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Anatomical, physiological and pathophysiological features of the lower urinary tract in gender and age aspects

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In the review article, based on the results of modern clinical and experimental studies, gender and age-related features of the anatomy, physiology and pathophysiology of the lower urinary tract are considered. The features of the structure and functioning of the urothelium, myoepithelium, neuroepithelium and endothelium of the lower urinary tract in men and women are described in detail. A separate section of the review is devoted to the peculiarities of hormonal regulation of the lower urinary tract, depending on gender and age.

Keywords: sexual dimorphism; lower urinary tract; detrusor; urethra; pelvic floor; urothelium; endothelium; myoepithelium; neuroepithelium; sex steroid hormones.

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Анатомо-физиологические и патофизиологические особенности нижних мочевых путей в гендерном и возрастном аспектах

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В обзорной статье на основе результатов современных клинко-экспериментальных исследований рассматриваются гендерные и возрастные особенности анатомии, физиологии и патофизиологии нижних мочевых путей. Подробно описаны особенности строения и функционирования уротелия, миотелия, нейротелия и эндотелия нижних мочевых путей у мужчин и женщин. Отдельный раздел обзора посвящен особенностям гормональной регуляции нижних мочевых путей в зависимости от половой принадлежности и возраста.

Ключевые слова: половой диморфизм; нижние мочевые пути; детрузор; уретра; тазовое дно; уротелий; эндотелий; миотелий; нейротелий; половые стероидные гормоны.

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INTRODUCTION

Sexual identity is a fundamental aspect of human physiology, which equally divides the entire living population into two, but this significant biological variable is rarely considered when developing basic physiological research, when transferring results from basic science into clinical research, or when developing personalized medical strategies [1]. Sexual differentiation is one of the most important biological characteristics of all living organisms, including humans, and it has a significant and sometimes key influence on the formation, development, and functioning of many organs and organ systems [2, 3].

The lower urinary tract (LUT) is one of the classic examples of sexual structural and functional dimorphism, which is actively studied to optimize modern diagnostics and pharmacotherapy of LUT diseases in relation to the gender identity of the patients [4–6]. Sexual dimorphism is characteristic of nearly all anatomical structures that form the LUT. However, there is a new hypothetical endocrine–autocrine–paracrine model of interactions

between the urothelium, afferent and efferent nerve terminals (neurothelium), vessels (endothelium), muscle cells (myothelium), and myofibroblasts of all LUT structures. According to this model, all these structures constitute a single anatomical and functional system, and its elements are in constant interaction and interinfluence with each other (Fig. 1) [7–11].

Sexual aspects of the LUT urothelium

The urothelium is a specialized epithelium that is located in its connective tissue plate (lamina propria) and lines the LUT inside in the form of 5–7 rows of cells represented by three layers. These layers appeared apical (one layer of umbrella cells, which are in direct contact with urine and serve as a physiological barrier between it and underlying tissues), intermediate (2–3 layers of intermediate cells), and basal (2–3 layers of basal cells) [12]. When the bladder is full, the umbrella cells stretch and flatten, and when the bladder is empty, the cells are cuboidal [13]. The umbrella cells of the superficial urothelium got their name because, in 70%–90% of the cases, peculiar outgrowths (plaques) are found on their

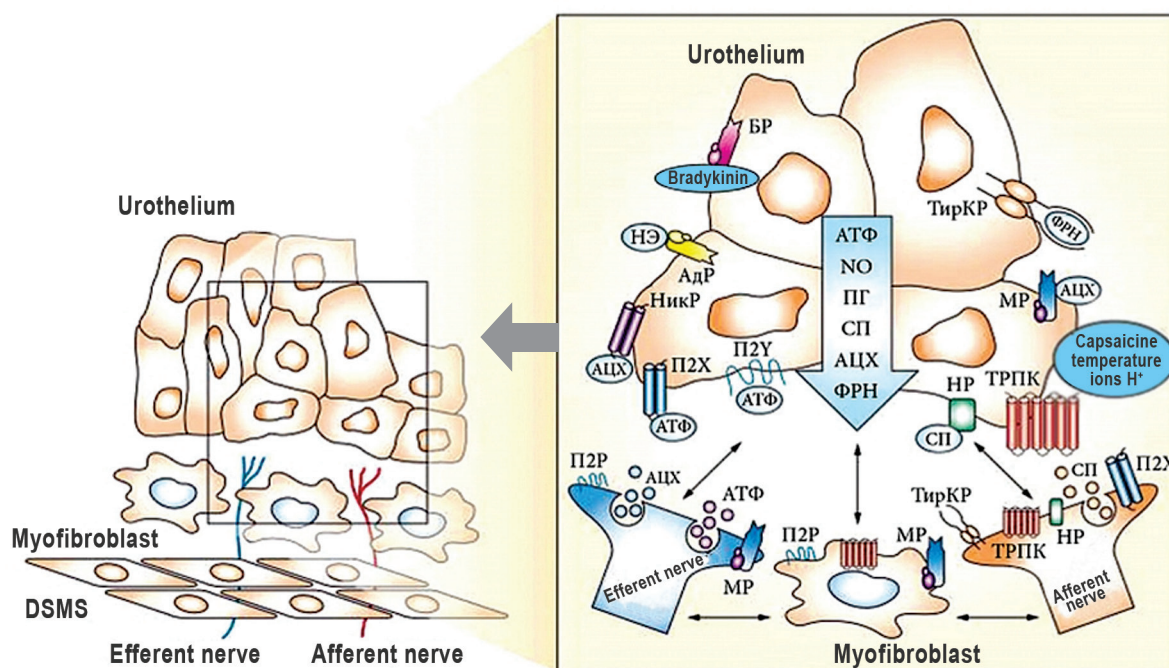


Fig. 1. A hypothetical model of interactions between urothelial cells (urothelium), afferent and efferent nerve endings (neurothelium), vessels (endothelium), smooth muscle cells and myofibroblasts of the lower urinary tract [10, 11]. NO – nitrogen oxide; АдР – adrenergic receptor; АТФ – adenosine triphosphate; АЦХ – acetylcholine; Бр – bradykinin receptor; ГКД – detrusor smooth muscle cell; МР – muscarinic receptor; НикР – nicotinic receptor; НР – neurokinin receptor; НЭ – norepinephrine; П2Р – purinergic receptor subtype 2; П2Х and П2У – purinergic receptors of the X and Y subtypes; ПГ – prostaglandins; СП – substance R; ТирКР – tyrosine kinase receptor with high affinity for nerve growth factor; ТРПК – transit receptor of potential channels; ФРН – nerve growth factor

Рис. 1. Гипотетическая модель взаимодействий между уротелиальными клетками (уротелием), афферентными и эфферентными нервными окончаниями (нейротелием), сосудами (эндотелием), гладкомышечными клетками и миофибробластами нижних мочевых путей [10, 11]. NO – оксид азота; АдР – адренергический рецептор; АТФ – аденозинтрифосфат; АЦХ – ацетилхолин; Бр – брадикининовый рецептор; ГКД – гладкомышечная клетка детрузора; МР – мускариновый рецептор; НикР – никотиновый рецептор; НР – нейрокининовый рецептор; НЭ – норэпинефрин; П2Р – пуринергический рецептор подтипа 2; П2Х и П2У – пуринергические рецепторы подтипов Х и У; ПГ – простагландины; СП – субстанция Р; ТирКР – тирозинкиназный рецептор с высокой аффинностью к фактору роста нервов; ТРПК – транзитный рецептор потенциальных каналов; ФРН – фактор роста нервов

membrane facing the lumen of the urinary tract, giving the cells a "toothed" appearance and consisting of uroplakins that form macromolecular structures (hexagonal shape), providing, along with tight junction proteins, the barrier function of the urothelium [14].

Currently, several types of uroplakins have been identified in the urothelium, namely, UP-Ia (molecular weight, 27 kDa), UP-Ib (molecular weight, 28 kDa), UP-II (molecular weight, 15 kDa), and UP-III (molecular weight, 47 kDa) [15–17]. Hu et al. [18] examined male and female rats and found that the identification of UP-II in these animals was associated with defective glycosylation, smaller urothelial plaques of umbrella cells, and increased their water permeability. Aboushwareb et al. [19] revealed specific sex differences in UP-II- and UP-III-knockout mice. Thus, male mouse knockout of UP-II showed signs of functional bladder decompensation, presenting as a decrease in effective pressure and an increase in residual urine. In female rats with the same knockout, no changes were detected. A study of the excitability of detrusor myocytes in uroplakin-knockout mice revealed sex differences, where female rats showed decreased excitability, while male ones showed no changes in myocyte excitability [19]. These data confirmed that uroplakin deficiency in the urothelium can induce bladder dysfunction.

In addition to its barrier function, the urothelium performs the most important sensory function, as it contains numerous ion channels and regulatory proteins of various receptors (i.e., adenosinergic, purinergic, adrenergic, bradykinin, neurokinin, muscarinic, and cholinergic), which ensure the interaction of urothelial cells with each other and unites them into a single functional system [20, 21]. The urothelium releases various small molecules and neurotransmitters in response to a mechanical or chemical stimulus. In addition, because the urothelium contains specialized acid-sensitive ion channels, it maintains a constant pH level and controls bladder sensitivity. The expressions of ASIC1 are more pronounced in male rats, while that of ASIC2 is more pronounced in female ones [22]. Although the impaired expression of ASIC1 channels in the intestine decreases its mechanical sensitivity, the functional consequences of this difference in the expression of these channels in the urinary bladder remain unknown [23].

The urothelium of the urinary bladder also plays an important role in the innate immune response when uropathogens are detected, and the higher incidence of LUT infections in women suggests a significant regulatory role of estrogen in this process. Studies on female mice and urothelial tissues of women have shown that estrogens mediate the defense mechanisms of the urothelium against *Escherichia coli* by regulating the activity of type β (ER- β) estrogen receptors identified in the urothelium; therefore, ER- β plays an important role in the

pathogenesis of inflammatory diseases of the bladder in women [24–26].

Recent studies have revealed that high-conductivity potassium channels (BK channels) in the umbrella cells of the surface urothelial layer are regulated by calcium and lipopolysaccharides [27, 28]. A study suggested a relationship between the activity of urothelial BK channels and the expression of estrogen receptors, i.e., ER- β type, in the implementation of the urothelial innate immune response in LUT infections in women, whereas estradiol increases the expression of BK channels, and the blockade of its effects led to the opposite effect in an experimental mouse model of oophorectomy [29].

Myoethelial sexual dimorphism of the LUT

Detrusor. The normal bladder is a reliable reservoir controlled by the nervous system for storing urine and its periodical emptying. It represents a complex of smooth muscle elements combined into a single functional syncytium [30, 31]. The detrusor muscle is thicker in men than in women because more pressure is required to empty the bladder through the longer male urethra [32]. The ratio between the smooth muscles and connective tissues in the detrusor is not different in women and men of any age, and studies have shown that the contractile ability of the human detrusor is independent of sex [12, 33].

The human bladder can be detected as early as week 10 of intrauterine development. Although the trigone is believed to be of mesodermal origin, the rest of the bladder originates from the endoderm; some studies have shown that the trigone also has an endodermal origin [34]. Favorito et al. [35] evaluated morphological differences in the detrusor smooth muscle of women and men and did not find differences in the volumetric density of fetal nerves, smooth muscle cells, or collagen at weeks 13–20 after conception [35].

Urethra. The muscle layer of the urethra contains oblique and longitudinal muscle fibers surrounded by circular fibers in both women and men. The muscular layer provides basic resistance to urine flow, which is further enhanced by the rich vascularization of the urethra [36]. In addition, $\alpha 1$ and $\alpha 2$ adrenergic receptors, consisting of several subtypes and regulating the contractility of smooth myocytes, contribute to the functions of the urethra in both sexes. Male rabbits have the same number of $\alpha 1$ and $\alpha 2$ adrenergic receptors, but female rabbits have a much higher density of $\alpha 2$ adrenergic receptors [37]. Alexandre et al. [38] examined the effect of various agonists and antagonists on urethral smooth muscles and revealed that phenylephrine, norepinephrine, potassium chloride, and stimulation with an electric field caused stronger muscle contractions in men. However, no sex differences were noted in response to the administration of N-nitro-L-arginine, atropine,

or a P2X1-purine receptor antagonist. The expressions of the RNAs of α 1A-adrenergic receptor and tyrosine hydroxylase in the urethra were significantly higher in men than in women. Therefore, α 1-adrenergic receptors may not be very important for the contraction and functionality of the urethra in women [38].

Oswald et al. [39] studied the intrauterine development of the internal urethral sphincter in 37 human fetuses and found that the volume of the internal sphincter is significantly greater in male fetuses than in female fetuses, which is partially caused by muscular hypertrophy and leads to a decrease in the urethral lumen [39]. This may be due to transient urethral obstruction distal to the bladder neck during the hormone-dependent growth stimulation, particularly by testosterone, in men. Jin et al. [40] showed that the differentiation of smooth muscle cells in the bladder and urethra is crucial in the development of mesonephric duct prolapse during intrauterine fetal development [40].

The striated musculature provides pelvic floor support and coordinates urinary initiation and bladder emptying. The architecture of the striated muscles in the detrusor and urethra coincides with the arrangement of muscle fibers and connective tissues [41, 42]. Striated muscle fibers have two types, namely, slow contraction (type I) and fast contraction (type II) muscle fibers [43]. Type I fibers have more acid-resistant ATPases, more mitochondria, thicker Z-disks, higher amounts of oxidative enzymes, and a contraction time of approximately 100 ms. Type II fibers have a higher concentration of alkali-resistant ATPases, fewer mitochondria, and a contraction time of approximately 30 ms [43, 44]. The fiber type of striated muscles affects their susceptibility to damage and repair and varies by sex. In rats, the female and male urethras differ macro- and microscopically. In contrast to typical skeletal muscles, the myofibrils of the female urethral sphincter are 3–5 times smaller in diameter than the striated pelvic floor muscles because these cells lack peripheral nuclear localization [45]. Instead, the nuclei of myocytes are located in the center, with sizes similar to the diameter of the fibril. Unlike other skeletal muscles, these cells have no attachment points and are in direct contact with the adjacent connective tissue [46]. Similar to skeletal muscles, the striated musculature of the rat urethra expresses desmin and dystrophin; desmin runs orthogonally to Z-shaped lines, outlining sarcomeres, while dystrophin is localized in the sarcolemma of myocytes in female rats [45]. In male rats, two longitudinal bands of connective tissue segment the sphincter into two lateral bundles, and myofibrils do not form myotendinous connections with adjacent connective tissue. The striated elements of the sphincter form a longitudinal thick layer visible in male rats, while the sphincter is thin and more rounded in female rats [46]. Bierinx et al. [47] found two types of

myofibrils in the urethra of male rats, which were characterized predominantly by type II “fast” fibers, similar to female rats; however, individual fibers taken near the urethral lumen contained slow contraction myofibrils (type I). In addition to the bladder neck, striated muscles are located in the middle part of the urethra, and recent studies using immunohistochemical methods have shown that rapid-contraction muscle fibers predominate among the fibers [48]. Chen et al. [49] identified fast contraction fibers in the proximal urethra in male rats.

In humans, the structure of the muscular apparatus of the urethra is significantly different between men and women. When using histochemical methods and electron microscopic analysis of samples obtained using cystourethrectomy, Gosling et al. [44] identified type I muscle fibers of 15–20 μ m in diameter (slow contraction fibers) in the urethra of both men and women and muscle fibers of both types in *m. levator ani* 3. Other authors revealed that striated muscles of the female urethra consist mainly of type I fibers [50–52], while the striated muscles extend from the male urethra through the prostate and consist of both slow and fast fibers of various diameters; however, its functional significance remains unknown [53, 54]. In humans, the identification of nitrergic nerve fibers in rhabdosphincter suggests that nitric oxide (NO) plays an important role in the control of striated muscles in the urethra, regulating their relaxation and contractility [55–57]. Ho et al. [53] showed the presence of NO synthases (NOS) in 86% of fast muscle fibers and in 29% of slow muscle fibers in the sarcolemma of myocytes of the male urethra [53].

Urogenital pelvic diaphragm. The pelvic floor musculature supports the abdominal and pelvic organs and regulates the mechanisms of the retention of feces and urine [58]. The muscular components of the pelvic floor include the levator muscles and coccygeal muscle [59]. *M. levator ani* contains a heterogeneous population of type I and II muscle fibers, but histological studies have shown a predominance of type I fibers, which clinically correlates with the static nature of the pelvic floor and its role in maintaining the internal organs of the abdominal cavity and pelvis [44]. A smaller population of type II fibers supports the pelvic floor during periods of increased abdominal pressure, and the number and diameter of these fibers decrease with age. However, their proportions are not different between men and women [60, 61]. Tobin et al. [62] found that the *m. levator ani* of a rat embryo exhibits sexual dimorphism; during antenatal development, male embryos contain significantly fewer motor units (153 versus 350), and each unit has a relatively smaller cross-sectional area (89.2 μ m² versus 120.8 μ m²) compared with that in female fetuses. With the postnatal development of rats, the cross-sectional area of the motor units decreased slightly in both sexes, but the number of motor units in male rats increased

rapidly to 2726 by day 6 after birth, which is caused by the influence of testosterone [62]. In their experimental studies, Niel et al. [63] showed that satellite cells (a population of myogenic stem cells) located on the periphery of the pelvic floor muscle fibers are influenced by androgens and can play a role in sexual dimorphism. Thus, the number of satellite cells in *m. levator ani* of newborn male rats was higher than that in newborn females. With prenatal exposure to testosterone, the number and size of satellite muscle cells in female rats increased; therefore, sex differences in muscle tissue found in developing rats are due to sexual dimorphism in satellite cells sensitive to androgens [63]. These animal studies are consistent with a study that analyzed sex differences in human fetuses and showed that, during development, *m. levator ani* forms as a thick muscle layer in boys, while it is a thin muscle in girls, and its bundles are integrated with connective tissue [64].

Sex characteristics of the regulation of endothelial function and blood supply to the LUT in terms of age

The endothelium mainly helps in maintaining vascular homeostasis by synthesizing and secreting substances involved in the expansion (vasodilators) and narrowing (vasoconstrictors) of the vessels. Progressive endothelial dysfunction, as one of the aspects of vascular aging, has been identified as a key initiating stage in the pathogenesis of atherosclerosis [65]. Endothelial function is controlled by several factors that differ in men and women, and it is associated with the expression of sex hormone receptors in the endothelium, which have a major effect on endothelial metabolism and consequently on the tone of the vascular wall and regional blood flow [65]. Thus, differences in the reaction of blood flow to the infusion of intraperitoneal acetylcholine in men and women have been established. In particular, in normotensive premenopausal women, acetylcholine-mediated vasodilation slightly decreased (by approximately 0.5% annually); however, disorders of endothelium-dependent vasodilation became evident only after menopause, in which the response to acetylcholine decreased faster (2.1% per year). In women, no sex differences in smooth muscle function response to sodium nitroprusside administration were noted, which confirmed the effect of menopause on endothelium-dependent rather than endothelium-independent vasodilation [66]. The well-known consequences of menopause on LUT are united by the term “genitourinary menopausal syndrome,” which includes the following conditions: decreased volume and speed of blood circulation in LUT structures, thinning of the mucous membranes and urothelium, decreased bactericidal and barrier function of the urothelium, hypoxia, ischemia, and fibrosis in pelvic

organs, which increase significantly the risk of vulvovaginal and cystourethral atrophic changes [67]. In women of reproductive age, due to cyclical changes in sex hormones during the menstrual cycle, the same cyclical changes in the blood flow of the LUT are observed, i.e., from the main pronounced arterial blood flow in phase 1 of the cycle (estrogen effects) to its minor decrease and development of the veno-discirculatory phenomena in the small pelvis in phase 2 (effects of progesterone). Moreover, cyclical changes in estrogen and progesterone levels in women are associated with increased severity of LUT symptoms (LUTS) immediately before menstruation [67]. Moderately obese postmenopausal women may be more susceptible to endothelial dysfunction than men and postmenopausal women without moderate obesity, whether or not they have type 2 diabetes mellitus [68]. In addition, the length of the urethra in women increases in the middle of the menstrual cycle at the peak of the effect of estrogen [69]. By contrast, in men with normal blood pressure levels, acetylcholine-induced vasodilation decreased with age (approximately 1.8% per year), which coincided with the average rate of the decrease in the level of testosterone secretion in men. This may indicate the role of testosterone in the synthesis of NO in the vascular endothelium and its pronounced vasoprotective effect. Numerous clinical and experimental studies have shown a significant relationship among low testosterone levels, pelvic atherosclerosis, ischemia, and fibrosis of the LUT in men [70–72].

The total area of the pelvic arterial bed in both men and women is significantly lower than the total density of all venous collectors, which determines a high frequency of the formation of venous congestion in this anatomical region with the involvement of the LUT in the process, often simulating urological pathology [73]. Moreover, in men, a relationship is found between the density of the vascular bed of the small pelvis and the blood flow rate in it and the level of testosterone [74]. According to recent reviews and meta-analyses, men with moderate-to-severe LUTS are at increased risk of most cardiovascular complications, and endothelial dysfunction connects them (Fig. 2) [75, 76].

Sex characteristics of pain reception in the LUT

The LUT has extensive innervation, represented by mixed, sensory, and motor autonomic nerve endings (sympathetic and parasympathetic) and motor somatic nerve fibers. Currently, sexual dimorphism in relation to the characteristics of pain reception and perception in men and women is well known, which leads to the conclusion that sex steroid hormones are indirectly involved in their mechanisms. Traditionally, estrogens are believed to be pro-pain hormones, and progesterone and testosterone are anti-pain hormones; therefore, evolutionarily, men are more resistant to acute and chronic pain [77, 78],

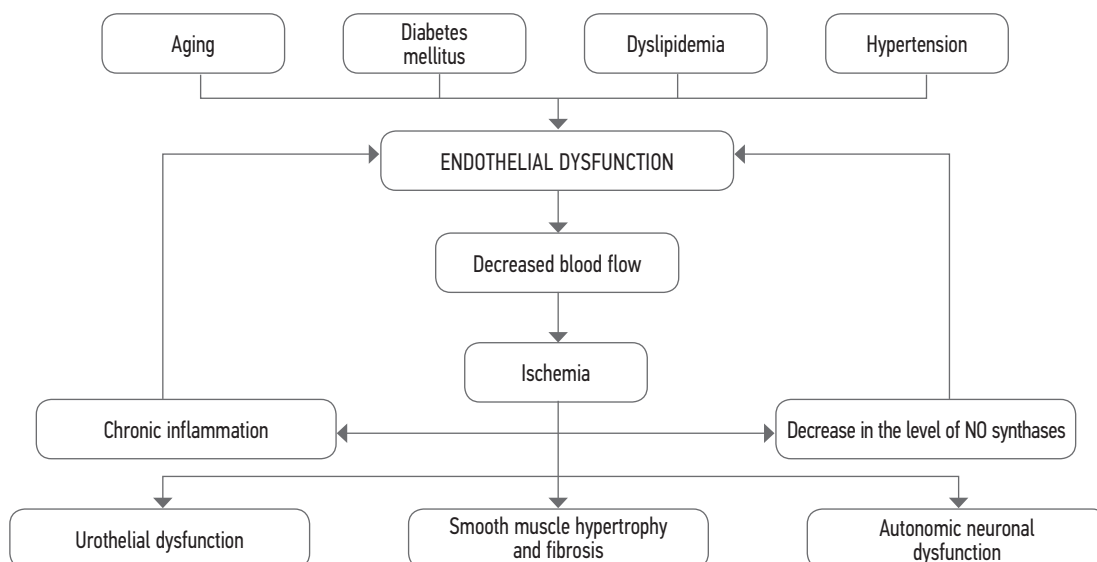


Fig. 2. Scheme of the relationship between endothelial dysfunction and lower urinary tract dysfunction in men [76]

Рис. 2. Схема связи между эндотелиальной дисфункцией и дисфункцией нижних мочевых путей у мужчин [76]

and women often have low pain tolerance [79–81]. A study showed differences between men and women in terms of how they recall the pain experience. Unlike women, men showed increased hypersensitivity to pain under the same painful experiences, and this may be mediated by decreased testosterone levels [82]. Studies analyzing sex differences in people with delayed muscle pain have not reported any significant sex differences [83]. A study of muscle pain induced by saline administration led to an increase in pressure point thresholds in men but not in women [84]. In an endotoxemia model, one of the ways to induce widespread inflammatory pain, a decrease in the initial pain thresholds of pressure in women was detected, but without sex differences after the development of inflammation [85]. The change in pain perception can be caused by various factors, in particular, age. Moreover, no differences in sensitivity to pain were found between older men and women [86], in contrast to the results in young adults [87]. From a mechanistic point of view, variations in pain perception may be associated with differences in brain activation patterns caused by muscle pain, since significant changes in signal intensity in the middle cingulate cortex and dorsolateral prefrontal cortex occur in a sex-dependent dimorphic pattern, which indicates important sex differences in the emotional perception of pain [88]. In diseases or injuries, muscle tissue releases various metabolites, cytokines, and growth factors, which can be accompanied by the infiltration of immune cells. These signals are combined with differential gene expression patterns and receptor interactions in both men and women. In the spinal cord, the increased signals from muscle afferents are possibly modulated by the increased immune reactivity of microglia in men and T cells in women. The perception of pain in the brain may also depend on sex-specific psychological

and emotional factors and may be accompanied by pain sensations in men, unlike those in women [89] (Fig. 3).

Sex differences have been found in healthy people and patients with chronic muscle pain; however, more research is required to elucidate the mechanisms underlying these phenomena.

Characteristics of the hormonal regulation of the LUT in sex and age aspects

Estrogen, progesterone, and testosterone receptors are present in the urinary tract of both sexes. The regulatory mechanisms of the expression of sex hormone receptors differ with age [90–94]. In female and male mice, myocytes and fibroblasts of the lamina propria of the urethral wall have a high density of estrogen receptors such as α (ER- α) and β (ER- β), progesterone receptors (PR), and androgen receptors [95]. In men, the striated muscle cells of the rhabdosphincter showed expression of androgen receptors, estrogen receptors of ER- β , and PR; in women, the expressions of ER- α receptors are predominantly associated with PR [95]. Moreover, experimental knockout of ER- α receptors in the LUT of female mice led to a decrease in several PR-positive cells in the urethra, which suggests that ER- α modulate PR expression in the female urethra; however, the expression disorders of neither ER- α nor ER- β changed the PR expression in the male urethra [95]. Cells expressing ER- β and PR receptors are found in the urethral epithelium, while ER- β^+ cells are detected in the epithelium of the urinary bladder and detrusor of both sexes [96]. In humans, the expression of ER- β receptors was only found in the urothelium of the female urinary bladder [97].

In female rabbits, injections of progesterone or testosterone decreased the capacity and compliance of the

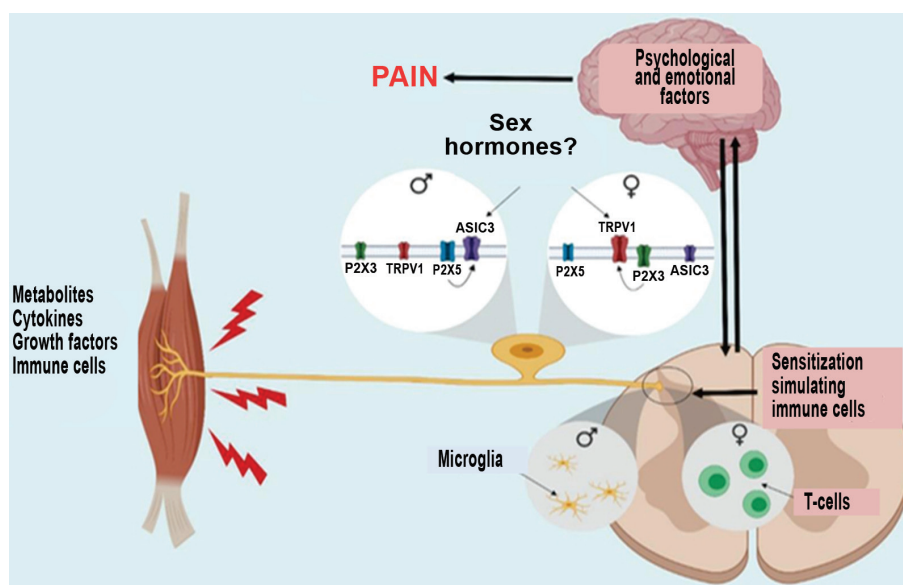


Fig. 3. Gender characteristics of the pathogenesis of muscle pain [89]. TRPV1 – transient receptor potential of vanilloid type 1; P2X3 – type 3 adenosine receptor; P2X5 – type 5 adenosine receptor; ASIC3 – type 3 acid-sensitive ion channel

Рис. 3. Гендерные особенности патогенеза мышечной боли [89]. TRPV1 — транзиторный рецепторный потенциал ваниллоидного типа 1; P2X3 — аденозиновый рецептор типа 3; P2X5 — аденозиновый рецептор типа 5; ASIC3 — кислотно-чувствительный ионный канал типа 3

bladder, whereas estrogen treatment increased them. In male rabbits, testosterone or estrogen injections increase significantly the bladder capacity, but progesterone treatment has no effect [98]. Numerous clinical studies have shown that testosterone therapy in men has significant positive direct and indirect effects on nearly all LUT structures, including the vascular endothelium, neurothelium, muscle structures, and urothelium [99, 100]. The indirect effect of testosterone on the LUT is due to several mechanisms, such as the regulation of neurons of the autonomic nervous system and the activity of cellular enzymes of Rho kinase and phosphodiesterase type 5. In addition, testosterone activates endothelial NOS in the pelvis, which can lead to vasodilation and relief of pelvic ischemia. Bladder blood flow is often reduced in patients with LUTS, and decreased blood flow in the bladder and aging-induced ischemia are associated with the development of anatomical and functional disorders in the detrusor [101–103]. In a laboratory model of testosterone deficiency in male rats, Zhang et al. [104] revealed the rapid development of fibrosis of the bladder wall, which led to a decrease in the volume and contractility of the bladder and an increase in the expression of procollagen I mRNA, which is one of the main markers of age-related fibrosis.

REFERENCES

1. Miller VM. Why are sex and gender important to basic physiology and translational and individualized medicine? *Am J Physiol Heart Circ Physiol.* 2014;306(6): H781–H788. DOI: 10.1152/ajpheart.00994.2013

CONCLUSION

Personalized medicine requires considering the sex of the patient to personalize the examination and potentially improve the results of any therapy. For basic science, the first step toward this aim is the provision of more evidence of the mechanistic and regulatory processes, which are both similar and completely different in men and women. Physiology has invariably made a decisive contribution to understanding the regulatory processes that develop in the body under healthy and disease states. In the future, its contribution will continue to form the basis for the development of personalized medicine. With the inclusion of scientific and practical medical problems to achieve scientific excellence in research and medical education, the disparities in healthcare and practical medicine between women and men will be significantly reduced, and the efficiency of treatment in patients of different sexes will improve, and the quality of its implementation will increase.

ADDITIONAL INFORMATION

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2. Institute of Medicine. Exploring the biological contributions to human health: does sex matter? Washington, DC: The National Academies Press; 2001. Available from: <https://www.nap.edu/read/10028/chapter/1>

3. Collins FS, Tabak LA. Policy: NIH plans to enhance reproducibility. *Nature*. 2014;505(7485):612–613. DOI: 10.1038/505612a
4. Losada L, Amundsen CL, Ashton-Miller J, et al. Expert panel recommendations on lower urinary tract health of women across their life span. *J Women's Health (Larchmt)*. 2016;25(11):1086–1096. DOI: 10.1089/jwh.2016.5895
5. Clayton JA, Collins FS. Policy: NIH to balance sex in cell and animal studies. *Nature*. 2014;509(7500):282–283. DOI: 10.1038/509282a
6. Abelson B, Sun D, Que L, et al. Sex differences in lower urinary tract biology and physiology. *Biol Sex Differences*. 2018;9(1):45–58. DOI: 10.1186/s13293-018-0204-8
7. Parsons CL. The role of the urinary epithelium in the pathogenesis of interstitial cystitis / prostatitis / urethritis. *Urol*. 2007;69(4):9–16. DOI: 10.1016/j.urology.2006.03.084
8. Russo GI, Castelli T, Urzi D, et al. Emerging links between non-neurogenic lower urinary tract symptoms secondary to benign prostatic obstruction, metabolic syndrome and its components: A systematic review. *Int J Urol*. 2015;22(11):982–990. DOI: 10.1111/iju.12877
9. Matthews CA. Risk factors for urinary, fecal, or double incontinence in women. *Curr Opin Obstet Gynecol*. 2014;26(5):393–397. DOI: 10.1097/GCO.0000000000000094
10. Birder LA, de Groat WC. Mechanisms of disease: involvement of the urothelium in bladder dysfunction. *Nat Clin Pract Urol*. 2007;4(1):46–54. DOI: 10.1038/ncpuro0672
11. Tyuzikov IA, Kalinchenko SYu. Endocrinological aspects of chronic cystitis in women. *Experimental and Clinical Urology*. 2016;(3):120–126.
12. Fry CH, Bayliss M, Young JS, Hussain M. Influence of age and bladder dysfunction on the contractile properties of isolated human detrusor smooth muscle. *BJU Int*. 2011;108(2): E91–E96. DOI: 10.1111/j.1464-410X.2010.09845.x
13. Khandelwal P, Abraham SN, Apodaca G. Cell biology and physiology of the uroepithelium. *Am J Physiol Ren Physiol*. 2009;297(6):F1477–1501. DOI: 10.1152/ajprenal.00327.2009
14. Walz T, Haner M, Wu XR, et al. Towards the molecular architecture of the asymmetric unit membrane of the mammalian urinary bladder epithelium: a closed “twisted ribbon” structure. *J Mol Biol*. 1995;248(5):887–900. DOI: 10.1006/jmbi.1995.0269
15. Lin JH, Wu XR, Kreibich G, Sun TT. Precursor sequence, processing, and urothelium-specific expression of a major 15-kDa protein subunit of asymmetric unit membrane. *J Biol Chem*. 1994;269(3):1775–1784.
16. Wu XR, Sun TT. Molecular cloning of a 47 kDa tissue-specific and differentiation dependent urothelial cell surface glycoprotein. *J Cell Sci*. 1993;106(Pt 1):31–43.
17. Yu J, Lin JH, Wu XR, Sun TT. Uroplakins Ia and Ib, two major differentiation products of bladder epithelium, belong to a family of four transmembrane domain (4TM) proteins. *J Cell Biol*. 1994;125:171–182. DOI: 10.1083/jcb.125.1.171
18. Hu P, Meyers S, Liang FX, et al. Role of membrane proteins in permeability barrier function: uroplakin ablation elevates urothelial permeability. *Am J Physiol Ren Physiol*. 2002;283(6): F1200–1207. DOI: 10.1152/ajprenal.00043.2002
19. Aboushwareb T, Zhou G, Deng FM, et al. Alterations in bladder function associated with urothelial defects in uroplakin II and IIIa knockout mice. *NeuroUrol Urodyn*. 2009;28(8):1028–1033. DOI: 10.1002/nau.20688
20. Apodaca G, Balestreire E, Birder LA. The uroepithelial-associated sensory web. *Kidney Int*. 2007;72(9):1057–1064. DOI: 10.1038/sj.ki.5002439
21. Birder LA. Urothelial signaling. *Handb Exp Pharmacol*. 2011;(202):207–231. DOI: 10.1007/978-3-642-16499-6_10
22. Kobayashi H, Yoshiyama M, Zakoji H, Takeda M, Araki I. Sex differences in the expression profile of acid-sensing ion channels in the mouse urinary bladder: a possible involvement in irritative bladder symptoms. *BJU Int*. 2009;104(11):1746–1751. DOI: 10.1111/j.1464-410X.2009.08658.x
23. Page AJ, Brierley SM, Martin CM, et al. Different contributions of ASIC channels 1a, 2, and 3 in gastrointestinal mechanosensory function. *Gut*. 2005;54(10):1408–1415. DOI: 10.1136/gut.2005.071084
24. Luthje P, Brauner H, Ramos NL, et al. Estrogen supports urothelial defense mechanisms. *Sci Transl Med*. 2013;5(190):190ra180. DOI: 10.1126/scitranslmed.3005574
25. Wang C, Symington JW, Ma E, et al. Estrogenic modulation of uropathogenic *Escherichia coli* infection pathogenesis in a murine menopause model. *Infect Immun*. 2013;81(3):733–739. DOI: 10.1128/IAI.01234-12
26. Tincello DG, Taylor AH, Spurling SM, Bell SC. Receptor isoforms that mediate estrogen and progestagen action in the female lower urinary tract. *J Urol*. 2009;181(3):1474–1482. DOI: 10.1016/j.juro.2008.10.104
27. Lu M, Li JR, Alvarez-Lugo L, et al. Lipopolysaccharide stimulates BK channel activity in bladder umbrella cells. *Am J Phys Cell Physiol*. 2018;314(6):643–653. DOI: 10.1152/ajpcell.00339.2017
28. Papavlassopoulos M, Stamme C, Thon L, et al. MaxiK blockade selectively inhibits the lipopolysaccharide-induced I kappa B-alpha / NF-kappa B signaling pathway in macrophages. *J Immunol*. 2006;177(6):4086–4093. DOI: 10.4049/jimmunol.177.6.4086
29. Acevedo-Alvarez M, Yeh J, Alvarez-Lugo L, et al. Mouse urothelial genes associated with voiding behavior changes after ovariectomy and bladder lipopolysaccharide exposure. *NeuroUrol Urodyn*. 2018;37(8):2398–2405. DOI: 10.1002/nau.23592
30. Andersson KE, Arner A. Urinary bladder contraction and relaxation: physiology and pathophysiology. *Physiol Rev*. 2004;84(3):935–986. DOI: 10.1152/physrev.00038.2003
31. DeLancey J, Gosling J, Creed K, et al. Gross anatomy and cell biology of the lower urinary tract. In: Abrams P, Cardozo L, Khoury S, Wein A, editors. *Second international consultation on incontinence*. Plymouth: Health Publication; 2002. P. 17–82.
32. Mangera A, Osman NI, Chapple CR. Anatomy of the lower urinary tract. *Surgery (Oxford)*. 2013;31(7):319–325. DOI: 10.1016/j.mpsur.2010.03.002
33. Lepor H, Machi G. Comparison of AUA symptom index in unselected males and females between fifty-five and seventy-nine years of age. *Urology*. 1993;42:36–40. DOI: 10.1016/0090-4295(93)90332-5
34. Tanaka ST, Ishii K, Demarco RT, et al. Endodermal origin of bladder trigone inferred from mesenchymal-epithelial interaction. *J Urol*. 2010;183:386–391. DOI: 10.1016/j.juro.2009.08.107

35. Favorito LA, Pazos HM, Costa SF, et al. Morphology of the fetal bladder during the second trimester: comparing genders. *J Pediatr Urol*. 2014;10(6):1014–1019. DOI: 10.1016/j.jpuro.2014.11.006
36. Baggish MS, Karram MM. *Atlas of Pelvic Anatomy and Gynecologic Surgery*. Philadelphia: Elsevier / Saunders; 2011.
37. Morita T, Latifpour J, O'Hollaren B, et al. Sex differences in function and distribution of alpha 1- and alpha 2-adrenoceptors in rabbit urethra. *Am J Phys*. 1987;252(6 pt 2): F1124–1128. DOI: 10.1152/ajprenal.1987.252.6.F1124
38. Alexandre EC, de Oliveira MG, Campos R, et al. How important is the alpha1-adrenoceptor in primate and rodent proximal urethra? Sex differences in the contribution of alpha1-adrenoceptor to urethral contractility. *Am J Physiol Ren Physiol*. 2017;312(6): F1026–1034. DOI: 10.1152/ajprenal.00013.2017
39. Oswald J, Heidegger I, Steiner E, et al. Gender-related fetal development of the internal urethral sphincter. *Urology*. 2013;82(6):1410–1415. DOI: 10.1016/j.urology.2013.03.096
40. Jin ZW, Abe H, Hinata N, et al. Descent of mesonephric duct to the final position of the vas deferens in human embryo and fetus. *Anat Cell Biol*. 2016;49(4):231–240. DOI: 10.5115/acb.2016.49.4.231
41. Downing K. Chapter Eight. Biochemistry and ultrastructure of pelvic floor tissues and organs. In: Hoyte L, Damaster M, editors. *Biomechanics of the female pelvic floor*. 1st edition. Elsevier; 2016. P. 181–208. DOI: 10.1016/B978-0-12-803228-2.00008-8
42. Frontera WR, Ochala J. Skeletal muscle: a brief review of structure and function. *Calcif Tissue Int*. 2015;96(3):183–195. DOI: 10.1007/s00223-014-9915-y
43. Dixon J, Gosling J. Structure and innervation in the human. In: *The physiology of the lower urinary tract*. London: Springer; 1987. P. 3–22.
44. Gosling JA, Dixon JS, Critchley HO, Thompson SA. A comparative study of the human external sphincter and peri-urethral levator ani muscles. *Br J Urol*. 1981;53(1):35–41. DOI: 10.1111/j.1464-410x.1981.tb03125.x
45. Praud C, Sebe P, Mondet F, Sebille A. The striated urethral sphincter in female rats. *Anat Embryol (Berl)*. 2003;207(2):169–175. DOI: 10.1007/s00429-003-0340-7
46. Lim SH, Wang TJ, Tseng GF, et al. The distribution of muscles fibers and their types in the female rat urethra: cytoarchitecture and three-dimensional reconstruction. *Anat Rec (Hoboken)*. 2013;296(10):1640–1649. DOI: 10.1002/ar.22740
47. Bierinx AS, Sebille A. The urethral striated sphincter in adult male rat. *Anat Embryol (Berl)*. 2006;211(5):435–441. DOI: 10.1007/s00429-006-0093-1
48. Buffini M, O'Halloran KD, O'Herlihy C, et al. Comparison of the contractile properties, oxidative capacities and fibre type profiles of the voluntary sphincters of continence in the rat. *J Anat*. 2010;217(3):187–195. DOI: 10.1111/j.1469-7580.2010.01263.x
49. Chen SL, Wu M, Henderson JP, et al. Genomic diversity and fitness of *E. coli* strains recovered from the intestinal and urinary tracts of women with recurrent urinary tract infection. *Sci Transl Med*. 2013;5(184):184ra160. DOI: 10.1126/scitranslmed.3005497
50. Benoit G, Quillard J, Jardin A. Anatomical study of the infra-montanal urethra in man. *J Urol*. 1988;139(4):866–868. DOI: 10.1016/s0022-5347(17)42664-4
51. Brading AF. The physiology of the mammalian urinary outflow tract. *Exp Physiol*. 1999;84(1):215–21.
52. Brading AF. The physiology of the mammalian urinary outflow tract. *Exp Physiol*. 1999;84:215–21. DOI: 10.1111/j.1469-445x.1999.tb00084.x
53. Ho KM, McMurray G, Brading AF, et al. Nitric oxide synthase in the heterogeneous population of intramural striated muscle fibres of the human membranous urethral sphincter. *J Urol*. 1998;159: 1091–1096.
54. Tokunaka S, Okamura K, Fujii H, Yachiku S. The proportions of fiber types in human external urethral sphincter: electrophoretic analysis of myosin. *Urol Res*. 1990;18(5):341–344. DOI: 10.1007/BF00300784
55. Bridgewater M, MacNeil HF, Brading AF. Regulation of tone in pig urethral smooth muscle. *J Urol*. 1993;150(1):223–228. DOI: 10.1016/s0022-5347(17)35451-4
56. Persson K, Andersson KE. Non-adrenergic, non-cholinergic relaxation and levels of cyclic nucleotides in rabbit lower urinary tract. *Eur J Pharmacol*. 1994;268(2):159–167. DOI: 10.1016/0922-4106(94)90185-6
57. Persson K, Igawa Y, Mattiasson A, Andersson KE. Effects of inhibition of the L-arginine/nitric oxide pathway in the rat lower urinary tract *in vivo* and *in vitro*. *Br J Pharmacol*. 1992;107(1):178–184. DOI: 10.1111/j.1476-5381.1992.tb14483.x
58. Livingston BP. Anatomy and neural control of the lower urinary tract and pelvic floor. *Top Geriatr Rehabil*. 2016;32(4):280–294. DOI: 10.1097/TGR.0000000000000123
59. Hull T, Zutshi M. Chapter 78. Pathophysiology, diagnosis, and treatment of defecatory dysfunction. In: *Female urology*. 3rd ed. Philadelphia: Elsevier; 2008. P. 761–72. DOI: 10.1016/B978-1-4160-2339-5.50127-0
60. Koelbl H, Strassegger H, Riss PA, Gruber H. Morphologic and functional aspects of pelvic floor muscles in patients with pelvic relaxation and genuine stress incontinence. *Obstet Gynecol*. 1989;74(5):789–795.
61. Jundt K, Kiening M, Fischer P, et al. Is the histomorphological concept of the female pelvic floor and its changes due to age and vaginal delivery correct? *NeuroUrol Urodyn*. 2005;24(1):44–50. DOI: 10.1002/nau.20080
62. Tobin C, Joubert Y. Testosterone-induced development of the rat levator ani muscle. *Dev Biol*. 1991;146(1):131–138. DOI: 10.1016/0012-1606(91)90453-a
63. Niel L, Willemsen KR, Volante SN, Monks DA. Sexual dimorphism and androgen regulation of satellite cell population in differentiating rat levator ani muscle. *Dev Neurobiol*. 2008;68(1):115–122. DOI: 10.1002/dneu.20580
64. Fritsch H, Frohlich B. Development of the levator ani muscle in human fetuses. *Early Hum Dev*. 1994;37(1):15–25. DOI: 10.1016/0378-3782(94)90143-0
65. Anderson TJ, Charbonneau F, Title LM, et al. Microvascular function predicts cardiovascular events in primary prevention: long-term results from the Firefighters and Their Endothelium (FATE) study. *Circulation*. 2011;123(2):163–169. DOI: 10.1161/CIRCULATIONAHA.110.953653

66. Taddei S, Virdis A, Ghiadoni L, et al. Menopause is associated with endothelial dysfunction in women. *Hypertension*. 1996;28(4):576–582. DOI: 10.1161/01.hyp.28.4.576
67. Robinson D, Toozs-Hobson P, Cardozo L. The effect of hormones on the lower urinary tract. *Menopause Int*. 2013;19(4):155–162. DOI: 10.1177/1754045313511398
68. Somani YB, Pawelczyk JA, De Souza MJ, et al. Aging women and their endothelium: Probing the relative role of estrogen on vasodilator function. *Am J Physiol Heart Circ Physiol*. 2019;317(2):H395–H404. DOI: 10.1152/ajpheart.00430.2018
69. Van Geelen JM, Doesburg WH, Thomas CM, Martin CB Jr. Urodynamic studies in the normal menstrual cycle: the relationship between hormonal changes during the menstrual cycle and the urethral pressure profile. *Am J Obstet Gynecol*. 1981;141(4):384–392. DOI: 10.1016/0002-9378(81)90599-8
70. Ansari MA, Begum D, Islam F. Serum sex steroids, gonadotrophins and sex hormonebinding globulin in prostatic hyperplasia. *Ann Saudi Med*. 2008;28(3):174–178. DOI: 10.5144/0256-4947.2008.174
71. Ito S, Juncos LA, Nushiro N., et al. Endothelium-derived relaxing factor modulates endothelin action in afferent arterioles. *Hypertension*. 1991;17(6 Pt. 2):1052–1056. DOI: 10.1161/01.hyp.17.6.1052
72. Tyuzikov IA. Pathogenetic Mechanisms of Influence of Testosterone Deficiency on Lower Urinary Tract Symptoms in Men. *Jeftektivnaja Farmakoterapija*. 2020;16(20):32–42. DOI 10.33978/2307-3586-2020-16-20-32-42
73. Riding DM, Hansrani V, McCollum C. Pelvic vein incompetence: clinical perspectives. *Vasc Health Risk Manag*. 2017;13:439–447. DOI: 10.2147/VHRM.S132827
74. Shigehara K, Namiki M. Late-onset hypogonadism syndrome and lower urinary tract symptoms. *Korean J Urol*. 2011;52(10):657–663. DOI: 10.4111/kju.2011.52.10.657
75. Gacci M, Corona G, Sebastianelli A, et al. Male Lower Urinary Tract Symptoms and Cardiovascular Events: A Systematic Review and Meta-analysis. *Eur Urol*. 2016;70(5):788–796. DOI: 10.1016/j.eururo.2016.07.007
76. Semczuk-Kaczmarek K, Piatek AE, Filip M, Szymański FM. Co-treatment of lower urinary tract symptoms and cardiovascular disease — where do we stand? *Cent European J Urol*. 2020;73(1):42–45. DOI: 10.5173/cej.2020.0029
77. Sorge RE, Totsch SK. Sex Differences in Pain. *J Neurosci Res*. 2017;95(6):1271–1281. DOI: 10.1002/jnr.23841
78. Li J, Baccei ML. Functional Organization of Cutaneous and Muscle Afferent Synapses onto Immature Spinal Lamina I Projection Neurons. *J Neurosci*. 2017;37(6):1505–1517. DOI: 10.1523/JNEUROSCI.3164-16.2016
79. Bueno CH, Pereira DD, Pattussi MP, et al. Gender differences in temporomandibular disorders in adult populational studies: A systematic review and meta-analysis. *J Oral Rehabil*. 2018;45(9):720–729. DOI: 10.1111/joor.12661
80. Mogil JS. Sex differences in pain and pain inhibition: multiple explanations of a controversial phenomenon. *Nat Rev Neurosci*. 2012;13(12):859–866. DOI: 10.1038/nrn3360
81. Rovner GS, Sunnerhagen KS, Bjorkdahl A, et al. Chronic pain and sex-differences; women accept and move, while men feel blue. *PLoS One*. 2017;12(4): e0175737. DOI: 10.1371/journal.pone.0175737
82. Martin LJ, Acland EL, Cho C, et al. Male-Specific Conditioned Pain Hypersensitivity in Mice and Humans. *Current Biol*. 2019;29(2):192–201. DOI: 10.1016/j.cub.2018.11.030
83. Dannecker EA, Liu Y, Rector RS, et al. Sex differences in exercise-induced muscle pain and muscle damage. *J Pain*. 2012;13(12):1242–1249. DOI: 10.1016/j.jpain.2012.09.014
84. Arendt-Nielsen L, Sluka KA, Nie HL. Experimental muscle pain impairs descending inhibition. *Pain*. 2008;140(3):465–471. DOI: 10.1016/j.pain.2008.09.027
85. Wegner A, Elsenbruch S, Rebernik L, et al. Inflammation-induced pain sensitization in men and women: does sex matter in experimental endotoxemia? *Pain*. 2015;156(10):1954–1964. DOI: 10.1097/j.pain.0000000000000256
86. Monroe TB, Fillingim RB, Bruehl SP, et al. Sex Differences in Brain Regions Modulating Pain Among Older Adults: A Cross-Sectional Resting State Functional Connectivity Study. *Pain Medicine*. 2018;19(9):1737–1747. DOI: 10.1093/pm/pnx084
87. Gupta A, Mayer EA, Fling C, et al. Sex-based differences in brain alterations across chronic pain conditions. *Journal of Neuroscience Research*. 2017;95(1–2):604–616. DOI: 10.1002/jnr.23856
88. Henderson LA, Gandevia SC, Macefield VG. Gender differences in brain activity evoked by muscle and cutaneous pain: a retrospective study of single-trial fMRI data. *Neuroimage*. 2008;39(4):1867–1876. DOI: 10.1016/j.neuroimage.2007.10.045
89. Queme LF, Jankowski MP. Sex differences and mechanisms of muscle pain. *Curr Opin Physiol*. 2019;11:1–6. DOI: 10.1016/j.cophys.2019.03.006
90. Blakeman PJ, Hilton P, Bulmer JN. Oestrogen and progesterone receptor expression in the female lower urinary tract, with reference to oestrogen status. *BJU Int*. 2000;86(1):32–38. DOI: 10.1046/j.1464-410x.2000.00724.x
91. Celayir S. Is there a “bladder sex”? The relation of different sex hormones and sex hormone receptors in bladder in childhood. *Med Hypotheses*. 2002;59(2):186–190. DOI: 10.1016/s0306-9877(02)00245-1
92. Keast JR, Saunders RJ. Testosterone has potent, selective effects on the morphology of pelvic autonomic neurons which control the bladder, lower bowel and internal reproductive organs of the male rat. *Neuroscience*. 1998;85(2):543–556. DOI: 10.1016/s0306-4522(97)00631-3
93. Makela S, Strauss L, Kuiper G, et al. Differential expression of estrogen receptors alpha and beta in adult rat accessory sex glands and lower urinary tract. *Mol Cell Endocrinol*. 2000;170(1–2):219–229. DOI: 10.1016/s0303-7207(00)00441-x
94. McKenna KE, Nadelhaft I. The organization of the pudendal nerve in the male and female rat. *J Comp Neurol*. 1986;248(4):532–549. DOI: 10.1002/cne.902480406
95. Savolainen S, Santti R, Streng T, et al. Sex specific expression of progesterone receptor in mouse lower urinary tract. *Mol Cell Endocrinol*. 2005;230(1–2):17–21. DOI: 10.1016/j.mce.2004.11.008

- 96.** Tincello DG, Taylor AH, Spurling SM, Bell SC. Receptor isoforms that mediate estrogen and progestagen action in the female lower urinary tract. *J Urol*. 2009;181(3):1474–1482. DOI: 10.1016/j.juro.2008.10.104
- 97.** Bødker A, Balslev E, Juul BR, et al. Estrogen receptors in the human male bladder, prostatic urethra, and prostate: an immunohistochemical and biochemical study. *Scand J Urol Nephrol*. 1995;29(2):161–165. DOI: 10.3109/00365599509180557
- 98.** Celayir S. Effects of different sex hormones on male rabbit urodynamics: an experimental study. *Horm Res*. 2003;60(5):215–220. DOI: 10.1159/000074034
- 99.** Rohrmann S, Nelson WG, Rifai N. Serum sex steroid hormones and lower urinary tract symptoms in Third National Health and Nutrition Examination Survey (NHANES III) *Urol*. 2007;69(4):708–713. DOI: 10.1016/j.urology.2007.01.011
- 100.** Navarro-Dorado J, Orensanz LM, Recio P. Mechanisms involved in testosterone-induced vasodilatation in pig prostatic small arteries. *Life Sci*. 2008;83(15–16):569–573. DOI: 10.1016/j.lfs.2008.08.009
- 101.** Mitterberger M, Pallwein L, Gradl J, et al. Persistent detrusor overactivity after transurethral resection of the prostate is associated with reduced perfusion of the urinary bladder. *BJU Int*. 2007;99(4):831–835. DOI: 10.1111/j.1464-410X.2006.06735.x
- 102.** McVary KT. Erectile dysfunction and lower urinary tract symptoms secondary to BPH. *Eur Urol*. 2005;47(6):838–845. DOI: 10.1016/j.eururo.2005.02.001
- 103.** Azadzoi KM, Tarcan T, Kozłowski R, et al. Overactivity and structural changes in the chronically ischemic bladder. *J Urol*. 1999;162(5):1768–1778.
- 104.** Zhang Y, Chen J, Hu L, Chen Z. Androgen deprivation induces bladder histological abnormalities and dysfunction via TGF- β in orchiectomized mature rats. *Tohoku J Exp Med*. 2012;226(2):121–128. DOI: 10.1620/tjem.226.121

СПИСОК ЛИТЕРАТУРЫ

- 1.** Miller V.M. Why are sex and gender important to basic physiology and translational and individualized medicine? // *Am J Physiol Heart Circ Physiol*. 2014. Vol. 306, No. 6. P. H781–H788. DOI: 10.1152/ajpheart.00994.2013
- 2.** Institute of Medicine. Exploring the biological contributions to human health: does sex matter? Washington, DC: The National Academies Press; 2001. Режим доступа: <https://www.nap.edu/read/10028/chapter/1>. Дата обращения: 09.08.2021.
- 3.** Collins F.S., Tabak L.A. Policy: NIH plans to enhance reproducibility // *Nature*. 2014. Vol. 505, No. 7485. P. 612–613. DOI: 10.1038/505612a
- 4.** Losada L., Amundsen C.L., Ashton-Miller J., et al. Expert panel recommendations on lower urinary tract health of women across their life span // *J Women's Health (Larchmt)*. 2016. Vol. 25, No. 11. P. 1086–1096; DOI: 10.1089/jwh.2016.5895
- 5.** Clayton J.A., Collins F.S. Policy: NIH to balance sex in cell and animal studies // *Nature*. 2014. Vol. 509, No. 7500. P. 282–283. DOI: 10.1038/509282a
- 6.** Abelson B., Sun D., Que L., et al. Sex differences in lower urinary tract biology and physiology // *Biol Sex Differences*. 2018. Vol. 9, No. 1. P. 45–58. DOI: 10.1186/s13293-018-0204-8
- 7.** Parsons C.L. The role of the urinary epithelium in the pathogenesis of interstitial cystitis / prostatitis / urethritis // *Urol*. 2007. Vol. 69, Suppl. 4. P. 9–16. DOI: 10.1016/j.urology.2006.03.084
- 8.** Russo G.I., Castelli T., Urzi D., et al. Emerging links between non-neurogenic lower urinary tract symptoms secondary to benign prostatic obstruction, metabolic syndrome and its components: A systematic review // *Int J Urol*. 2015. Vol. 22, No. 11. P. 982–990. DOI: 10.1111/iju.12877
- 9.** Matthews C.A. Risk factors for urinary, fecal, or double incontinence in women // *Curr Opin Obstet Gynecol*. 2014. Vol. 26, No. 5. P. 393–397. DOI: 10.1097/GCO.0000000000000094
- 10.** Birder L.A., de Groat W.C. Mechanisms of disease: involvement of the urothelium in bladder dysfunction // *Nat Clin Pract Urol*. 2007. Vol. 4, No. 1. P. 46–54. DOI: 10.1038/ncpuro0672
- 11.** Тюзиков И.А. Эндокринологические аспекты хронического цистита у женщин. Часть 1. Общие эндокринологические аспекты // *Экспериментальная и клиническая урология*. 2016. № 3. С. 120–126.
- 12.** Fry C.H., Bayliss M., Young J.S., Hussain M. Influence of age and bladder dysfunction on the contractile properties of isolated human detrusor smooth muscle // *BJU Int*. 2011. Vol. 108, No. 2. P. E91–E96. DOI: 10.1111/j.1464-410X.2010.09845.x
- 13.** Khandelwal P., Abraham S.N., Apodaca G. Cell biology and physiology of the uroepithelium // *Am J Physiol Ren Physiol*. 2009. Vol. 297, No. 6. P. F1477–1501. DOI: 10.1152/ajprenal.00327.2009
- 14.** Walz T., Haner M., Wu X.R., et al. Towards the molecular architecture of the asymmetric unit membrane of the mammalian urinary bladder epithelium: a closed «twisted ribbon» structure // *J Mol Biol*. 1995. Vol. 248, No. 5. P. 887–900. DOI: 10.1006/jmbi.1995.0269
- 15.** Lin J.H., Wu X.R., Kreibich G., Sun T.T. Precursor sequence, processing, and urothelium-specific expression of a major 15-kDa protein subunit of asymmetric unit membrane // *J Biol Chem*. 1994. Vol. 269, No. 3. P. 1775–1784.
- 16.** Wu X.R., Sun T.T. Molecular cloning of a 47 kDa tissue-specific and differentiation dependent urothelial cell surface glycoprotein // *J Cell Sci*. 1993. Vol. 106, No. 1. P. 31–43.
- 17.** Yu J., Lin J.H., Wu X.R., Sun T.T. Uroplakins Ia and Ib, two major differentiation products of bladder epithelium, belong to a family of four transmembrane domain (4TM) proteins // *J Cell Biol*. 1994. Vol. 125. P. 171–182. DOI: 10.1083/jcb.125.1.171
- 18.** Hu P., Meyers S., Liang F.X., et al. Role of membrane proteins in permeability barrier function: uroplakin ablation elevates urothelial

- permeability // *Am J Physiol Ren Physiol*. 2002. Vol. 283, No. 6. P. F1200–1207. DOI: 10.1152/ajprenal.00043.2002
19. Aboushwareb T., Zhou G., Deng FM, et al. Alterations in bladder function associated with urothelial defects in uroplakin II and IIIa knockout mice // *Neurourol Urodyn*. 2009. Vol. 28, No. 8. P. 1028–1033. DOI: 10.1002/nau.20688
20. Apodaca G., Balestreire E., Birder L.A. The uroepithelial-associated sensory web // *Kidney Int*. 2007. Vol. 72, No. 9. P. 1057–1064. DOI: 10.1038/sj.ki.5002439
21. Birder L.A. Urothelial signaling // *Handb Exp Pharmacol*. 2011. No. 202. P. 207–231. DOI: 10.1007/978-3-642-16499-6_10
22. Kobayashi H., Yoshiyama M., Zakoji H., et al. Sex differences in the expression profile of acid-sensing ion channels in the mouse urinary bladder: a possible involvement in irritative bladder symptoms // *BJU Int*. 2009. Vol. 104, No. 11. P. 1746–1751. DOI: 10.1111/j.1464-410X.2009.08658.x
23. Page A.J., Brierley S.M., Martin C.M., et al. Different contributions of ASIC channels 1a, 2, and 3 in gastrointestinal mechanosensory function // *Gut*. 2005. Vol. 54, No. 10. P. 1408–1415. DOI: 10.1136/gut.2005.071084
24. Luthje P., Brauner H., Ramos N.L., et al. Estrogen supports urothelial defense mechanisms // *Sci Transl Med*. 2013. Vol. 5, No. 190. P. 190ra180. DOI: 10.1126/scitranslmed.3005574
25. Wang C., Symington J.W., Ma E., et al. Estrogenic modulation of uropathogenic *Escherichia coli* infection pathogenesis in a murine menopause model // *Infect Immun*. 2013. Vol. 81, No. 3. P. 733–739. DOI: 10.1128/IAI.01234-12
26. Tincello D.G., Taylor A.H., Spurling S.M., Bell S.C. Receptor isoforms that mediate estrogen and progesterone action in the female lower urinary tract // *J Urol*. 2009. Vol. 181, No. 3. P. 1474–1482. DOI: 10.1016/j.juro.2008.10.104
27. Lu M., Li J.R., Alvarez-Lugo L., et al. Lipopolysaccharide stimulates BK channel activity in bladder umbrella cells // *Am J Phys Cell Physiol*. 2018. Vol. 314, No. 6. P. 643–653. DOI: 10.1152/ajpcell.00339.2017
28. Papavlassopoulos M., Stamme C., Thon L., et al. MaxiK blockade selectively inhibits the lipopolysaccharide-induced I kappa B-alpha / NF-kappa B signaling pathway in macrophages // *J Immunol*. 2006. Vol. 177, No. 6. P. 4086–4093. DOI: 10.4049/jimmunol.177.6.4086
29. Acevedo-Alvarez M., Yeh J., Alvarez-Lugo L., et al. Mouse urothelial genes associated with voiding behavior changes after ovariectomy and bladder lipopolysaccharide exposure // *Neurourol Urodyn*. 2018. Vol. 37, No. 8. P. 2398–2405. DOI: 10.1002/nau.23592
30. Andersson K.E., Arner A. Urinary bladder contraction and relaxation: physiology and pathophysiology // *Physiol Rev*. 2004. Vol. 84, No. 3. P. 935–986. DOI: 10.1152/physrev.00038.2003
31. DeLancey J., Gosling J., Creed K., et al. Gross anatomy and cell biology of the lower urinary tract. In: Abrams P., Cardozo L., Khoury S., Wein A., editors. Second international consultation on incontinence. Plymouth: Health Publication; 2002. P. 17–82.
32. Mangera A., Osman N.I., Chapple C.R. Anatomy of the lower urinary tract // *Surgery (Oxford)*. 2013. Vol. 31, No. 7. P. 319–325. DOI: 10.1016/j.mpsur.2010.03.002
33. Lepor H., Machi G. Comparison of AUA symptom index in unselected males and females between fifty-five and seventy-nine years of age // *Urology*. 1993. Vol. 42. P. 36–40. DOI: 10.1016/0090-4295(93)90332-5
34. Tanaka S.T., Ishii K., Demarco R.T., et al. Endodermal origin of bladder trigone inferred from mesenchymal-epithelial interaction // *J Urol*. 2010. Vol. 183, No. 1. P. 386–391. DOI: 10.1016/j.juro.2009.08.107
35. Favorito L.A., Pazos H.M., Costa S.F., et al. Morphology of the fetal bladder during the second trimester: comparing genders // *J Pediatr Urol*. 2014. Vol. 10, No. 6. P. 1014–1019. DOI: 10.1016/j.jpuro.2014.11.006
36. Baggish M.S., Karram M.M. Atlas of Pelvic Anatomy and Gynecologic Surgery. Philadelphia: Elsevier/Saunders, 2011.
37. Morita T., Latifpour J., O'Hollaren B., et al. Sex differences in function and distribution of alpha 1- and alpha 2-adrenoceptors in rabbit urethra // *Am J Phys*. 1987. Vol. 252, No. 6, pt 2. P. F1124–1128. DOI: 10.1152/ajprenal.1987.252.6.F1124
38. Alexandre E.C., de Oliveira M.G., Campos R., et al. How important is the alpha1-adrenoceptor in primate and rodent proximal urethra? Sex differences in the contribution of alpha1-adrenoceptor to urethral contractility // *Am J Physiol Ren Physiol*. 2017. Vol. 312, No. 6. P. F1026–1034. DOI: 10.1152/ajprenal.00013.2017
39. Oswald J., Heidegger I., Steiner E., et al. Gender-related fetal development of the internal urethral sphincter // *Urology*. 2013. Vol. 82, No. 6. P. 1410–1415. DOI: 10.1016/j.urology.2013.03.096
40. Jin Z.W., Abe H., Hinata N., et al. Descent of mesonephric duct to the final position of the vas deferens in human embryo and fetus // *Anat Cell Biol*. 2016. Vol. 49, No. 4. P. 231–240. DOI: 10.5115/acb.2016.49.4.231
41. Downing K. Chapter Eight; Biochemistry and ultrastructure of pelvic floor tissues and organs. In: Biomechanics of the female pelvic floor. 1st edition. Hoyte L., Damaster M., eds. Elsevier, 2016. P. 181–208. DOI: 10.1016/B978-0-12-803228-2.00008-8
42. Frontera W.R., Ochala J. Skeletal muscle: a brief review of structure and function // *Calcif Tissue Int*. 2015. Vol. 96. P. 183–195. DOI: 10.1007/s00223-014-9915-y
43. Dixon J., Gosling J. Structure and innervation in the human. In: The physiology of the lower urinary tract. London: Springer, 1987. P. 3–22.
44. Gosling J.A., Dixon J.S., Critchley H.O., Thompson S.A. A comparative study of the human external sphincter and periurethral levator ani muscles // *Br J Urol*. 1981. Vol. 53, No. 1. P. 35–41. DOI: 10.1111/j.1464-410x.1981.tb03125.x
45. Praud C., Sebe P., Mondet F., Sebille A. The striated urethral sphincter in female rats // *Anat Embryol (Berl)*. 2003. Vol. 207, No. 2. P. 169–175. DOI: 10.1007/s00429-003-0340-7
46. Lim S.H., Wang T.J., Tseng G.F., et al. The distribution of muscle fibers and their types in the female rat urethra: cytoarchitecture and three-dimensional reconstruction // *Anat Rec (Hoboken)*. 2013. Vol. 296, No. 10. P. 1640–1649. DOI: 10.1002/ar.22740
47. Bierinx A.S., Sebille A. The urethral striated sphincter in adult male rat // *Anat Embryol (Berl)*. 2006. Vol. 211, No. 5. P. 435–441. DOI: 10.1007/s00429-006-0093-1

- 48.** Buffini M., O'Halloran K.D., O'Herlihy C., et al. Comparison of the contractile properties, oxidative capacities and fibre type profiles of the voluntary sphincters of continence in the rat // *J Anat.* 2010. Vol. 217, No. 3. P. 187–195. DOI: 10.1111/j.1469-7580.2010.01263.x
- 49.** Chen S.L., Wu M., Henderson J.P., et al. Genomic diversity and fitness of *E. coli* strains recovered from the intestinal and urinary tracts of women with recurrent urinary tract infection // *Sci Transl Med.* 2013. Vol. 5. No. 184. P. 184ra160. DOI: 10.1126/scitranslmed.3005497
- 50.** Benoit G., Quillard J., Jardin A. Anatomical study of the inframontanal urethra in man // *J Urol.* 1988. Vol. 139, No. 4. P. 866–868. DOI: 10.1016/s0022-5347(17)42664-4
- 51.** Brading AF. The physiology of the mammalian urinary outflow tract // *Exp Physiol.* 1999. Vol. 84. P. 215–221.
- 52.** Ho K.M., McMurray G., Brading A., Noble J. The human female striated urethral sphincter consists of a heterogeneous population of muscle fibres // *Br J Urol.* 1997. Vol. 79. P. 13.
- 53.** Ho K.M., McMurray G., Brading A.F., et al. Nitric oxide synthase in the heterogeneous population of intramural striated muscle fibres of the human membranous urethral sphincter // *J Urol.* 1998. Vol. 159. P. 1091–1096.
- 54.** Tokunaka S., Okamura K., Fujii H., Yachiku S. The proportions of fiber types in human external urethral sphincter: electrophoretic analysis of myosin // *Urol Res.* 1990. Vol. 18, No. 5. P. 341–344. DOI: 10.1007/BF00300784
- 55.** Bridgewater M., MacNeil H.F., Brading A.F. Regulation of tone in pig urethral smooth muscle // *J Urol.* 1993. Vol. 150, No. 1. P. 223–228. DOI: 10.1016/s0022-5347(17)35451-4
- 56.** Persson K., Andersson K.E. Non-adrenergic, non-cholinergic relaxation and levels of cyclic nucleotides in rabbit lower urinary tract // *Eur J Pharmacol.* 1994. Vol. 268, No. 2. P. 159–167. DOI: 10.1016/0922-4106(94)90185-6
- 57.** Persson K., Igawa Y., Mattiasson A., Andersson K.E. Effects of inhibition of the L-arginine / nitric oxide pathway in the rat lower urinary tract *in vivo* and *in vitro* // *Br J Pharmacol.* 1992. Vol. 107, No. 1. P. 178–184. DOI: 10.1111/j.1476-5381.1992.tb14483.x
- 58.** Livingston B.P. Anatomy and neural control of the lower urinary tract and pelvic floor // *Top Geriatr Rehabil.* 2016. Vol. 32, No. 4. P. 280–294. DOI:10.1097/TGR.000000000000123
- 59.** Hull T., Zutshi M. Chapter 78. Pathophysiology, diagnosis, and treatment of defecatory dysfunction. In: *Female urology*. 3rd ed. Philadelphia: Elsevier, 2008. P. 761–72. DOI: 10.1016/B978-1-4160-2339-5.50127-0
- 60.** Koelbl H., Strassegger H., Riss P.A., Gruber H. Morphologic and functional aspects of pelvic floor muscles in patients with pelvic relaxation and genuine stress incontinence // *Obstet Gynecol.* 1989. Vol. 74. P. 789–795.
- 61.** Jundt K., Kiening M., Fischer P., et al. Is the histomorphological concept of the female pelvic floor and its changes due to age and vaginal delivery correct? // *Neurourol Urodyn.* 2005. Vol. 24, No. 1. P. 44–50. DOI: 10.1002/nau.20080
- 62.** Tobin C., Joubert Y. Testosterone-induced development of the rat levatorani muscle // *Dev Biol.* 1991. Vol. 146, No. 1. P. 131–138. DOI: 10.1016/0012-1606(91)90453-a
- 63.** Niel L., Willemsen K.R., Volante S.N., Monks D.A. Sexual dimorphism and androgen regulation of satellite cell population in differentiating rat levator ani muscle // *Dev Neurobiol.* 2008. Vol. 68, No. 1. P. 115–122. DOI: 10.1002/dneu.20580
- 64.** Fritsch H., Frohlich B. Development of the levator ani muscle in human fetuses // *Early Hum Dev.* 1994. Vol. 37, No. 1. P. 15–25. DOI: 10.1016/0378-3782(94)90143-0
- 65.** Anderson T.J., Charbonneau F., Title L.M., et al. Microvascular function predicts cardiovascular events in primary prevention: long-term results from the Firefighters and Their Endothelium (FATE) study // *Circulation.* 2011. Vol. 123, No. 2. P. 163–169. DOI: 10.1161/CIRCULATIONAHA.110.953653
- 66.** Taddei S., Virdis A., Ghiadoni L., et al. Menopause is associated with endothelial dysfunction in women // *Hypertension.* 1996. Vol. 28, No. 4. P. 576–582. DOI: 10.1161/01.hyp.28.4.576
- 67.** Robinson D., Tooze-Hobson P., Cardozo L. The effect of hormones on the lower urinary tract // *Menopause Int.* 2013. Vol. 19, No. 4. P. 155–162. DOI: 10.1177/1754045313511398
- 68.** Somani Y.B., Pawelczyk J.A., De Souza M.J., et al. Aging women and their endothelium: Probing the relative role of estrogen on vasodilator function // *Am J Physiol Heart Circ Physiol.* 2019. Vol. 317, No. 2. P. H395–H404. DOI: 10.1152/ajpheart.00430.2018
- 69.** Van Geelen J.M., Doesburg W.H., Thomas C.M., Martin C.B. Urodynamic studies in the normal menstrual cycle: the relationship between hormonal changes during the menstrual cycle and the urethral pressure profile // *Am J Obstet Gynecol.* 1981. Vol. 141, No. 4. P. 384–392. DOI: 10.1016/0002-9378(81)90599-8
- 70.** Ansari M.A., Begum D., Islam F. Serum sex steroids, gonadotrophins and sex hormonebinding globulin in prostatic hyperplasia // *Ann Saudi Med.* 2008. Vol. 28, No. 3. P. 174–178. DOI: 10.5144/0256-4947.2008.174
- 71.** Ito S., Juncos L.A., Nushiro N., et al. Endothelium-derived relaxing factor modulates endothelin action in afferent arterioles // *Hypertension.* 1991. Vol. 17, 6 Pt. 2. P. 1052–1056. DOI: 10.1161/01.hyp.17.6.1052
- 72.** Тюзиков И.А. Патогенетические механизмы влияния дефицита тестостерона на симптомы нижних мочевых путей у мужчин // Эффективная фармакотерапия. 2020. Т. 16, № 20. С. 32–42. DOI 10.33978/2307-3586-2020-16-20-32-42
- 73.** Riding D.M., Hansrani V., McCollum C. Pelvic vein incompetence: clinical perspectives. // *Vasc Health Risk Manag.* 2017. Vol. 13. P. 439–447. DOI: 10.2147/VHRM.S132827
- 74.** Shigehara K., Namiki M. Late-onset hypogonadism syndrome and lower urinary tract symptoms // *Korean J Urol.* 2011. Vol. 52, No. 10. P. 657–663. DOI: 10.4111/kju.2011.52.10.657
- 75.** Gacci M., Corona G., Sebastianelli A., et al. Male Lower Urinary Tract Symptoms and Cardiovascular Events: A Systematic Review and Meta-analysis // *Eur Urol.* 2016. Vol. 70, No. 5. P. 788–796. DOI: 10.1016/j.eururo.2016.07.007
- 76.** Semczuk-Kaczmarek K., Płatek A.E., Filip M. Szymański FM. Co-treatment of lower urinary tract symptoms and cardiovascular disease — where do we stand? // *Cent European J Urol.* 2020. Vol. 73, No. 1. P. 42–45. DOI: 10.5173/ceju.2020.0029

- 77.** Sorge R.E., Totsch S.K. Sex Differences in Pain // *J Neurosci Res.* 2017. Vol. 95, No. 6. P. 1271–1281. DOI: 10.1002/jnr.23841
- 78.** Li J., Baccei M.L. Functional Organization of Cutaneous and Muscle Afferent Synapses onto Immature Spinal Lamina I Projection Neurons // *J Neurosci.* 2017. Vol. 37, No. 6. P. 1505–1517. DOI: 10.1523/JNEUROSCI.3164-16.2016
- 79.** Bueno C.H., Pereira D.D., Pattussi M.P., et al. Gender differences in temporomandibular disorders in adult populational studies: A systematic review and meta-analysis // *J Oral Rehabil.* 2018. Vol. 45, No. 9. P. 720–729. DOI: 10.1111/joor.12661
- 80.** Mogil J.S. Sex differences in pain and pain inhibition: multiple explanations of a controversial phenomenon // *Nat Rev Neurosci.* 2012. Vol. 13, No. 12. P. 859–866. DOI: 10.1038/nrn3360
- 81.** Rovner G.S., Sunnerhagen K.S., Bjorkdahl A., et al. Chronic pain and sex-differences; women accept and move, while men feel blue // *PLoS One.* 2017. Vol. 12, No. (4). P. e0175737. DOI: 10.1371/journal.pone.0175737
- 82.** Martin L.J., Acland E.L., Cho C., et al. Male-Specific Conditioned Pain Hypersensitivity in Mice and Humans // *Current Biol.* 2019. Vol. 29, No. 2. P. 192–201. DOI: 10.1016/j.cub.2018.11.030
- 83.** Dannecker E.A., Liu Y., Rector R.S., et al. Sex differences in exercise-induced muscle pain and muscle damage // *J Pain.* 2012. Vol. 13, No. 12. P. 1242–1249. DOI: 10.1016/j.jpain.2012.09.014
- 84.** Arendt-Nielsen L., Sluka K.A., Nie H.L. Experimental muscle pain impairs descending inhibition // *Pain.* 2008. Vol. 140, No. 3. P. 465–471. DOI: 10.1016/j.pain.2008.09.027
- 85.** Wegner A., Elsenbruch S., Rebernik L., et al. Inflammation-induced pain sensitization in men and women: does sex matter in experimental endotoxemia? // *Pain.* 2015. Vol. 156, No. 10. P. 1954–1964. DOI: 10.1097/j.pain.0000000000000256
- 86.** Monroe T.B., Fillingim R.B., Bruehl S.P., et al. Sex Differences in Brain Regions Modulating Pain Among Older Adults: A Cross-Sectional Resting State Functional Connectivity Study // *Pain Medicine.* 2018. Vol. 19, No. 9. P. 1737–1747. DOI: 10.1093/pm/pnx084
- 87.** Gupta A., Mayer E.A., Fling C., et al. Sex-based differences in brain alterations across chronic pain conditions // *Journal of Neuroscience Research.* 2017. Vol. 95, No. 1–2. P. 604–616. DOI: 10.1002/jnr.23856
- 88.** Henderson L.A., Gandevia S.C., Macefield V.G. Gender differences in brain activity evoked by muscle and cutaneous pain: a retrospective study of single-trial fMRI data // *Neuroimage.* 2008. Vol. 39, No. 4. P. 1867–1876. DOI: 10.1016/j.neuroimage.2007.10.045
- 89.** Queme L.F., Jankowski M.P. // *Curr Opin Physiol.* 2019. Vol. 11. P. 1–6. DOI: 10.1016/j.cophys.2019.03.006
- 90.** Blakeman P.J., Hilton P., Bulmer J.N. Oestrogen and progesterone receptor expression in the female lower urinary tract, with reference to oestrogen status // *BJU Int.* 2000. Vol. 86, No. 1. P. 32–38. DOI: 10.1046/j.1464-410x.2000.00724.x
- 91.** Celayir S. Is there a “bladder sex”? The relation of different sex hormones and sex hormone receptors in bladder in childhood // *Med Hypotheses.* 2002. Vol. 59, No. 2. P. 186–190. DOI: 10.1016/s0306-9877(02)00245-1
- 92.** Keast J.R., Saunders R.J. Testosterone has potent, selective effects on the morphology of pelvic autonomic neurons which control the bladder, lower bowel and internal reproductive organs of the male rat // *Neuroscience.* 1998. Vol. 85, No. 2. P. 543–556. DOI: 10.1016/s0306-4522(97)00631-3
- 93.** Makela S., Strauss L., Kuiper G., et al. Differential expression of estrogen receptors alpha and beta in adult rat accessory sex glands and lower urinary tract // *Mol Cell Endocrinol.* 2000. Vol. 170, No. 1–2. P. 219–229. DOI: 10.1016/s0303-7207(00)00441-x
- 94.** McKenna K.E., Nadelhaft I. The organization of the pudendal nerve in the male and female rat // *J Comp Neurol.* 1986. Vol. 248, No. 4. P. 532–549. DOI: 10.1002/cne.902480406
- 95.** Savolainen S., Santti R., Streng T., et al. Sex specific expression of progesterone receptor in mouse lower urinary tract // *Mol Cell Endocrinol.* 2005. Vol. 230, No. 1–2. P. 17–21. DOI: 10.1016/j.mce.2004.11.008
- 96.** Tincello D.G., Taylor A.H., Spurling S.M., Bell S.C. Receptor isoforms that mediate estrogen and progestagen action in the female lower urinary tract // *J Urol.* 2009. Vol. 181, No. 3. P. 1474–1482. DOI: 10.1016/j.juro.2008.10.104
- 97.** Bødker A., Balslev E., Juul B.R., et al. Estrogen receptors in the human male bladder, prostatic urethra, and prostate: an immunohistochemical and biochemical study // *Scand J Urol Nephrol.* 1995. Vol. 29, No. 2. P. 161–165. DOI: 10.3109/00365599509180557
- 98.** Celayir S. Effects of different sex hormones on male rabbit urodynamics: an experimental study // *Horm Res.* 2003. Vol. 60, No. 5. P. 215–220. DOI: 10.1159/000074034
- 99.** Rohrmann S., Nelson W.G., Rifai N. Serum sex steroid hormones and lower urinary tract symptoms in Third National Health and Nutrition Examination Survey (NHANES III) // *Urol.* 2007. Vol. 69, No. 4. P. 708–713. DOI: 10.1016/j.urology.2007.01.011
- 100.** Navarro-Dorado J., Orensanz L.M., Recio P. Mechanisms involved in testosterone-induced vasodilatation in pig prostatic small arteries // *Life Sci.* 2008. Vol. 83, No. 15–16. P. 569–573. DOI: 10.1016/j.lfs.2008.08.009
- 101.** Mitterberger M., Pallwein L., Gradl J., et al. Persistent detrusor overactivity after transurethral resection of the prostate is associated with reduced perfusion of the urinary bladder // *BJU Int.* 2007. Vol. 99, No. 4. P. 831–835. DOI: 10.1111/j.1464-410X.2006.06735.x
- 102.** McVary K.T. Erectile dysfunction and lower urinary tract symptoms secondary to BPH // *Eur Urol.* 2005. Vol. 47, No. 6. P. 838–845. DOI: 10.1016/j.eururo.2005.02.001
- 103.** Azadzoi K.M., Tarcan T., Kozłowski R., et al. Overactivity and structural changes in the chronically ischemic bladder // *J Urol.* 1999. Vol. 162, No. 5. P. 1768–1778.
- 104.** Zhang Y., Chen J., Hu L., Chen Z. Androgen deprivation induces bladder histological abnormalities and dysfunction via TGF- β in orchietomized mature rats // *Tohoku J Exp. Med.* 2012. Vol. 226, No. 2. P. 121–128. DOI: 10.1620/tjem.226.121

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