neonatal sepsis. *Bulletin of the V.A. Almazov Federal Centre of Heart, Blood and Endocrinology*. 2012;(3):41–52. EDN: PCCWWL
7. Ivanov DO, Shabalov NP, Petrenko YuV. Neonatal sepsis. Neonatal sepsis. Experience of the hypothesis. *Children's Medicine of the North-West*. 2012;3(3):37–45. EDN: PWVKID
8. Ivzhits MA, Zyryanov SK, Ushkalova EA, et al. The use of antimicrobial drugs in preterm newborn: the experience of creating the formulary. *Good Clinical Practice*. 2016;(3):56–65. EDN: XHYKZZ
9. Antonov AG, Amirkhanova DY, Astasheva IB, et al. *Selected clinical recommendations on neonatology*. Ed. by Baibarina EN, Degtyarev DN. Moscow: GEOTAR-Media; 2016. 240 p. (In Russ.)
10. Ionov OV, Sharafutdinova DR, Balashova EN, et al. Necrotizing enterocolitis in extremely low birth weight infants and associated risk factors: a retrospective analysis. *Neonatology: News, Views, Education*. 2023;11(1):29–41. EDN: EHHGDS doi: 10.33029/2308-2402-2023-11-1-28-41
11. Ionov OV, Krokshina KN, Gorbacheva LM, et al. Leukocytosis: a new important diagnostic marker for inflammatory infection in premature neonates older than 72 hours of age. *Neonatology: News, Views, Education*. 2016;(1):81–88.
12. Ionov OV, Degtyareva AV, Levadnaya AV, et al. Clinical and laboratory manifestations of congenital infectious and inflammatory diseases in extremely low and very low birth weight infants. *Obstetrics and Gynecology*. 2014;(10):66–71. EDN: SXRJXT
13. Knyazeva TP. Causes and risk factors of premature rupture membranes. *Far Eastern Medical Journal*. 2016;(2):128–135. (In Russ.) EDN: WCAXRZ
14. Kozlov RS, Dehnic AV, editors. *Handbook of antimicrobial therapy*. Issue 2. MAKMAX; 2010. 416 c. (In Russ.)
15. Kozlov SN, Strachunsky LS. *Modern antimicrobial chemotherapy: a guide for doctors*. Moscow: Med Inform Agency; 2009. 448 p. (In Russ.) EDN: QLUROV
16. Kosenkova EG, Lysenko IM. *Clinical and diagnostic criteria for the implementation of intrauterine infection in newborns and children of the first year of life: a monograph*. Vitebsk: VSMU; 2016. P. 184–200.

17. Lobzin YuV, Vasiliev VV. Key aspects congenital infection. *Journal of Infectology*. 2014;6(3):5–14. EDN: STQTXJ
18. Volodin NN, Degtyarev DN, editors. *Neonatology: a national guideline*. Vol. 2. Moscow: GEOTAR-Media; 2023. 768 p.
19. Ovsyannikov DYU, Boytsova EV, Zhestkova MA, et al. *Neonatal pulmonology: Monograph*. Ed. by Ovsyannikov DYU. Moscow: Seven-Print; 2022. 168 p. EDN: NGFFJV
20. Ovsyannikov DYU, Kozlov VV, Stuklov NI, et al. *Pediatrics: textbook*. Volume 2: *Otorhinolaryngology, pulmonology, haematology, immunology*. Ed. by Ovsyannikov DYU. Moscow: RUDN, 2022. 592 p. EDN: EWEMYE
21. Ovsyannikov DYU, Boitsova EV. Pneumonia in newborns. *Pediatrics. Consilium Medicum*. 2021;3(3):214–223. EDN: ENPMBP doi: 10.26442/26586630.2021.3.201060
22. Ovsyannikov DYU, Kravchuk DA, Nikolaeva DYU. Clinical pathophysiology of the respiratory system in preterm infants. *Neonatology: News, Views, Education*. 2018;6(3(21)):74–98. EDN: MHCYTZ doi: 10.24411/2308-2408-2402-2018-13003
23. Bryant KA, Kuzman-Cottrill DA. *Manual on the prevention of infectious diseases in pediatrics*. Ed. by Osmanov IM, Borzakova SN. Moscow: GEOTAR-Media; 2021. 333 p. (In Russ.)
24. Savicheva AM. Vaginal microbiome and virome — features of interaction. *Problems of Medical Mycology*. 2023;25(2):172. EDN: NSIDPM
25. Savicheva AM, Sokolovsky EV, Ailamazyan EK. *Infectious-inflammatory diseases in obstetrics and gynaecology*. Manual for doctors. Ailamazyan EK, editor. Moscow GEOTAR-Media; 2016. 320 p. (In Russ.) EDN: WJAQZH
26. Samsygina GA. *Neonatal sepsis*. Moscow: GEOTAR-Media; 2020. 192 p. (In Russ.)
27. Samsygina GA. On predisposing factors and risk factors for the development of neonatal sepsis and modern approaches to its treatment. *Pediatrics. Journal named after G.N. Speransky*. 2012;91(3):32–37. (In Russ.) EDN: OZMAVH
28. Zinserling VA. The importance of morphological investigations in diagnostics and study of infections. *Tissue microbiology. Journal Infectology*. 2018;10(3):124–132. EDN: VMVIXY doi: 10.22625/2072-6732-2018-10-3-124-132
29. Zinserling VA, Melnikova VF. *Perinatal infections. (Questions of pathogenesis, morphological diagnosis and clinical and morphological comparisons)*. Practical Guide. Saint Petersburg: Elby; 2002. 352 p. (In Russ.)
30. Chernyakhovsky OB, Abramova IV. Intrauterine infections in the newborn, risk factors. *Russian Bulletin of Perinatology and Pediatrics*. 2009;54(1):88. EDN: KPVMXZ
31. Shabalov NP, Ivanov DO, Shabalova NN. Sepsis of newborns. *Medical Academic Journal*. 2001;1(3):81–86. (In Russ.) EDN: WDQJYF
32. Shabalov NP. Obschebiological problem: patterns and consequences of perinatal human infection. *Pediatrics. Journal named after G.N. Speransky*. 2012;91(3):26–31. EDN: OZMAUX
33. Shabalov NP, Sofronova LN. *Neonatology: textbook*. Vol. 2. Moscow: GEOTAR-Media; 2020. 752 p. (In Russ.) EDN: ANAYGR doi: 10.33029/9704-5771-9-NEO-2020-1-752
34. Shabalov NP, Sofronova LN. *Neonatology: textbook*. Vol. 1. Moscow: GEOTAR-Media; 2020. 720 p. (In Russ.) EDN: WUSRND doi: 10.33029/9704-5770-2-NEO-2020-1-720
35. Al-Matary A, Al Sulaiman M, Al-Otaiby S, et al. Association between the timing of antibiotics administration and outcome of neonatal sepsis. *J Infect Public Health*. 2022;15(6):643–647. doi: 10.1016/j.jiph.2022.05.004
36. Al-Lawama M, Aljbouir H, Tanash A, Badran E. Intravenous Colistin in the treatment of multidrug-resistant *Acinetobacter* in neonates. *Ann Clin Microbiol Antimicrob*. 2016;15:8. doi: 10.1186/s12941-016-0126-4
37. EARSNet [Internet]. Annual report of the european antimicrobial resistance surveillance network stockholm: ECDC; 2017. [Cited 2024 Oct 08]. Available from: <https://www.ecdc.europa.eu/en/about-us/networks/disease-networks-and-laboratory-networks/ears-net-data>
38. Anugu NR, Khan S. Comparing the diagnostic accuracy of procalcitonin and C-reactive protein in neonatal sepsis: a systematic review. *Cureus*. 2021;13(11):e19485. doi: 10.7759/cureus.19485
39. Balamuth F, Weiss SL, Fitzgerald JC, et al. Protocolized treatment is associated with decreased organ dysfunction in pediatric severe sepsis. *Pediatr Crit Care Med*. 2016;17(9):817–822. doi: 10.1097/PCC.0000000000000858
40. Barichello T, Generoso JS, Singer M, Dal-Pizzol F. Biomarkers for sepsis: more than just fever and leukocytosis — a narrative review. *Crit Care*. 2022;26(1):14. doi: 10.1186/s13054-021-03862-5
41. Bell EF, Acarregui MJ. Restricted versus liberal water intake for preventing morbidity and mortality in preterm infants. *Cochrane Database Syst Rev*. 2014;2014(12):CD000503. doi: 10.1002/14651858.CD000503
42. Berardi A, Buffagni AM, Rossi C, et al. Serial physical examinations, a simple and reliable tool for managing neonates at risk for early-onset sepsis. *World J Clin Pediatr*. 2016;5(4):358–364. doi: 10.5409/wjcp.v5.i4.358
43. Beucher G, Charlier C, Cazanave C. Diagnosis and management of intra-uterine infection: CNGOF preterm premature rupture of membranes guidelines. *Gynecol Obstet Fertil Senol*. 2018;46(12):1054–1067. (In French). doi: 10.1016/j.gofs.2018.10.022
44. Bianchini S, Rigotti E, Nicoletti L, et al. Surgical antimicrobial prophylaxis in neonates and children with special high-risk conditions: A RAND/UCLA appropriateness method consensus study. *Antibiotics (Basel)*. 2022;11(2):246. doi: 10.3390/antibiotics11020246
45. Bedetti L, Miselli F, Minotti C, et al. Lumbar puncture and meningitis in infants with proven early- or late-onset sepsis: an Italian prospective multicenter observational study. *Microorganisms*. 2023;11(6):1546. doi: 10.3390/microorganisms11061546
46. Berardi A, Zinani I, Bedetti L, et al. Should we give antibiotics to neonates with mild non-progressive symptoms? A comparison of serial clinical observation and the neonatal sepsis risk calculator. *Front Pediatr*. 2022;10:882416. doi: 10.3389/fped.2022.882416
47. Cantey JB, Lee JH. Biomarkers for the diagnosis of neonatal sepsis. *Clin Perinatol*. 2021;48(2):215–227. doi: 10.1016/j.clp.2021.03.012
48. Çelik HT, Portakal O, Yiğit Ş, et al. Efficacy of new leukocyte parameters versus serum C-reactive protein, procalcitonin, and interleukin-6 in the diagnosis of neonatal sepsis. *Pediatr Int*. 2016;58(2):119–125. doi: 10.1111/ped.12754
49. Clements KE, Fisher M, Quaye K, et al. Surgical site infections in the NICU. *J Pediatr Surg*. 2016;51(9):1405–1408. doi: 10.1016/j.jpedsurg.2016.04.002
50. Collins A, Weitkamp JH, Wynn JL. Why are preterm newborns at increased risk of infection? *Arch Dis Child Fetal Neonatal Ed*. 2018;103(4):F391–F394. doi: 10.1136/archdischild-2017-313595

51. Conde-Agudelo A, Romero R, Jung EJ, Garcia Sánchez ÁJ. Management of clinical chorioamnionitis: an evidence-based approach. *Am J Obstet Gynecol.* 2020;223(6):848–869. doi: 10.1016/j.ajog.2020.09.044
52. Coyne R, Hughes W, Purtill H, et al. Influence of an early human milk diet on the duration of parenteral nutrition and incidence of late-onset sepsis in very low birthweight (VLBW) infants: a systematic review. *Breastfeed Med.* 2024;19(6):425–434. doi: 10.1089/bfm.2023.0290
53. Craig AM, Dotters-Katz S, Kuller JA, Thompson JL. Listeria in pregnancy: a review. *Obstet Gynecol Surv.* 2019;74(6):362–368. doi: 10.1097/OGX.0000000000000683
54. Czikk MJ, McCarthy FP, Murphy KE. Chorioamnionitis: from pathogenesis to treatment. *Clin Microbiol Infect.* 2011;17(9):1304–1311. doi: 10.1111/j.1469-0691.2011.03574.x
55. Davis AL, Carcillo JA, Aneja RK, et al. American college of critical care medicine clinical practice parameters for hemodynamic support of pediatric and neonatal septic shock. *Crit Care Med.* 2017;45(6):1061–1093. doi: 10.1097/CCM.00000000000002425
56. De Rose DU, Ronchetti MP, Tzialla C, et al. Editorial: Congenital and perinatal infections: How to prevent sequelae in neonates and children. *Front Pediatr.* 2023;11:1142636. doi: 10.3389/fped.2023.1142636
57. Dempsey E, Chen MF, Kokottis T, et al. Outcome of neonates less than 30 weeks gestation with histologic chorioamnionitis. *Am J Perinatol.* 2005;22(3):155–159. doi: 10.1055/s-2005-865020
58. Dior UP, Kogan L, Eventov-Friedman S, et al. Very high intrapartum fever in term pregnancies and adverse obstetric and neonatal outcomes. *Neonatology.* 2016;109(1):62–68. doi: 10.1159/000440938
59. Donà D, Barbieri E, Daverio M, et al. Implementation and impact of pediatric antimicrobial stewardship programs: a systematic scoping review. *Antimicrob Resist Infect Control.* 2020;9(1):3. doi: 10.1186/s13756-019-0659-3
60. Eschborn S, Weitkamp JH. Procalcitonin versus C-reactive protein: review of kinetics and performance for diagnosis of neonatal sepsis. *J Perinatol.* 2019;39(7):893–903. doi: 10.1038/s41372-019-0363-4
61. Evans IVR, Phillips GS, Alpern ER, et al. Association between the New York sepsis care mandate and in-hospital mortality for pediatric sepsis. *JAMA.* 2018;320(4):358–367. doi: 10.1001/jama.2018.9071
62. Freud A, Wainstock T, Sheiner E, et al. Maternal chorioamnionitis & long term neurological morbidity in the offspring. *Eur J Paediatr Neurol.* 2019;23(3):484–490. doi: 10.1016/j.ejpn.2019.03.005
63. Fuchs A, Bielicki J, Mathur S, et al. Reviewing the WHO guidelines for antibiotic use for sepsis in neonates and children. *Paediatr Int Child Health.* 2018;38(S1):S3–S15. doi: 10.1080/20469047.2017.1408738
64. García-Muñoz Rodrigo F, Galán Henríquez G, Figueras Aloy J, García-Alix Pérez A. Outcomes of very-low-birth-weight infants exposed to maternal clinical chorioamnionitis: a multicentre study. *Neonatology.* 2014;106(3):229–234. doi: 10.1159/000363127. Epub 2014 Jul 5. Erratum in: *Neonatology.* 2015;107(1):42. PMID: 25011418.
65. García-Muñoz Rodrigo F, Galán Henríquez GM, Ospina CG. Morbidity and mortality among very-low-birth-weight infants born to mothers with clinical chorioamnionitis. *Pediatr Neonatol.* 2014;55(5):381–386. doi: 10.1016/j.pedneo.2013.12.007
66. Giannoni E, Dimopoulou V, Klingenberg C, et al. Analysis of antibiotic exposure and early-onset neonatal sepsis in Europe, North America, and Australia. *JAMA Netw Open.* 2022;5(11):e2243691. doi: 10.1001/jamanetworkopen.2022.43691
67. Hedegaard SS, Wisborg K, Hvas AM. Diagnostic utility of biomarkers for neonatal sepsis — a systematic review. *Infect Dis (Lond).* 2015;47(3):117–124. doi: 10.3109/00365548.2014.971053
68. Higgins RD, Saade G, Polin RA, et al. Evaluation and management of women and newborns with a maternal diagnosis of chorioamnionitis: summary of a workshop. *Obstet Gynecol.* 2016;127(3):426–436. doi: 10.1097/AOG.0000000000001246
69. Hofer N, Müller W, Resch B. Neonates presenting with temperature symptoms: role in the diagnosis of early onset sepsis. *Pediatr Int.* 2012;54(4):486–490. doi: 10.1111/j.1442-200X.2012.03570.x
70. Hofer N, Zacharias E, Müller W, Resch B. An update on the use of C-reactive protein in early-onset neonatal sepsis: current insights and new tasks. *Neonatology.* 2012;102(1):25–36. doi: 10.1159/000336629
71. Hornik CP, Benjamin DK, Becker KC, et al. Use of the complete blood cell count in early-onset neonatal sepsis. *Pediatr Infect Dis J.* 2012;31(8):799–802. doi: 10.1097/INF.0b013e318256905c
72. Huttner A, Harbarth S, Carlet J, et al. Antimicrobial resistance: a global view from the 2013 World Healthcare-Associated Infections Forum. *Antimicrob Resist Infect Control.* 2013;2:31. doi: 10.1186/2047-2994-2-31
73. Imdad A, Rehman F, Davis E, et al. Effects of neonatal nutrition interventions on neonatal mortality and child health and development outcomes: A systematic review. *Campbell Syst Rev.* 2021;17(1):e1141. doi: 10.1002/cl2.1141
74. Gao K, Fu J, Guan X, et al. Incidence, bacterial profiles, and antimicrobial resistance of culture-proven neonatal sepsis in South China. *Infect Drug Resist.* 2019;12:3797–3805.
75. Joshi NS, Gupta A, Allan JM, et al. Clinical monitoring of well-appearing infants born to mothers with chorioamnionitis. *Pediatrics.* 2018;141(4):e20172056. doi: 10.1542/peds.2017-2056
76. Kim CJ, Romero R, Chaemsathong P, et al. Acute chorioamnionitis and funisitis: definition, pathologic features, and clinical significance. *Am J Obstet Gynecol.* 2015;213(S4):S29–S52. doi: 10.1016/j.ajog.2015.08.040
77. Kim MJ, Romero R, Gervasi MT, et al. Widespread microbial invasion of the chorioamniotic membranes is a consequence and not a cause of intra-amniotic infection. *Lab Invest.* 2009;89(8):924–936. doi: 10.1038/labinvest.2009.49
78. Kimpton JA, Verma A, Thakkar D, et al. Comparison of NICE Guideline CG149 and the sepsis risk calculator for the management of early-onset sepsis on the postnatal ward. *Neonatology.* 2021;118(5):562–568. doi: 10.1159/000518059
79. Kondo Y, Umemura Y, Hayashida K, et al. Diagnostic value of procalcitonin and presepsin for sepsis in critically ill adult patients: a systematic review and meta-analysis. *J Intensive Care.* 2019;7:22. doi: 10.1186/s40560-019-0374-4
80. Korang SK, Safi S, Nava C, et al. Antibiotic regimens for early-onset neonatal sepsis. *Cochrane Database Syst. Rev.* 2021;5(5):CD013837. doi: 10.1002/14651858.CD013837.pub2
81. Kuhn P, Dheu C, Bolender C, et al. Incidence and distribution of pathogens in early-onset neonatal sepsis in the era of antenatal antibiotics. *Paediatr Perinat Epidemiol.* 2010;24(5):479–487. doi: 10.1111/j.1365-3016.2010.01132.x

82. Kuzniewicz MW, Puopolo KM, Fischer A, et al. A quantitative, risk-based approach to the management of neonatal early-onset sepsis. *JAMA Pediatr.* 2017;171(4):365–371. doi: 10.1001/jamapediatrics.2016.4678
83. Kuzniewicz MW, Mukhopadhyay S, Li S, et al. Time to positivity of neonatal blood cultures for early-onset sepsis. *Pediatr Infect Dis J.* 2020;39(7):634–640. doi: 10.1097/INF.0000000000002632
84. Lu B, Zhang Y, Li C, et al. The utility of presepsin in diagnosis and risk stratification for the emergency patients with sepsis. *Am J Emerg Med.* 2018;36(8):1341–1345. doi: 10.1016/j.ajem.2017.12.038
85. Maddaloni C, De Rose DU, Santisi A, et al. The emerging role of presepsin (P-SEP) in the diagnosis of sepsis in the critically ill infant: a literature review. *Int J Mol Sci.* 2021;22(22):12154. doi: 10.3390/ijms222212154
86. Makkar M, Gupta C, Pathak R, et al. Performance evaluation of hematologic scoring system in early diagnosis of neonatal sepsis. *J Clin Neonatol.* 2013;2(1):25–29. doi: 10.4103/2249-4847.109243
87. Makkar N, Soneja M, Arora U, et al. Prognostic utility of biomarker levels and clinical severity scoring in sepsis: a comparative study. *J Investig Med.* 2022;70(6):1399–1405. doi: 10.1136/jim-2021-002276
88. Manurung TN, Wungu CDK, Utomo MT. The role of breast milk on reducing the risk of neonatal sepsis in preterm and low birth weight infants: a systematic review and meta-analysis. *Pharmacognosy Journal.* 2022;14(6):1067–1074. doi: 10.5530/pj.2022.14.211
89. Marks L, de Waal K, Ferguson JK. Time to positive blood culture in early onset neonatal sepsis: A retrospective clinical study and review of the literature. *J Paediatr Child Health.* 2020;56(9):1371–1375. doi: 10.1111/jpc.14934
90. Mateus T, Silva J, Maia RL, Teixeira P. Listeriosis during pregnancy: a public health concern. *ISRN Obstet Gynecol.* 2013;2013:851712. doi: 10.1155/2013/851712
91. Miller JM, Binnicker MJ, Campbell S, et al. guide to utilization of the microbiology laboratory for diagnosis of infectious diseases: 2024 update by the infectious diseases society of America (IDSA) and the American society for microbiology (ASM). *Clin Infect Dis.* 2024; ciae104. doi: 10.1093/cid/ciae104
92. Mjelle AB, Guthe HJT, Reigstad H, et al. Serum concentrations of C-reactive protein in healthy term-born Norwegian infants 48–72 hours after birth. *Acta Paediatr.* 2019;108(5):849–854. doi: 10.1111/apa.14578
93. Moffett SM, Kitts HL, Henderson SJ. Medication therapy for early-onset neonatal sepsis. *AACN Adv Crit Care.* 2016;27(3):253–258. doi: 10.4037/aacnacc2016503
94. Murphy K, Weiner J. Use of leukocyte counts in evaluation of early-onset neonatal sepsis. *Pediatr Infect Dis J.* 2012;31(1):16–19. doi: 10.1097/INF.0b013e31822ffc17
95. Lamont RF, Sobel J, Mazaki-Tovi S, et al. Listeriosis in human pregnancy: a systematic review. *J Perinat Med.* 2011;39(3):227–236. doi: 10.1515/jpm.2011.035
96. Young TE, Mangum OB. *Neofax: A manual of drugs used in neonatal care.* American Society of Hospital Pharmacists; 2000. 272 p.
97. Neonatal infection: antibiotics for prevention and treatment. NICE. 2021. 81 p. Available from: <http://nice.org.uk/guidance/ng195>
98. Newman TB, Draper D, Puopolo KM, et al. Combining immature and total neutrophil counts to predict early onset sepsis in term and late preterm newborns: use of the I/T2. *Pediatr Infect Dis J.* 2014;33(8):798–802. doi: 10.1097/INF.0000000000000297
99. Ofman G, Vasco N, Cantey JB. Risk of early-onset sepsis following preterm, prolonged rupture of membranes with or without chorioamnionitis. *Am J Perinatol.* 2016;33(4):339–342. doi: 10.1055/s-0035-1556758
100. Ozdemir AA, Elgormus Y. Diagnostic value of presepsin in detection of early-onset neonatal sepsis. *Am J Perinatol.* 2017;34(6):550–556. doi: 10.1055/s-0036-1593851
101. Pammi M, Flores A, Versalovic J, Leeflang MM. Molecular assays for the diagnosis of sepsis in neonates. *Cochrane Database Syst Rev.* 2017;2(2):CD011926. doi: 10.1002/14651858.CD011926.pub2
102. Paul R, Neuman MI, Monuteaux MC, Melendez E. Adherence to PALS sepsis guidelines and hospital length of stay. *Pediatrics.* 2012;130(2):e273–e280. doi: 10.1542/peds.2012-0094
103. Pierrakos C, Velissaris D, Bisdorff M, et al. Biomarkers of sepsis: time for a reappraisal. *Crit Care.* 2020;24(1):287. doi: 10.1186/s13054-020-02993-5
104. Poggi C, Bianconi T, Gozzini E, et al. Presepsin for the detection of late-onset sepsis in preterm newborns. *Pediatrics.* 2015;135(1):68–75. doi: 10.1542/peds.2014-1755
105. Poggi C, Lucenteforte E, Petri D, et al. Presepsin for the diagnosis of neonatal early-onset sepsis: a systematic review and meta-analysis. *JAMA Pediatr.* 2022;176(8):750–758. doi: 10.1001/jamapediatrics.2022.1647
106. Polin RA; Committee on fetus and newborn. Management of neonates with suspected or proven early-onset bacterial sepsis. *Pediatrics.* 2012;129(5):1006–1015. doi: 10.1542/peds.2012-0541
107. Pontrelli G, De Crescenzo F, Buzzetti R, et al. Accuracy of serum procalcitonin for the diagnosis of sepsis in neonates and children with systemic inflammatory syndrome: a meta-analysis. *BMC Infect Dis.* 2017;17(1):302. doi: 10.1186/s12879-017-2396-7
108. Pugaloni F, De Rose DU, Kipfmüller F, et al. Assessment of hemodynamic dysfunction in septic newborns by functional echocardiography: a systematic review. *Pediatr Res.* 2024;95(6):1422–1431. doi: 10.1038/s41390-024-03045-2
109. Puopolo KM, Draper D, Wi S, et al. Estimating the probability of neonatal early-onset infection on the basis of maternal risk factors. *Pediatrics.* 2011;128(5):e1155–e1163. doi: 10.1542/peds.2010-3464
110. Raines DA, Wagner A, Salinas A. Intraamniotic infection and the term neonate. *Neonatal Netw.* 2017;36(6):385–387. doi: 10.1891/0730-0832.36.6.385
111. Rees CA, Lim J, Westbrook AL, et al. Systematic review and meta-analysis of the diagnostic value of four biomarkers in detecting neonatal sepsis in low- and middle-income countries. *BMJ Paediatr Open.* 2023;7(1):e001627. doi: 10.1136/bmjpo-2022-001627
112. Ohis RK, Yoder MC. *Hematology, immunology and infection disease: neonatology questions and controversies.* Philadelphia: Elsevier Health Sciences, 2008. 312 p.
113. Rojas-Reyes MX, Morley CJ, Soll R. Prophylactic versus selective use of surfactant in preventing morbidity and mortality in preterm infants. *Cochrane Database Syst Rev.* 2012;(3):CD000510. doi: 10.1002/14651858
114. Romero R, Gomez-Lopez N, Winters AD, et al. Evidence that intra-amniotic infections are often the result of an ascending invasion — a molecular microbiological study. *J Perinat Med.* 2019;47(9):915–931. doi: 10.1515/jpm-2019-0297
115. Saboohi E, Saeed F, Khan RN, Khan MA. Immature to total neutrophil ratio as an early indicator of early neonatal sepsis. *Pak J Med Sci.* 2019;35(1):241–246. doi: 10.12669/pjms.35.1.99

116. Sarkar S, Bhagat I, DeCristofaro JD, et al. A study of the role of multiple site blood cultures in the evaluation of neonatal sepsis. *J Perinatol*. 2006;26(1):18–22. doi: 10.1038/sj.jp.7211410
117. Schlapbach LJ, Weiss SL, Wolf J. Reducing collateral damage from mandates for time to antibiotics in pediatric sepsis—primum non nocere. *JAMA Pediatr*. 2019;173(5):409–410. doi: 10.1001/jamapediatrics.2019.0174
118. Seyoum K, Sahiledengle B, Kene C, et al. Determinants of neonatal sepsis among neonates admitted to neonatal intensive care units in Ethiopian hospitals: A systematic review and meta-analysis. *Heliyon*. 2023;9(9):e20336. doi: 10.1016/j.heliyon.2023.e20336
119. Sgro M, Kobylanski A, Yudin MH, et al. Population-based study of early-onset neonatal sepsis in Canada. *Paediatr Child Heal*. 2019;24(2):e66–e73. doi: 10.1093/pch/pxy018
120. Shane AL, Sánchez PJ, Stoll BJ. Neonatal sepsis. *Lancet*. 2017;390(10104):1770–1780. doi: 10.1016/S0140-6736(17)31002-4.
121. Singer M, Deutschman CS, Seymour CW, et al. The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA*. 2016;315(8):801–810. doi: 10.1001/jama.2016.0287
122. Stoll BJ, Hansen NI, Sánchez PJ, et al. Early onset neonatal sepsis: The burden of group B streptococcal and *E. coli* disease continues. *Pediatrics*. 2011;127(5):817–826. doi: 10.1542/peds.2010-2217
123. Sturrock S, Sadoo S, Nanyunja C, Le Doare K. Improving the treatment of neonatal sepsis in resource-limited settings: gaps and recommendations. *Res Rep Trop Med*. 2023;14:121–134. doi: 10.2147/RRTM.S410785
124. Sun B, Liang LF, Li J, et al. A meta-analysis of interleukin-6 as a valid and accurate index in diagnosing early neonatal sepsis. *Int Wound J*. 2019;16(2):527–533. doi: 10.1111/iwj.13079
125. Tita AT, Andrews WW. Diagnosis and management of clinical chorioamnionitis. *Clin Perinatol*. 2010;37(2):339–354. doi: 10.1016/j.clp.2010.02.003
126. Tuzun F, Ozkan H, Cetinkaya M, et al. Is European Medicines Agency (EMA) sepsis criteria accurate for neonatal sepsis diagnosis or do we need new criteria? *PLoS One*. 2019;14(6):e0218002. doi: 10.1371/journal.pone.0218002
127. van Leeuwen LM, Fourie E, van den Brink G, et al. Diagnostic value of maternal, cord blood and neonatal biomarkers for early-onset sepsis: a systematic review and meta-analysis. *Clin Microbiol Infect*. 2024;30(7):850–857. doi: 10.1016/j.cmi.2024.03.005
128. Varghese A, Blaschke AJ, Korgenski EK, Crandall H. Neonatal early-onset sepsis due to haemophilus influenzae in utero. *Pediatr Infect Dis J*. 2023;42(3):e90–e92. doi: 10.1097/INF.0000000000003795
129. Vatne A, Klingenberg C, Rettedal S, Øymar K. Early-onset sepsis in neonates — a population-based study in South-West Norway from 1996 to 2018. *Front Pediatr*. 2021;9:634798. doi: 10.3389/fped.2021.634798
130. Walker S, Datta A, Massoumi RL, et al. Antibiotic stewardship in the newborn surgical patient: A quality improvement project in the neonatal intensive care unit. *Surgery*. 2017;162(6):1295–1303. doi: 10.1016/j.surg.2017.07.021
131. Wiechers C, Bernhard W, Goelz R, et al. Optimizing early neonatal nutrition and dietary pattern in premature infants. *Int J Environ Res Public Health*. 2021;18(14):7544. doi: 10.3390/ijerph18147544
132. Wing EJ, Gregory SH. *Listeria monocytogenes*: clinical and experimental update. *J Infect Dis*. 2002;185(Suppl 1):S18–S24. doi: 10.1086/338465
133. Yapicioğlu H, Özlü F, Sertdemir Y. Are vital signs indicative for bacteremia in newborns? *J Matern Fetal Neonatal Med*. 2015;28(18):2244–2249. doi: 10.3109/14767058.2014.983896
134. Yochpaz S, Friedman N, Zirkov S, et al. C-reactive protein in early-onset neonatal sepsis — a cutoff point for CRP value as a predictor of early-onset neonatal sepsis in term and late preterm infants early after birth? *J Matern Fetal Neonatal Med*. 2022;35(23):4552–4557. doi: 10.1080/14767058.2020.1856068
135. Zachariah P, Saiman L. Expanding antimicrobial stewardship strategies for the NICU: Management of surgical site infections, perioperative prophylaxis, and culture negative sepsis. *Semin Perinatol*. 2020;44(8):151327. doi: 10.1016/j.semperi.2020.151327

СПИСОК ЛИТЕРАТУРЫ

1. Иванов Д.О., Петренко Ю.В., Яковлева Е.Е. и др. Алгоритмы проведения антибиотикопрофилактики и антибактериальной терапии в структурных подразделениях многопрофильного детского стационара: учебное пособие. Санкт-Петербург: СПбГПМУ, 2024. 192 с.
2. Александрович Ю.С., Иванов Д.О., Пшениснов К.В. Сепсис новорожденных. Санкт-Петербург: СПбГПМУ, 2018, 174 с.
3. Вельков В.В. Использование биомаркера пресепсина для ранней и высокоспецифичной диагностики сепсиса. Раны и раневые инфекции // Журнал имени проф. Б.М. Костюченка. 2015. Т. 2, № 1. С. 53–82. EDN: TUGSBZ
4. Злоказов М.Д., Любимова А.В., Техова И.Г., и др. Проблемы выявления и учета инфекций, специфичных для перинатального периода у новорожденных детей // Эпидемиология и вакцинопрофилактика. 2018. Т. 17, № 5. С. 71–77. EDN: YQXVCP doi: 10.31631/2073-3046-2018-17-5-71-77
5. Злоказов М.Д. Эпидемиологические особенности возникновения бактериальных инфекций, специфичных для перинатального периода у новорожденных детей: дис. ... канд. мед. наук. Санкт-Петербург, 2020. 131 с. EDN: EJIEEN. Режим доступа: <https://www.dissercat.com/content/epidemiologicheskie-osobennosti-vozniknoveniya-bakterialnykh-infektsii-spetsifichnykh-dlya> Дата обращения: 07.10.2024.
6. Иванов Д.О., Петренко Ю.В., Курзина Е.А., Петрова Н.А. Показатели клинического анализа крови у новорожденных, заболевших неонатальным сепсисом // Бюллетень Федерального Центра сердца, крови и эндокринологии им. В.А. Алмазова 2012. № 3. С. 41–52. EDN: PCCWWL
7. Иванов Д.О., Шабалов Н.П., Петренко Ю.В. Неонатальный сепсис. Опыт построения гипотезы // Детская медицина Северо-Запада. 2012. Т. 3, № 3. С. 37–45. EDN: PWVKID

8. Ивжиц М.А., Зырянов С.К., Ушкалова Е.А., и др. Использование антибактериальных препаратов у недоношенных новорожденных: опыт создания формуляра // *Качественная клиническая практика*. 2016. № 3. С. 56–65. EDN: XHYKZZ
9. Антонов А.Г., Амирханова Д.Ю., Асташева И.Б., и др. Избранные клинические рекомендации по неонатологии / под ред. Е.Н. Байбариной, Д.Н. Дегтярева. Москва: ГЭОТАР-Медиа, 2016. 240 с.
10. Ионов О.В., Шарафутдинова Д.Р., Балашова Е.Н., и др. Факторы, ассоциированные с развитием некротизирующего энтероколита у новорожденных с экстремально низкой массой тела при рождении: ретроспективный анализ // *Неонатология: новости, мнения, обучение*. 2023. Т. 11, № 1. С. 29–41. EDN: ENHGDS doi: 10.33029/2308-2402-2023-11-1-28-41
11. Ионов О.В., Крохина К.Н., Горбачева Л.М., и др. Является ли лейкоцитоз значимым диагностическим маркером инфекционно-воспалительных заболеваний у недоношенных новорожденных в возрасте старше 72 ч жизни? // *Неонатология: новости, мнения, обучение*. 2016. № 1, С. 81–88.
12. Ионов О.В., Дегтярева А.В., Левадная А.В., и др. Клинико-лабораторные проявления врожденных инфекционно-воспалительных заболеваний у детей с экстремально низкой и очень низкой массой тела при рождении // *Акушерство и гинекология*. 2014. № 10. С. 66–71. EDN: SXRJXT
13. Князева Т.П. Причины и факторы риска преждевременного разрыва плодных оболочек // *Дальневосточный медицинский журнал*. 2016. № 2. С. 128–135. EDN: WCAXRZ
14. Справочник по антимикробной терапии. Выпуск 2 / под ред. Р.С. Козлова, А.В. Дехнич. Смоленск: МАКМАХ, 2010. 416 с.
15. Козлов С.Н., Страчунский Л.С. Современная антимикробная химиотерапия: руководство для врачей. Москва: Мед. Информ. Агенство, 2009. 448 с. EDN: QLUROV
16. Косенкова Е.Г., Лысенко И.М. Клинико-диагностические критерии реализации внутриутробного инфицирования у новорожденных и детей первого года жизни: монография. Витебск: ВГМУ, 2016. С. 184–200.
17. Лобзин Ю.В., Васильев В.В. Ключевые аспекты проблемы врожденных инфекций // *Журнал инфектологии*. 2014. Т. 6, № 3. С. 5–14. EDN: STQTXJ
18. Неонатология: национальное руководство. В 2 томах. Т. 2. / под ред. Н.Н. Володина, Д.Н. Дегтярева. 2-е изд., перераб. и доп. Москва: ГЭОТАР-Медиа, 2023. 768 с.
19. Овсянников Д.Ю., Бойцова Е.В., Жесткова М.А., и др. Неонатальная пульмонология: монография / под ред. Д.Ю. Овсянникова. Москва: Севен-Принт, 2022. 168 с. EDN: NGFFJV
20. Овсянников Д.Ю., Бойцова Е.В., Стуков Н.И., и др. Педиатрия: учебник: в 5 томах. Том 2: Оториноларингология, пульмонология, гематология, иммунология / под ред. Д.Ю. Овсянникова. Москва: РУДН, 2022. 592 с. EDN: EWEMYE
21. Овсянников Д.Ю., Бойцова Е.В. Пневмонии у новорожденных детей. Педиатрия // *Consilium Medicum*. 2021. № 3. С. 214–223. EDN: ENPMBP doi: 10.26442/26586630.2021.3.201060
22. Овсянников Д.Ю., Кравчук Д.А., Николаева Д.Ю. Клиническая патофизиология органов дыхания недоношенных детей // *Неонатология: новости, мнения, обучение*. 2018. Т. 6. № 3(21). С. 74–98. EDN: MHCYTZ doi: 10.24411/2308-2402-2018-13003
23. Брайант К.А., Кузман-Коттрилл Д.А. Руководство по профилактике инфекционных заболеваний в педиатрии / под ред. И.М. Османова, С.Н. Борзаковой. Москва: ГЭОТАР-Медиа, 2021. 333 с.
24. Савичева А.М. Вагинальный микробиом и виром — особенности взаимодействия // *Проблемы медицинской микологии*. 2023. Т. 25, № 2. С. 172. EDN: NSIDPM
25. Савичева А.М., Соколовский Е.В., Айламазян Э.К. Инфекционно-воспалительные заболевания в акушерстве и гинекологии. Руководство для врачей / под ред. Э.К. Айламазяна. Москва: ГЭОТАР-Медиа, 2016. 320 с. EDN: WJAQZN
26. Самсыгина Г.А. Неонатальный сепсис. Москва: ГЭОТАР-Медиа, 2020. 192 с.
27. Самсыгина Г.А. О предрасполагающих факторах и факторах риска развития неонатального сепсиса и о современных подходах его лечения // *Педиатрия. Журнал им. Г.Н. Сперанского*. 2012. Т. 91, № 3. С. 32–37. EDN: OZMAVN
28. Цинзерлинг В.А. Значение морфологических исследований в диагностике и изучении патогенеза инфекций. Тканевая микробиология // *Журнал инфектологии*. 2018. Т. 10, № 3. С. 124–132. EDN: VMVIXY doi: 10.22625/2072-6732-2018-10-3-124-132
29. Цинзерлинг В.А., Мельникова В.Ф. Перинатальные инфекции (вопросы патогенеза, морфологической диагностики и клинико-морфологических сопоставлений). Практическое руководство. Санкт-Петербург: Элби, 2002. 352 с.
30. Черняховский О.Б., Абрамова И.В., Полянчикова О.Л. Внутриутробные инфекции у новорожденных, факторы риска // *Российский вестник перинатологии и педиатрии*. 2009. Т. 54, № 1. С. 88. EDN: KPYMXZ
31. Шабалов Н.П., Иванов Д.О., Шабалова Н.Н. Сепсис новорожденных // *Медицинский академический журнал*. 2001. Т. 1, № 3. С. 81–86. EDN: WDQJYF
32. Шабалов Н.П. Общебиологическая проблема: закономерности и последствия перинатального инфицирования человека // *Педиатрия. Журнал им. Г.Н. Сперанского*. 2012, Т. 91, № 3. С. 26–31. EDN: OZMAUX
33. Шабалов Н.П. Софронова Л.Н. Неонатология: учебное пособие. В 2 томах. 7-е изд., перераб. и доп. Т. 2. Москва: ГЭОТАР-Медиа, 2020. 752 с. EDN: ANAYGR doi: 10.33029/9704-5771-9-NEO-2020-1-752
34. Шабалов Н.П., Софронова Л.Н. Неонатология: учебное пособие. В 2 томах. 7-е изд., перераб. и доп. Т. 1. Москва: ГЭОТАР-Медиа, 2020. 720 с. EDN: WUSRND doi: 10.33029/9704-5770-2-NEO-2020-1-720
35. Al-Matary A., Al Sulaiman M., Al-Otaiby S., et al. Association between the timing of antibiotics administration and outcome of neonatal sepsis // *J Infect Public Health*. 2022. Vol. 15, N 6. P. 643–647. doi: 10.1016/j.jiph.2022.05.004
36. Al-Lawama M., Aljbour H., Tanash A., Badran E. Intravenous Colistin in the treatment of multidrug-resistant *Acinetobacter* in neonates // *Ann Clin Microbiol Antimicrob*. 2016. Vol. 15. P. 8. doi: 10.1186/s12941-016-0126-4
37. EARSNet [Электронный ресурс]. Annual Report of the European Antimicrobial Resistance Surveillance Network Stockholm: ECDC; 2017. Режим доступа: <https://www.ecdc.europa.eu/en/about-us/networks/disease-networks-and-laboratory-networks/ears-net-data> Дата обращения: 08.10.2024.

38. Anugu N.R., Khan S. Comparing the diagnostic accuracy of procalcitonin and C-reactive protein in neonatal sepsis: a systematic review // *Cureus*. 2021. Vol. 13, N 11. P. e19485. doi: 10.7759/cureus.19485
39. Balamuth F., Weiss S.L., Fitzgerald J.C., et al. Protocolized treatment is associated with decreased organ dysfunction in pediatric severe sepsis // *Pediatr Crit Care Med*. 2016. Vol. 17, N 9. P. 817–822. doi: 10.1097/PCC.0000000000000858
40. Barichello T., Generoso J.S., Singer M., Dal-Pizzol F. Biomarkers for sepsis: more than just fever and leukocytosis — a narrative review // *Crit Care*. 2022. Vol. 26, N 1. P. 14. doi: 10.1186/s13054-021-03862-5
41. Bell E.F., Acarregui M.J. Restricted versus liberal water intake for preventing morbidity and mortality in preterm infants // *Cochrane Database Syst Rev*. 2014. Vol. 2014, N 12. P. CD000503. doi: 10.1002/14651858.CD000503
42. Berardi A., Buffagni A.M., Rossi C., et al. Serial physical examinations, a simple and reliable tool for managing neonates at risk for early-onset sepsis // *World J Clin Pediatr*. 2016. Vol. 5, N 4. P. 358–364. doi: 10.5409/wjcp.v5.i4.358
43. Beucher G., Charlier C., Cazanave C. Infection intra-utérine: diagnostic et traitement. RPC rupture prématurée des membranes avant terme CNGOF // *Gynecol Obstet Fertil Senol*. 2018. Vol. 46, N 12. P. 1054–1067. doi: 10.1016/j.gofs.2018.10.022
44. Bianchini S., Rigotti E., Nicoletti L., et al. Surgical antimicrobial prophylaxis in neonates and children with special high-risk conditions: A RAND/UCLA appropriateness method consensus study // *Antibiotics (Basel)*. 2022. Vol. 11, N 2. P. 246. doi: 10.3390/antibiotics11020246
45. Bedetti L., Miselli F., Minotti C., et al. Lumbar puncture and meningitis in infants with proven early- or late-onset sepsis: an Italian prospective multicenter observational study // *Microorganisms*. 2023. Vol. 11, N 6. P. 1546. doi: 10.3390/microorganisms11061546
46. Berardi A., Zinani I., Bedetti L., et al. Should we give antibiotics to neonates with mild non-progressive symptoms? A comparison of serial clinical observation and the neonatal sepsis risk calculator // *Front Pediatr*. 2022. Vol. 10. P. 882416. doi: 10.3389/fped.2022.882416
47. Cantey J.B., Lee J.H. Biomarkers for the diagnosis of neonatal sepsis // *Clin Perinatol*. 2021. Vol. 48, N 2. P. 215–227. doi: 10.1016/j.clp.2021.03.012
48. Çelik H.T., Portakal O., Yiğit Ş., et al. Efficacy of new leukocyte parameters versus serum C-reactive protein, procalcitonin, and interleukin-6 in the diagnosis of neonatal sepsis // *Pediatr Int*. 2016. Vol. 58, N 2. P. 119–125. doi: 10.1111/ped.12754
49. Clements K.E., Fisher M., Quaye K., et al. Surgical site infections in the NICU // *J Pediatr Surg*. 2016. Vol. 51, N 9. P. 1405–1408. doi: 10.1016/j.jpedsurg.2016.04.002
50. Collins A., Weitkamp J.H., Wynn J.L. Why are preterm newborns at increased risk of infection? // *Arch Dis Child Fetal Neonatal Ed*. 2018. Vol. 103, N 4. P. F391–F394. doi: 10.1136/archdischild-2017-313595
51. Conde-Agudelo A., Romero R., Jung E.J., Garcia Sánchez Á.J. Management of clinical chorioamnionitis: an evidence-based approach // *Am J Obstet Gynecol*. 2020. Vol. 223, N 6. P. 848–869. doi: 10.1016/j.ajog.2020.09.044
52. Coyne R., Hughes W., Purtill H., et al. Influence of an early human milk diet on the duration of parenteral nutrition and incidence of late-onset sepsis in very low birthweight (VLBW) infants: a systematic review // *Breastfeed Med*. 2024. Vol. 19, N 6. P. 425–434. doi: 10.1089/bfm.2023.0290
53. Craig A.M., Dotters-Katz S., Kuller J.A., Thompson J.L. Listeriosis in pregnancy: a review // *Obstet Gynecol Surv*. 2019. Vol. 74, N 6. P. 362–368. doi: 10.1097/OGX.0000000000000683
54. Czikk M.J., McCarthy F.P., Murphy K.E. Chorioamnionitis: from pathogenesis to treatment // *Clin Microbiol Infect*. 2011. Vol. 17, N 9. P. 1304–1311. doi: 10.1111/j.1469-0691.2011.03574.x
55. Davis A.L., Carcillo J.A., Aneja R.K., et al. American college of critical care medicine clinical practice parameters for hemodynamic support of pediatric and neonatal septic shock // *Crit Care Med*. 2017. Vol. 45, N 6. P. 1061–1093. doi: 10.1097/CCM.0000000000002425
56. De Rose D.U., Ronchetti M.P., Tzialla C., et al. Editorial: Congenital and perinatal infections: How to prevent sequelae in neonates and children // *Front Pediatr*. 2023. Vol. 11. P. 1142636. doi: 10.3389/fped.2023.1142636
57. Dempsey E., Chen M.F., Kokottis T., et al. Outcome of neonates less than 30 weeks gestation with histologic chorioamnionitis // *Am J Perinatol*. 2005. Vol. 22, N 3. P. 155–159. doi: 10.1055/s-2005-865020
58. Dior U.P., Kogan L., Eventov-Friedman S., et al. Very high intrapartum fever in term pregnancies and adverse obstetric and neonatal outcomes // *Neonatology*. 2016. Vol. 109, N 1. P. 62–68. doi: 10.1159/000440938
59. Donà D., Barbieri E., Daverio M., et al. Implementation and impact of pediatric antimicrobial stewardship programs: a systematic scoping review // *Antimicrob Resist Infect Control*. 2020. Vol. 9, N 1. P. 3. doi: 10.1186/s13756-019-0659-3
60. Eschborn S., Weitkamp J.H. Procalcitonin versus C-reactive protein: review of kinetics and performance for diagnosis of neonatal sepsis // *J Perinatol*. 2019. Vol. 39, N 7. P. 893–903. doi: 10.1038/s41372-019-0363-4
61. Evans I.V.R., Phillips G.S., Alpern E.R., et al. Association between the New York sepsis care mandate and in-hospital mortality for pediatric sepsis // *JAMA*. 2018. Vol. 320, N 4. P. 358–367. doi: 10.1001/jama.2018.9071
62. Freud A., Wainstock T., Sheiner E., et al. Maternal chorioamnionitis & long term neurological morbidity in the offspring // *Eur J Paediatr Neurol*. 2019. Vol. 23, N 3. P. 484–490. doi: 10.1016/j.ejpn.2019.03.005
63. Fuchs A., Bielicki J., Mathur S., et al. Reviewing the WHO guidelines for antibiotic use for sepsis in neonates and children // *Paediatr Int Child Health*. 2018. Vol. 38, Suppl 1. P. S3–S15. doi: 10.1080/20469047.2017.1408738
64. García-Muñoz Rodrigo F., Galán Henríquez G., Figueras Aloy J., García-Alix Pérez A. Outcomes of very-low-birth-weight infants exposed to maternal clinical chorioamnionitis: a multicentre study // *Neonatology*. 2014. Vol. 106, N 3. P. 229–234. doi: 10.1159/000363127. Epub 2014 Jul 5. Erratum in: *Neonatology*. 2015. Vol. 107, N 1. P. 42. PMID: 25011418.
65. García-Muñoz Rodrigo F., Galán Henríquez G.M., Ospina C.G. Morbidity and mortality among very-low-birth-weight infants born to mothers with clinical chorioamnionitis // *Pediatr Neonatol*. 2014. Vol. 55, N 5. P. 381–386. doi: 10.1016/j.pedneo.2013.12.007
66. Giannoni E., Dimopoulou V., Klingenberg C., et al. Analysis of antibiotic exposure and early-onset neonatal sepsis in Europe, North America, and Australia. *JAMA Netw Open*. 2022. Vol. 5, N 11. P. e2243691. doi: 10.1001/jamanetworkopen.2022.43691

67. Hedegaard S.S., Wisborg K., Hvas A.M. Diagnostic utility of biomarkers for neonatal sepsis — a systematic review // *Infect Dis (Lond)*. 2015. Vol. 47, N 3. P. 117–124. doi: 10.3109/00365548.2014.971053
68. Higgins R.D., Saade G., Polin R.A., et al. Evaluation and management of women and newborns with a maternal diagnosis of chorioamnionitis: summary of a workshop // *Obstet Gynecol*. 2016. Vol. 127, N 3. P. 426–436. doi: 10.1097/AOG.0000000000001246
69. Hofer N., Müller W., Resch B. Neonates presenting with temperature symptoms: role in the diagnosis of early onset sepsis // *Pediatr Int*. 2012. Vol. 54, N 4. P. 486–490. doi: 10.1111/j.1442-200X.2012.03570.x
70. Hofer N., Zacharias E., Müller W., Resch B. An update on the use of C-reactive protein in early-onset neonatal sepsis: current insights and new tasks // *Neonatology*. 2012. Vol. 102, N 1. P. 25–36. doi: 10.1159/000336629
71. Hornik C.P., Benjamin D.K., Becker K.C., et al. Use of the complete blood cell count in early-onset neonatal sepsis // *Pediatr Infect Dis J*. 2012. Vol. 31, N 8. P. 799–802. doi: 10.1097/INF.0b013e318256905c
72. Huttner A., Harbarth S., Carlet J., et al. Antimicrobial resistance: a global view from the 2013 World Healthcare-Associated Infections Forum // *Antimicrob Resist Infect Control*. 2013. Vol. 2. P. 31. doi: 10.1186/2047-2994-2-31
73. Imdad A., Rehman F., Davis E., et al. Effects of neonatal nutrition interventions on neonatal mortality and child health and development outcomes: A systematic review // *Campbell Syst Rev*. 2021. Vol. 17, N 1. P. e1141. doi: 10.1002/cl2.1141
74. Gao K., Fu J., Guan X., et al. Incidence, bacterial profiles, and antimicrobial resistance of culture-proven neonatal sepsis in South China // *Infect Drug Resist*. 2019. Vol. 12. P. 3797–3805.
75. Joshi N.S., Gupta A., Allan J.M., et al. Clinical monitoring of well-appearing infants born to mothers with chorioamnionitis // *Pediatrics*. 2018. Vol. 141, N 4. P. e20172056. doi: 10.1542/peds.2017-2056
76. Kim C.J., Romero R., Chaemsaihong P., et al. Acute chorioamnionitis and funisitis: definition, pathologic features, and clinical significance // *Am J Obstet Gynecol*. 2015. Vol. 213, Suppl 4. P. S29–S52. doi: 10.1016/j.ajog.2015.08.040
77. Kim M.J., Romero R., Gervasi M.T., et al. Widespread microbial invasion of the chorioamniotic membranes is a consequence and not a cause of intra-amniotic infection // *Lab Invest*. 2009. Vol. 89, N 8. P. 924–936. doi: 10.1038/labinvest.2009.49
78. Kimpton J.A., Verma A., Thakkar D., et al. Comparison of NICE Guideline CG149 and the sepsis risk calculator for the management of early-onset sepsis on the postnatal ward // *Neonatology*. 2021. Vol. 118, N 5. P. 562–568. doi: 10.1159/000518059
79. Kondo Y., Umemura Y., Hayashida K., et al. Diagnostic value of procalcitonin and presepsin for sepsis in critically ill adult patients: a systematic review and meta-analysis // *J Intensive Care*. 2019. Vol. 7. P. 22. doi: 10.1186/s40560-019-0374-4
80. Korang S.K., Safi S., Nava C., et al. Antibiotic regimens for early-onset neonatal sepsis // *Cochrane Database Syst Rev*. 2021. Vol. 2021, N 5. P. CD013837. doi: 10.1002/14651858.CD013837.pub2
81. Kuhn P., Dheu C., Bolender C., et al. Incidence and distribution of pathogens in early-onset neonatal sepsis in the era of antenatal antibiotics // *Paediatr Perinat Epidemiol*. 2010. Vol. 24, N 5. P. 479–487. doi: 10.1111/j.1365-3016.2010.01132.x
82. Kuzniewicz M.W., Puopolo K.M., Fischer A., et al. A quantitative, risk-based approach to the management of neonatal early-onset sepsis // *JAMA Pediatr*. 2017. Vol. 171, N 4. P. 365–371. doi: 10.1001/jamapediatrics.2016.4678
83. Kuzniewicz M.W., Mukhopadhyay S., Li S., et al. Time to positivity of neonatal blood cultures for early-onset sepsis // *Pediatr Infect Dis J*. 2020. Vol. 39, N 7. P. 634–640. doi: 10.1097/INF.0000000000002632
84. Lu B., Zhang Y., Li C., et al. The utility of presepsin in diagnosis and risk stratification for the emergency patients with sepsis // *Am J Emerg Med*. 2018. Vol. 36, N 8. P. 1341–1345. doi: 10.1016/j.ajem.2017.12.038
85. Maddaloni C., De Rose D.U., Santisi A., et al. The emerging role of presepsin (P-SEP) in the diagnosis of sepsis in the critically ill infant: a literature review // *Int J Mol Sci*. 2021. Vol. 22, N 22. P. 12154. doi: 10.3390/ijms222212154
86. Makkar M., Gupta C., Pathak R., et al. Performance evaluation of hematologic scoring system in early diagnosis of neonatal sepsis // *J Clin Neonatol*. 2013. Vol. 2, N 1. P. 25–29. doi: 10.4103/2249-4847.109243
87. Makkar N., Soneja M., Arora U., et al. Prognostic utility of biomarker levels and clinical severity scoring in sepsis: a comparative study // *J Investig Med*. 2022. Vol. 70, N 6. P. 1399–1405. doi: 10.1136/jim-2021-002276
88. Manurung T.N., Wungu C.D.K., Utomo M.T. The role of breast milk on reducing the risk of neonatal sepsis in preterm and low birth weight infants: a systematic review and meta-analysis // *Pharmacognosy Journal*. 2022. Vol. 14, N 6. P. 1067–1074. doi: 10.5530/pj.2022.14.211
89. Marks L., de Waal K., Ferguson J.K. Time to positive blood culture in early onset neonatal sepsis: A retrospective clinical study and review of the literature // *J Paediatr Child Health*. 2020. Vol. 56, N 9. P. 1371–1375. doi: 10.1111/jpc.14934
90. Mateus T., Silva J., Maia R.L., Teixeira P. Listeriosis during pregnancy: a public health concern // *ISRN Obstet Gynecol*. 2013. Vol. 2013. P. 851712. doi: 10.1155/2013/851712
91. Miller J.M., Binnicker M.J., Campbell S., et al. Guide to utilization of the microbiology laboratory for diagnosis of infectious diseases: 2024 update by the infectious diseases society of America (IDSA) and the American Society for Microbiology (ASM) // *Clin Infect Dis*. 2024. P. ciae104. doi: 10.1093/cid/ciae104
92. Mjelle AB, Guthe HJT, Reigstad H, et al. Serum concentrations of C-reactive protein in healthy term-born Norwegian infants 48–72 hours after birth // *Acta Paediatr*. 2019. Vol. 108, N 5. P. 849–854. doi: 10.1111/apa.14578
93. Moffett S.M., Kitts H.L., Henderson S.J. Medication therapy for early-onset neonatal sepsis // *AACN Adv Crit Care*. 2016. Vol. 27, N 3. P. 253–258. doi: 10.4037/aacnacc2016503
94. Murphy K., Weiner J. Use of leukocyte counts in evaluation of early-onset neonatal sepsis // *Pediatr Infect Dis J*. 2012. Vol. 31, N 1. P. 16–19. doi: 10.1097/INF.0b013e31822ffcf7
95. Lamont R.F., Sobel J., Mazaki-Tovi S., et al. Listeriosis in human pregnancy: a systematic review // *J Perinat Med*. 2011. Vol. 39, N 3. P. 227–236. doi: 10.1515/jpm.2011.035
96. Young T.E., Mangum O.B. Neofax: A manual of drugs used in neonatal care. American Society of Hospital Pharmacists, 2000. 272 p.
97. Neonatal infection: antibiotics for prevention and treatment. NICE. 2021. 81 p. Режим доступа: <http://nice.org.uk/guidance/ng195>
98. Newman T.B., Draper D., Puopolo K.M., et al. Combining immature and total neutrophil counts to predict early onset sepsis in term and late preterm newborns: use of the I/T2 // *Pediatr Infect Dis J*. 2014. Vol. 33, N 8. P. 798–802. doi: 10.1097/INF.0000000000000297

99. Ofman G., Vasco N., Cantey J.B. Risk of early-onset sepsis following preterm, prolonged rupture of membranes with or without chorioamnionitis // *Am J Perinatol.* 2016. Vol. 33, N 4. P. 339–342. doi: 10.1055/s-0035-1556758
100. Ozdemir A.A., Elgormus Y. Diagnostic value of presepsin in detection of early-onset neonatal sepsis // *Am J Perinatol.* 2017. Vol. 34, N 6. P. 550–556. doi: 10.1055/s-0036-1593851
101. Pammi M., Flores A., Versalovic J., Leeflang M.M. Molecular assays for the diagnosis of sepsis in neonates // *Cochrane Database Syst Rev.* 2017. Vol. 2, N 2. P. CD011926. doi: 10.1002/14651858.CD011926.pub2
102. Paul R., Neuman M.I., Monuteaux M.C., Melendez E. Adherence to PALS sepsis guidelines and hospital length of stay // *Pediatrics.* 2012. Vol. 130, N 2. P. e273–e280. doi: 10.1542/peds.2012-0094
103. Pierrakos C., Velissaris D., Bisdorff M., et al. Biomarkers of sepsis: time for a reappraisal // *Crit Care.* 2020. Vol. 24, N 1. P. 287. doi: 10.1186/s13054-020-02993-5
104. Poggi C., Bianconi T., Gozzini E., et al. Presepsin for the detection of late-onset sepsis in preterm newborns // *Pediatrics.* 2015. Vol. 135, N 1. P. 68–75. doi: 10.1542/peds.2014-1755
105. Poggi C., Lucenteforte E., Petri D., et al. Presepsin for the diagnosis of neonatal early-onset sepsis: a systematic review and meta-analysis // *JAMA Pediatr.* 2022. Vol. 176, N 8. P. 750–758. doi: 10.1001/jamapediatrics.2022.1647
106. Polin R.A.; Committee on Fetus and Newborn. Management of neonates with suspected or proven early-onset bacterial sepsis // *Pediatrics.* 2012. Vol. 129, N 5. P. 1006–1015. doi: 10.1542/peds.2012-0541
107. Pontrelli G., De Crescenzo F., Buzzetti R., et al. Accuracy of serum procalcitonin for the diagnosis of sepsis in neonates and children with systemic inflammatory syndrome: a meta-analysis // *BMC Infect Dis.* 2017. Vol. 17, N 1. P. 302. doi: 10.1186/s12879-017-2396-7
108. Pugaloni F., De Rose D.U., Kipfmüller F., et al. Assessment of hemodynamic dysfunction in septic newborns by functional echocardiography: a systematic review // *Pediatr Res.* 2024. Vol. 95, N 6. P. 1422–1431. doi: 10.1038/s41390-024-03045-2
109. Puopolo K.M., Draper D., Wi S., et al. Estimating the probability of neonatal early-onset infection on the basis of maternal risk factors // *Pediatrics.* 2011. Vol. 128, N 5. P. e1155–e1163. doi: 10.1542/peds.2010-3464
110. Raines D.A., Wagner A., Salinas A. Intraamniotic infection and the term neonate // *Neonatal Netw.* 2017. Vol. 36, N 6. P. 385–387. doi: 10.1891/0730-0832.36.6.385
111. Rees C.A., Lim J., Westbrook A.L., et al. Systematic review and meta-analysis of the diagnostic value of four biomarkers in detecting neonatal sepsis in low- and middle-income countries // *BMJ Paediatr Open.* 2023. Vol. 7, N 1. P. e001627. doi: 10.1136/bmjpo-2022-001627
112. Robin Ohis R.K., Yoder M.C. Hematology, immunology and infection disease: neonatology questions and controversies. Philadelphia: Elsevier Health Sciences, 2008. 312 p.
113. Rojas-Reyes M.X., Morley C.J., Soll R. Prophylactic versus selective use of surfactant in preventing morbidity and mortality in preterm infants // *Cochrane Database Syst Rev.* 2012. Vol. N 3. P. CD000510. doi: 10.1002/14651858
114. Romero R., Gomez-Lopez N., Winters A.D., et al. Evidence that intra-amniotic infections are often the result of an ascending invasion — a molecular microbiological study // *J Perinat Med.* 2019. Vol. 47, N 9. P. 915–931. doi: 10.1515/jpm-2019-0297
115. Saboohi E., Saeed F., Khan R.N., Khan M.A. Immature to total neutrophil ratio as an early indicator of early neonatal sepsis // *Pak J Med Sci.* 2019. Vol. 35, N 1. P. 241–246. doi: 10.12669/pjms.35.1.99
116. Sarkar S., Bhagat I., DeCristofaro J.D., et al. A study of the role of multiple site blood cultures in the evaluation of neonatal sepsis // *J Perinatol.* 2006. Vol. 26, N 1. P. 18–22. doi: 10.1038/sj.jp.7211410
117. Schlapbach L.J., Weiss S.L., Wolf J. Reducing collateral damage from mandates for time to antibiotics in pediatric sepsis — *primum non nocere* // *JAMA Pediatr.* 2019. Vol. 173, N 5. P. 409–410. doi: 10.1001/jamapediatrics.2019.0174
118. Seyoum K., Sahledengle B., Kene C., et al. Determinants of neonatal sepsis among neonates admitted to neonatal intensive care units in Ethiopian hospitals: A systematic review and meta-analysis // *Heliyon.* 2023. Vol. 9, N 9. P. e20336. doi: 10.1016/j.heliyon.2023.e20336
119. Sgro M., Kobylanski A., Yudin M.H., et al. Population-based study of early-onset neonatal sepsis in Canada // *Paediatr Child Heal.* 2019. Vol. 24, N 2. P. e66–e73. doi: 10.1093/pch/pxy018
120. Shane AL, Sánchez PJ, Stoll BJ. Neonatal sepsis // *Lancet.* 2017. Vol. 390, N 10104. P. 1770–1780. doi: 10.1016/S0140-6736(17)31002-4
121. Singer M., Deutschman C.S., Seymour C.W., et al. The third international consensus definitions for sepsis and septic shock (Sepsis-3) // *JAMA.* 2016. Vol. 315, N 8. P. 801–810. doi: 10.1001/jama.2016.0287
122. Stoll B.J., Hansen N.I., Sánchez P.J., et al. Early onset neonatal sepsis: The burden of group B streptococcal and *E. coli* disease continues // *Pediatrics.* 2011. Vol. 127, N 5. P. 817–826. doi: 10.1542/peds.2010-2217
123. Sturrock S., Sadoo S., Nanyunja C., Le Doare K. Improving the treatment of neonatal sepsis in resource-limited settings: gaps and recommendations // *Res Rep Trop Med.* 2023. Vol. 14. P. 121–134. doi: 10.2147/RRTM.S410785
124. Sun B., Liang L.F., Li J., et al. A meta-analysis of interleukin-6 as a valid and accurate index in diagnosing early neonatal sepsis // *Int Wound J.* 2019. Vol. 16, N 2. P. 527–533. doi: 10.1111/iwj.13079
125. Tita A.T., Andrews W.W. Diagnosis and management of clinical chorioamnionitis // *Clin Perinatol.* 2010. Vol. 37, N 2. P. 339–354. doi: 10.1016/j.clp.2010.02.003
126. Tuzun F., Ozkan H., Cetinkaya M., et al. Is European Medicines Agency (EMA) sepsis criteria accurate for neonatal sepsis diagnosis or do we need new criteria? // *PLoS One.* 2019. Vol. 14, N 6. P. e0218002. doi: 10.1371/journal.pone.0218002
127. van Leeuwen L.M., Fourie E., van den Brink G., et al. Diagnostic value of maternal, cord blood and neonatal biomarkers for early-onset sepsis: a systematic review and meta-analysis // *Clin Microbiol Infect.* 2024. Vol. 30, N 7. P. 850–857. doi: 10.1016/j.cmi.2024.03.005
128. Varghese A., Blaschke A.J., Korgenski E.K., Crandall H. Neonatal early-onset sepsis due to *Haemophilus influenzae* in Utah // *Pediatr Infect Dis J.* 2023. Vol. 42, N 3. P. e90–e92. doi: 10.1097/INF.0000000000003795
129. Vatne A., Klingenberg C., Rettedal S., Øymar K. Early-onset sepsis in neonates — a population-based study in South-West Norway from 1996 to 2018 // *Front Pediatr.* 2021. Vol. 9. P. 634798. doi: 10.3389/fped.2021.634798
130. Walker S., Datta A., Massoumi R.L., et al. Antibiotic stewardship in the newborn surgical patient: A quality improvement project in the neonatal intensive care unit // *Surgery.* 2017. Vol. 162, N 6. P. 1295–1303. doi: 10.1016/j.surg.2017.07.021

- 131.** Wiechers C., Bernhard W., Goelz R., et al. Optimizing early neonatal nutrition and dietary pattern in premature infants // *Int J Environ Res Public Health*. 2021. Vol. 18, N 14. P. 7544. doi: 10.3390/ijerph18147544
- 132.** Wing E.J., Gregory S.H. *Listeria monocytogenes*: clinical and experimental update // *J Infect Dis*. 2002. Vol. 185, Suppl 1. P. S18–S24. doi: 10.1086/338465
- 133.** Yapıcıoğlu H., Özlü F., Sertdemir Y. Are vital signs indicative for bacteremia in newborns? // *J Matern Fetal Neonatal Med*. 2015. Vol. 28, N 18. P. 2244–2249. doi: 10.3109/14767058.2014.983896
- 134.** Yochpaz S., Friedman N., Zirkin S., et al. C-reactive protein in early-onset neonatal sepsis — a cutoff point for CRP value as a predictor of early-onset neonatal sepsis in term and late preterm infants early after birth? // *J Matern Fetal Neonatal Med*. 2022. Vol. 35, N 23. P. 4552–4557. doi: 10.1080/14767058.2020.1856068
- 135.** Zachariah P., Saiman L. Expanding antimicrobial stewardship strategies for the NICU: Management of surgical site infections, perioperative prophylaxis, and culture negative sepsis // *Semin Perinatol*. 2020. Vol. 44, N 8. P. 151327. doi: 10.1016/j.semperi.2020.151327

AUTHORS' INFO

Dmitry O. Ivanov, MD, PhD, Dr. Sci. (Medicine), Professor, Chief neonatologist of the Ministry of Health of the Russian Federation, Head of the Department of Neonatology with Courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Rector, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0002-0060-4168; eLibrary SPIN: 4437-9626; e-mail: delopro@gpmu.org.ru

ОБ АВТОРАХ

Дмитрий Олегович Иванов, д-р мед. наук, профессор, главный неонатолог Минздрава России, заведующий кафедрой неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ректор, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0002-0060-4168; eLibrary SPIN: 4437-9626; e-mail: delopro@gpmu.org.ru

AUTHORS' INFO

***Alexandra S. Panchenko**, MD, PhD, Dr. Sci. (Medicine), Professor of the Department of Neonatology with courses of Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation; address: 2 Litovskaya st., Saint Petersburg, 194100, Russia; ORCID: 0000-0003-2313-3941; eLibrary SPIN: 9552-2350; e-mail: sashawomen18@mail.ru

Larisa A. Fedorova, MD, PhD, Associate Professor of the Department of Neonatology with courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0001-9747-762X; eLibrary SPIN: 5474-0902; e-mail: arslarissa@rambler.ru

Ekaterina N. Balashova, MD, PhD, Assistant of the Department of Neonatology, Research Center for Obstetrics, Gynecology and Perinatology, Ministry of Healthcare of the Russian Federation, Moscow, Russia; ORCID: 0000-0002-3741-0770; eLibrary SPIN: 1335-1489; e-mail: katbal99@gmail.com

Elena V. Bem, MD, PhD, Assistant of the Department of Neonatology with courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0009-0008-9337-5667; eLibrary SPIN: 2811-5938; e-mail: e.bohm@inbox.ru

Nazar M. Zelenin, anesthesiologist-resuscitator, Head of the Department, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; eLibrary SPIN: 1101-2442; e-mail: Piorun1944@gmail.com

Irina V. Myznikova, Head of the Department of Pathology of Newborns and Young Children of the Perinatal Center, Assistant of the Department of Neonatology with courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0002-8624-1854; e-mail: irinayurko2014@yandex.ru

Marina I. Levadneva, Head of the Department of Newborn Physiology at the Perinatal Center, Assistant of the Department of Neonatology with Courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0001-6716-7567; eLibrary SPIN: 2780-6674; e-mail: m-lev11@yandex.ru

ОБ АВТОРАХ

***Александра Сергеевна Панченко**, д-р мед. наук, профессор кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России; адрес: Россия, 194100, Санкт-Петербург, ул. Литовская, д. 2; ORCID: 0000-0003-2313-3941; eLibrary SPIN: 9552-2350; e-mail: sashawomen18@mail.ru

Лариса Арзумановна Федорова, канд. мед. наук, доцент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0001-9747-762X; eLibrary SPIN: 5474-0902; e-mail: arslarissa@rambler.ru

Екатерина Николаевна Балашова, канд. мед. наук, ведущий научный сотрудник отделения реанимации и интенсивной терапии им. проф. А.Г. Антонова, доцент кафедры неонатологии института неонатологии и педиатрии, ФГБУ «Национальный медицинский исследовательский центр акушерства, гинекологии и перинатологии им. акад. В.И. Кулакова» Минздрава России, Москва, Россия; ORCID: 0000-0002-3741-0770; eLibrary SPIN: 1335-1489; e-mail: katbal99@gmail.com

Елена Венедиктовна Бем, канд. мед. наук, ассистент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0009-0008-9337-5667; eLibrary SPIN: 2811-5938; e-mail: e.bohm@inbox.ru

Назар Мартович Зеленин, врач анестезиолог-реаниматолог, заведующий отделением реанимации новорожденных Перинатального центра, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; eLibrary SPIN: 1101-2442; e-mail: Piorun1944@gmail.com

Ирина Владимировна Мызникова, заведующая отделением патологии новорожденных и детей раннего возраста Перинатального центра, ассистент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0002-8624-1854; e-mail: irinayurko2014@yandex.ru

Марина Ивановна Леваднева, заведующая отделением физиологии новорожденных Перинатального центра, ассистент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0001-6716-7567; eLibrary SPIN: 2780-6674; e-mail: m-lev11@yandex.ru

* Corresponding author / Автор, ответственный за переписку

AUTHORS' INFO

Anna S. Nabieva, MD, PhD, Head of the Epidemiological Department, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0002-2519-7589; eLibrary SPIN: 4579-1621; e-mail: hamatum@bk.ru

Svetlana E. Pavlova, Assistant Professor of the Department of Neonatology with courses of Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0001-5423-0950; eLibrary SPIN: 1761-4768; e-mail: svetlanapav.spb@mail.ru

Larisa A. Romanova, MD, PhD, Associate Professor of the Department of Neonatology with courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0003-0828-352X; eLibrary SPIN: 6460-5491; e-mail: l_romanova2011@mail.ru

Galina N. Chumakova, MD, PhD, Dr. Sci. (Medicine), Professor of the Department of Neonatology with courses in Neurology and Obstetrics and Gynecology of Faculty of Retraining and Additional Professional Education, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0001-5353-4610; eLibrary SPIN: 1821-0198; e-mail: zelchum-neo@yandex.ru

Ekaterina E. Yakovleva, MD, PhD, Head of the Department of Clinical Pharmacology, Associate Professor of the Department of Pharmacology with the course of Clinical Pharmacology and Pharmacoeconomics, Saint Petersburg State Pediatric Medical University, Ministry of Health of the Russian Federation, Saint Petersburg, Russia; ORCID: 0000-0002-0270-0217; eLibrary SPIN: 6728-7340; e-mail: eeiakovleva@mail.ru

Tamara V. Belousova, MD, PhD, Dr. Sci. (Medicine), Professor, Honored Doctor of Russia, Head of the Department of Pediatrics and Neonatology, Novosibirsk State Medical University, Ministry of Health of the Russian Federation, Novosibirsk, Russia; ORCID: 0000-0002-4234-9353; eLibrary SPIN: 9651-0155; e-mail: belousovatv03@yandex.ru

Irina Ya. Izvekova, MD, PhD, Dr. Sci. (Medicine), Professor of the Department of Infectious Diseases, Novosibirsk State Medical University, Ministry of Health of the Russian Federation, Novosibirsk, Russia; ORCID: 0009-0003-6617-6797; e-mail: izvekova@inbox.ru

Dmitry Yu. Ovsyannikov, MD, PhD, Dr. Sci. (Medicine), Head of the Department of Pediatrics, Peoples' Friendship University of Russia named after Patrice Lumumba, Moscow, Russia; ORCID: 0000-0002-4961-384X; eLibrary SPIN: 5249-5760; e-mail: mdovsyannikov@yahoo.com

ОБ АВТОРАХ

Анна Сергеевна Набиева, канд. мед. наук, заведующий эпидемиологическим отделом, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава Российской Федерации, Санкт-Петербург, Россия; ORCID: 0000-0002-2519-7589; eLibrary SPIN: 4579-1621; e-mail: hamatum@bk.ru

Светлана Евгеньевна Павлова, ассистент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0001-5423-0950; eLibrary SPIN: 1761-4768; e-mail: svetlanapav.spb@mail.ru

Лариса Андреевна Романова, канд. мед. наук, доцент кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава Российской Федерации, Санкт-Петербург, Россия; ORCID: 0000-0003-0828-352X; eLibrary SPIN: 6460-5491; e-mail: l_romanova2011@mail.ru

Галина Николаевна Чумакова, д-р мед. наук, профессор кафедры неонатологии с курсами неврологии и акушерства-гинекологии ФП и ДПО, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0001-5353-4610; eLibrary SPIN: 1821-0198; e-mail: zelchum-neo@yandex.ru

Екатерина Евгеньевна Яковлева, канд. мед. наук, заведующая отделением клинической фармакологии, доцент кафедры фармакологии с курсом клинической фармакологии и фармакоэкономики, ФГБОУ ВО «Санкт-Петербургский государственный педиатрический медицинский университет» Минздрава России, Санкт-Петербург, Россия; ORCID: 0000-0002-0270-0217; eLibrary SPIN: 6728-7340; e-mail: eeiakovleva@mail.ru

Тамара Владимировна Белоусова, д-р мед. наук, профессор, Заслуженный врач России, заведующая кафедрой педиатрии и неонатологии, ФГБОУ ВО «Новосибирский государственный медицинский университет» Минздрава России, Новосибирск, Россия; ORCID: 0000-0002-4234-9353; eLibrary SPIN: 9651-0155; e-mail: belousovatv03@yandex.ru

Ирина Яковлевна Извекова, д-р мед. наук, профессор кафедры инфекционных болезней, ФГБОУ ВО «Новосибирский государственный медицинский университет» Минздрава России, Новосибирск, Россия; ORCID: 0009-0003-6617-6797; e-mail: izvekova@inbox.ru

Дмитрий Юрьевич Овсянников, д-р мед. наук, заведующий кафедрой педиатрии, ФГАОУ ВО «Российский университет дружбы народов им. Патриса Лумумбы», Москва, Россия; ORCID: 0000-0002-4961-384X; eLibrary SPIN: 5249-5760; e-mail: mdovsyannikov@yahoo.com